Global Theme on Agroecosystems Report no. 8

Vermicomposting: Recycling Wastes into Valuable Organic Fertilizer



Citation: Nagavallemma KP, Wani SP, Stephane Lacroix, Padmaja VV, Vineela C, Babu Rao M and Sahrawat KL. 2004. Vermicomposting: Recycling wastes into valuable organic fertilizer. Global Theme on Agrecosystems Report no. 8. Patancheru 502 324, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics. 20 pp.

Abstract

The large quantity of organic waste, nearly 700 million t yr⁻¹, generated in India is either burned or land filled posing a problem of safe disposal. To mitigate this problem all the waste can be converted into highly valuable nutrient-rich compost in an environment friendly manner. Vermicomposting is one of the best methods of composting any kind of organic matter, which could provide a 'win-win' solution to tackle the problem of safe disposal of waste and also provide most needed plant nutrients for sustainable productivity.

Vermicompost improves growth, quality and yield of different field crops, flower and fruit crops. Vermicomposting contributes to recycling of nitrogen and augments soil physico-chemical as well as biological properties. Microbial biodiversity was checked and higher diversity was recorded in the partially decomposed organic material for the vermicompost than in the vermicompost. All kinds of organic material can be used for vermicomposting however, *Gliricidia*, tobacco leaves and chicken droppings are not suitable for earthworm multiplication but can be composted with earthworms. The optimum temperature for vermicomposting is about 20–30°C and moisture content ranges from 32 to 60% only. It is a very simple process and easy to practice as well as cost-effective pollution abatement technology.

The training programs for women self-help groups (SHGs) covered technical aspects of making vermicompost and its application to various crops. These programs have been conducted by ICRISAT with support from the Asian Development Bank (ADB), Sir Dorabji Tata Trust and District Water Management Agency (DWMA) in Adarsha watershed (Kothapally) in Andhra Pradesh, Madhya Pradesh and eastern Rajasthan. A noxious weed, *Parthenium hysterophorus* (locally referred as *vayyari bhama* or congress weed) was found abundantly on field bunds in Kothapally and other regions of Andhra Pradesh, which inhibited the crop growth and caused environmental pollution. Some case studies of women who have come forward to utilize this weed as raw material for vermicomposting, a safe weed disposal mechanism, have been presented in this report.

The opinions expressed in this publication are those of the authors and do not necessarily reflect those of ICRISAT, ADB, Andhra Pradesh Rural Livelihoods Programme (APRLP) or Sir Dorabji Tata Trust. The designations employed and the presentation of material in this publication do not imply the expression of any opinion whatsoever on the part of ICRISAT, ADB, APRLP or Sir Dorabji Tata Trust concerning the legal status of any country, territory, city or area, or concerning the delimitation of its frontiers or boundaries. Where trade names are used, this does not constitute endorsement of or discrimination against any product by ICRISAT, ADB, APRLP or Sir Dorabji Tata Trust.

Global Theme on Agroecosystems Report no. 8

Vermicomposting: Recycling Wastes into Valuable Organic Fertilizer

KP Nagavallemma, SP Wani, Stephane Lacroix, VV Padmaja, C Vineela, M Babu Rao and KL Sahrawat



International Crops Research Institute for the Semi-Arid Tropics Patancheru 502 324, Andhra Pradesh, India

> Asian Development Bank 0401 Metro Manila 0980 Manila, The Philippines

Andhra Pradesh Rural Livelihoods Programme Hyderabad 500 030, Andhra Pradesh, India

> Sir Dorabji Tata Trust Mumbai 400 001, Maharashtra, India

> > 2004

About authors

KP Nagavallemma, Formerly Visiting Scientist, Global Theme on Agroecosystems, International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru 502 324, Andhra Pradesh, India

SP Wani, Principal Scientist (Watersheds) and Regional Theme Coordinator (Asia), Global Theme on Agroecosystems, ICRISAT, Patancheru 502 324, Andhra Pradesh, India

Stephane Lacroix, Formerly, Research Fellow, Global Theme on Agroecosystems, ICRISAT, Patancheru 502 324, Andhra Pradesh, India

VV Padmaja, Visiting Scientist, Global Theme on Agroecosystems, ICRISAT, Patancheru 502 324, Andhra Pradesh, India

C Vineela, Research Associate, Global Theme on Agroecosystems, ICRISAT, Patancheru 502 324, Andhra Pradesh, India

M Babu Rao, Scientific Associate, Global Theme on Agroecosystems, ICRISAT, Patancheru 502 324, Andhra Pradesh, India

KL Sahrawat, Visiting Scientist, Global Theme on Agroecosystems, ICRISAT, Patancheru 502 324, Andhra Pradesh, India

Acknowledgments

This report is based on the research conducted at ICRISAT, Patancheru and Kothapally village (Adarsha watershed), Ranga Reddy district, Andhra Pradesh, India. The contribution from farmers who are important partners in Adarsha watershed is greatly acknowledged. This work is a part of the project RETA-6067 "Farmer Participatory Watershed Management for Reducing Poverty and Land Degradation in SAT Asia", supported by the Asian Development Bank (ADB), the project "Combating Land Degradation and Increasing Productivity in Madhya Pradesh and Eastern Rajasthan" supported by Sir Dorabji Tata Trust and the watershed project of the Andhra Pradesh Rural Livelihoods Programme (APRLP) supported by the Government of Andhra Pradesh and Department for International Development (DFID), India. The support of these organizations is greatly acknowledged. Dr Radha D Kale, GKVK, University of Agricultural Sciences (UAS), Bangalore, India provided the earthworm culture to initiate these studies and her help is gratefully acknowledged. We also acknowledge Dr Vasantha Rao, Consultant for his contribution in identifying different fungal species. We are indebted to Ms Sheila Vijayakumar for editing the manuscript and Mr KNV Satyanarayana for incorporating the editorial corrections and page-setting the manuscript.

Contents

Background	
What is Vermicomposting?	
Importance of vermicompost	
Types of earthworms	
Earthworm multiplication	
Temperature changes during the process	
Methods of Vermicomposting	
Pits below the ground	
Heaping above the ground	
Tanks above the ground	
Cement rings	
Commercial model	
Materials Required for Vermicomposting	
Vermicompost Preparation	
Steps in the process	
Precautions during the process	
How to Use Vermicompost?	
Biodiversity in Vermicompost	
Vermicomposting: A Livelihood Micro-enterprise for Rural Women	
Case Studies	
Adarsha watershed, Kothapally	
APRLP watershed village14	
Tata-ICRISAT-ICAR project	
Conclusions	
Pafarancas 15	

Background

Environmental degradation is a major threat confronting the world, and the rampant use of chemical fertilizers contributes largely to the deterioration of the environment through depletion of fossil fuels, generation of carbon dioxide (CO₂) and contamination of water resources. It leads to loss of soil fertility due to imbalanced use of fertilizers that has adversely impacted agricultural productivity and causes soil degradation. Now there is a growing realization that the adoption of ecological and sustainable farming practices can only reverse the declining trend in the global productivity and environment protection (Aveyard 1988, Wani and Lee 1992, Wani et al. 1995).

On one hand tropical soils are deficient in all necessary plant nutrients and on the other hand large quantities of such nutrients contained in domestic wastes and agricultural byproducts are wasted. It is estimated that in cities and rural areas of India nearly 700 million t organic waste is generated annually which is either burned or land filled (Bhiday 1994). Such large quantities of organic wastes generated also pose a problem for safe disposal. Most of these organic residues are burned currently or used as land fillings. In nature's laboratory there are a number of organisms (micro and macro) that have the ability to convert organic waste into valuable resources containing plant nutrients and organic matter, which are critical for maintaining soil productivity. Microorganisms and earthworms are important biological organisms helping nature to maintain nutrient flows from one system to another and also minimize environmental degradation. The earthworm population is about 8–10 times higher in uncultivated area. This clearly indicates that earthworm population decreases with soil degradation and thus can be used as a sensitive indicator of soil degradation. In this report a simple biotechnological process, which could provide a 'win-win' solution to tackle the problem of safe disposal of waste as well as the most needed plant nutrients for sustainable productivity is described (Wani 2002).

What is Vermicomposting?

Vermicomposting is a simple biotechnological process of composting, in which certain species of earthworms are used to enhance the process of waste conversion and produce a better end product. Vermicomposting differs from composting in several ways (Gandhi et al. 1997). It is a mesophilic process, utilizing microorganisms and earthworms that are active at 10–32°C (not ambient temperature but temperature within the pile of moist organic material). The process is faster than composting; because the material passes through the earthworm gut, a significant but not yet fully understood transformation takes place, whereby the resulting earthworm castings (worm manure) are rich in microbial activity and plant growth regulators, and fortified with pest repellence attributes as well! In short, earthworms, through a type of biological alchemy, are capable of transforming garbage into 'gold' (Vermi Co 2001, Tara Crescent 2003).

Importance of vermicompost

Source of plant nutrients

Earthworms consume various organic wastes and reduce the volume by 40–60%. Each earthworm weighs about 0.5 to 0.6 g, eats waste equivalent to its body weight and produces cast equivalent to about 50% of the waste it consumes in a day. These worm castings have been analyzed for chemical and biological properties. The moisture content of castings ranges between 32 and 66% and the pH is

around 7.0. The worm castings contain higher percentage (nearly twofold) of both macro and micronutrients than the garden compost (Table 1).

Table 1.	Nutrient	composition of	vermicom	post and	garden	compost.
IUDIC I	1 dutilette	composition of	, crimicomi	post und	Suracii	compost

Nutrient element	Vermicompost (%)	Garden compost (%)
Organic carbon	9.8–13.4	12.2
Nitrogen	0.51-1.61	0.8
Phosphorus	0.19-1.02	0.35
Potassium	0.15-0.73	0.48
Calcium	1.18-7.61	2.27
Magnesium	0.093-0.568	0.57
Sodium	0.058-0.158	< 0.01
Zinc	0.0042-0.110	0.0012
Copper	0.0026-0.0048	0.0017
Iron	0.2050-1.3313	1.1690
Manganese	0.0105-0.2038	0.0414

From earlier studies also it is evident that vermicompost provides all nutrients in readily available form and also enhances uptake of nutrients by plants. Sreenivas et al. (2000) studied the integrated effect of application of fertilizer and vermicompost on soil available nitrozen (N) and uptake of ridge gourd (*Luffa acutangula*) at Rajendranagar, Andhra Pradesh, India. Soil available N increased significantly with increasing levels of vermicompost and highest N uptake was obtained at 50% of the recommended fertilizer rate plus 10 t ha⁻¹ vermicompost. Similarly, the uptake of N, phosphorus (P), potassium (K) and magnesium (Mg) by rice (*Oryza sativa*) plant was highest when fertilizer was applied in combination with vermicompost (Jadhav et al. 1997).

Plant growth promoting activity

Growth promoting activity of vermicompost was tested using a plant bioassay method. The plumule length of maize (*Zea mays*) seedling was measured 48 h after soaking in vermicompost water and in normal water. The marked difference in plumule length of maize seedlings indicated that plant growth promoting hormones are present in vermicompost (Table 2).

Table 2. Plumule length of maize seedlings.

Treatment	Initial length (cm)	Final length (cm)
Tank water	16.5	16.6
Vermicompost water	17.6	18.6

Improved crop growth and yield

Vermicompost plays a major role in improving growth and yield of different field crops, vegetables, flower and fruit crops. The application of vermicompost gave higher germination (93%) of mung bean (*Vigna radiata*) compared to the control (84%). Further, the growth and yield of mung bean was also significantly higher with vermicompost application. Likewise, in another pot experiment, the fresh and dry matter yields of cowpea (*Vigna unguiculata*) were higher when soil was amended with vermicompost than with biodigested slurry (Karmegam et al. 1999, Karmegam and Daniel 2000).

The efficiency of vermicompost was evaluated in a field study by Desai et al. (1999). They stated that the application of vermicompost along with fertilizer N gave higher dry matter (16.2 g plant⁻¹) and grain yield (3.6 t ha⁻¹) of wheat (*Triticum aestivum*) and higher dry matter yield (0.66 g plant⁻¹) of the following coriander (*Coriandrum sativum*) crop in sequential cropping system. Similarly, a positive response was obtained with the application of vermicompost to other field crops such as sorghum (*Sorghum bicolor*) (Patil and Sheelavantar 2000) and sunflower (*Helianthus annuus*) (Devi and Agarwal 1998, Devi et al. 1998).

Application of vermicompost at 5 t ha⁻¹ significantly increased yield of tomato (*Lycopersicon esculentum*) (5.8 t ha⁻¹) in farmers' fields in Adarsha watershed, Kothapally, Andhra Pradesh compared to control (3.5 t ha⁻¹). Similarly, greenhouse studies at Ohio State University in Columbus, Ohio, USA have indicated that vermicompost enhances transplant growth rate of vegetables. Amendment of vermicompost with a transplant grown without vermicompost had the highest amount of red marketable fruit at harvest. In addition, there were no symptoms of early blight lesions on the fruit at harvest. The yield of pea (*Pisum sativum*) was also higher with the application of vermicompost (10 t ha⁻¹) along with recommended N, P and K than with these fertilizers alone (Reddy et al. 1998). Vadiraj et al. (1998) reported that application of vermicompost produced herbage yields of coriander cultivars that were comparable to those obtained with chemical fertilizers.

The fresh weight of flowers such as *Chrysanthemum chinensis* increased with the application of different levels of vermicompost. Also, the number of flowers per plant (26), flower diameter (6 cm) and yield (0.5 t ha⁻¹) were maximum with the application of 10 t ha⁻¹ of vermicompost along with 50% of recommended dose of NPK fertilizer. However, the vase life of flowers (11 days) was high with the combined application of vermicompost at 15 t ha⁻¹ and 50% of recommended dose of NPK fertilizer (Nethra et al. 1999).

Reduction in soil C:N ratio

Vermicomposting converts household waste into compost within 30 days, reduces the C:N ratio and retains more N than the traditional methods of preparing composts (Gandhi et al. 1997). The C:N ratio of the unprocessed olive cake, vermicomposted olive cake and manure were 42, 29 and 11, respectively. Both the unprocessed olive cake and vermicomposted olive cake immobilized soil N throughout the study duration of 91 days. Cattle manure mineralized an appreciable amount of N during the study. The prolonged immobilization of soil N by the vermicomposted olive cake was attributed to the C:N ratio of 29 and to the recalcitrant nature of its C and N composition. The results suggest that for use of vermicomposted dry olive cake as an organic soil amendment, the management of vermicomposting process should be so adjusted as to ensure more favorable N mineralization-immobilization (Thompson and Nogales 1999).

Role in nitrogen cycle

Earthworms play an important role in the recycling of N in different agroecosystems, especially under *jhum* (shifting cultivation) where the use of agrochemicals is minimal. Bhadauria and Ramakrishnan (1996) reported that during the fallow period intervening between two crops at the same site in 5- to 15-year *jhum* system, earthworms participated in N cycle through cast-egestion, mucus production and dead tissue decomposition. Soil N losses were more pronounced over a period of 15-year *jhum* system. The total soil N made available for plant uptake was higher than the total input of N to the soil through the addition of slashed vegetation, inorganic and organic manure, recycled crop residues and weeds.

Improved soil physical, chemical and biological properties

Limited studies on vermicompost indicate that it increases macropore space ranging from 50 to 500 μ m, resulting in improved air-water relationship in the soil which favorably affect plant growth (Marinari et al. 2000). The application of organic matter including vermicompost favorably affects soil pH, microbial population and soil enzyme activities (Maheswarappa et al. 1999). It also reduces the proportion of water-soluble chemical species, which cause possible environmental contamination (Mitchell and Edwards 1997).

Types of earthworms

Earthworms are invertebrates. There are nearly 3600 types of earthworms in the world and they are mainly divided into two types: (1) burrowing; and (2) non-burrowing. The burrowing types *Pertima elongata* and *Pertima asiatica* live deep in the soil. On the other hand, the non-burrowing types *Eisenia fetida* and *Eudrilus eugenae* live in the upper layer of soil surface. The burrowing types are pale, 20 to 30 cm long and live for 15 years. The non-burrowing types are red or purple and 10 to 15 cm long but their life span is only 28 months.

The non-burrowing earthworms eat 10% soil and 90% organic waste materials; these convert the organic waste into vermicompost faster than the burrowing earthworms. They can tolerate temperatures ranging from 0 to 40°C but the regeneration capacity is more at 25 to 30°C and 40–45% moisture level in the pile. The burrowing type of earthworms come onto the soil surface only at night. These make holes in the soil up to a depth of 3.5 m and produce 5.6 kg casts by ingesting 90% soil and 10% organic waste.

Earthworm multiplication

Numerous organic materials have been evaluated for growth and reproduction of earthworms as these materials directly affect the efficacy of vermicompost. Nogales et al. (1999) evaluated the suitability of dry olive cake, municipal biosolids and cattle manure as substrates for vermicomposting. They reported that larger weights of newly hatched earthworms were obtained in substrate containing dry olive cake. In another study, maize straw was found to be the most suitable feed material compared to soybean (*Glycine max*) straw, wheat straw, chickpea (*Cicer arientinum*) straw and city refuse for the tropical epigeic earthworm, *Perionyx excavatus* (Manna et al. 1997).

Zajonc and Sidor (1990) evaluated and compared various non-standard materials for the preparation of vermicompost. A mixture of cotton waste with cattle manure in the ratio of 1:5 was found to be the best. The use of grape cake alone increased earthworm weight slightly. Tobacco (*Nicotiana tabacum*) waste, used as substrate, increased earthworm weight but the earthworms failed to reproduce. A mixture of tobacco waste with rabbit manure in the ratio of 1:5 was found to be lethal to the earthworms.

A multiplication trial was conducted at the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Andhra Pradesh with three kinds of earthworm cultures (Eisenia fetida, Eudrilus eugenae and Perionyx excavatus) using wheat straw, chickpea straw, tree leaves (Peltophorum sp) and Parthenium mixed with cow dung as feed materials. There was an increase in earthworm population and size during incubation for 90 days. The three types of earthworms multiplied 12 to 18 times when grown individually using legume tree leaves and cow dung mixture as

raw material (Table 3). However, mixed culture (of all three species) showed higher multiplication rate (27 times) than the individual species.

Further studies on earthworm multiplication were also conducted at ICRISAT using tree leaves and Gliricidia stems mixed with cattle manure as feed material (Table 4). The earthworm population decreased when grown in mixture of Gliricidia stems and cattle manure. These results indicated that Gliricidia loppings could not be used for multiplication of earthworms. Gliricidia bark is known to possess toxic properties as it is used as rat poisoning bait.

In another multiplication study at ICRISAT, there was maximum increase in earthworm population (570%) and weight (109%) when grown in a feed material containing tree leaves (3 kg) and cow dung (6 kg). In contrast, mortality of earthworms (about 7 to 22%) was observed by growing them in a feed material containing soil (Table 5).

All these studies indicated that Gliricidia and tobacco leaves are not suitable for multiplication of earthworms. Perhaps the alkaloids and other principal compounds present in these leaves may effect the survival of earthworms. Also, soil and rabbit manure should not be mixed with earthworm feed material.

Table 3. Multiplication trial of earthworm species at ICRISAT, Patancheru, India in 2000¹.

Earthworm species	Initial population	Final population	Increase (%)
Mixed culture	900	15950	1612 (27) ²
Eisenia fetida	90	1036	1051 (12)
Eudrilus eugenae	55	1007	1731 (18)
Perionyx excavatus	85	1192	1302 (14)

^{1.} Mixture of legume tree leaves and cow dung was used as substrate.

Table 4. Multiplication trials of earthworms using different organic materials at ICRISAT, Patancheru, India during 2000-02.

		Ini	tial	Fin	nal¹
Earthworm species	Feed material	Population	Weight (g)	Population	Weight (g)
Eisenia fetida	Tree leaves (15 kg)	345	20	2510	207
•	Cattle manure (15 kg)	510	207	1159	207
	Cattle manure (3 kg) +	1255	101	1000	50
	Gliricidia stem (6 kg)				
Eudrilus eugenae	Tree leaves (15 kg)	311	21	2986	334
	Cattle manure (15 kg)	2986	334	1522	216
	Cattle manure (3 kg) +	2707	230	2249	100
	Gliricidia stem (6 kg)				
Perionyx excavatus	Tree leaves (15 kg)	409	29	2707	230
-	Cattle manure (15 kg)	2707	230	2650	187
	Cattle manure (3 kg) +	3356	365	1000	50
	Gliricidia stem (6 kg)				
1. At 90 days after incuba	tion.				

^{2.} Values in parentheses indicate increase in number of times at 90 days after incubation.

Table 5. Multiplication trials of mixed culture of earthworms using soil and other organic substrates at ICRISAT, Patancheru, India, 2000–02.

	Initial		Final		Increase ¹ (%)	
Feed material	Number	Weight (g)	Number	Weight (g)	Number	Weight
Cow dung (15 kg)	500	89	750	163	50	83
Tree leaves (3 kg) + cow dung (3 kg)	500	95	1545	125	21	32
Tree leaves (3 kg) + cow dung (6 kg)	500	110	3351	230	570	109
Pigeonpea leaves + pod shells +	500	98	2230	187	346	90
tree leaves (2 kg) + cow dung (2 kg)						
Pigeonpea leaves + pod shells +	500	115	1490	193	198	68
tree leaves (2 kg) + cow dung (4 kg)						
Soil $(5 \text{ kg}) + \text{cow dung } (5 \text{ kg})$	1000	90	784	87	-22	-3
Soil $(5 \text{ kg}) + \text{cow dung } (5 \text{ kg}) +$	1000	75	1023	241	2	223
pigeonpea leaves (1 kg)						
Soil (5 kg) + cow dung (5 kg) +	1000	160	929	170	-7	-6
tree leaves (1 kg)						

1. At 90 days after incubation

Temperature changes during the process

Change in temperature was observed during the process of vermicomposting (from 5 to 65 days) with different farm residues (*Parthenium* and grass). In the beginning of the process, ie, up to 15 days, the temperature was high (32 to 33°C) in both *Parthenium* and grass substrates when compared to outside temperature (26 to 30°C). Later, there was a gradual decrease in temperature, which reached a minimum of about 24°C. However, higher temperature was recorded in *Parthenium* compost (decline from 32.8 to 27.5°C) than in grass compost (decline from 31.5 to 26.8°C) during the whole period of digestion process. Generally more heat was evolved from control treatment (without earthworms) than the vermicompost treatments (with earthworms). From these studies, it was suggested that the most suitable period for releasing the earthworms into organic residues would be between 15 and 20 days after heaping of the organic residues when the temperature is about 25°C (Fig. 1).

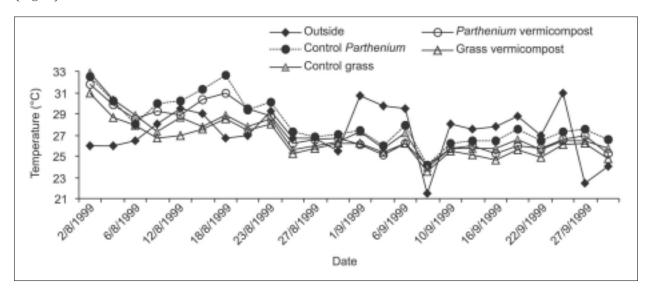


Figure 1. Temperature changes during biodigestion.

Methods of Vermicomposting

Pits below the ground

Pits made for vermicomposting are 1 m deep and 1.5 m wide. The length varies as required.

Heaping above the ground

The waste material is spread on a polythene sheet placed on the ground and then covered with cattle dung. Sunitha et al. (1997) compared the efficacy of pit and heap methods of preparing vermicompost under field conditions. Considering the biodegradation of wastes as the criterion, the heap method of preparing vermicompost was better than the pit method. Earthworm population was high in the heap method, with a 21-fold increase in *Eudrilus eugenae* as compared to 17-fold increase in the pit method. Biomass production was also higher in the heap method (46-fold increase) than in the pit method (31-fold). Consequent production of vermicompost was also higher in the heap method (51 kg) than in the pit method (40 kg).

Tanks above the ground

Tanks made up of different materials such as normal bricks, hollow bricks, shabaz stones, asbestos sheets and locally available rocks were evaluated for vermicompost preparation. Tanks can be constructed with the dimensions suitable for operations. At ICRISAT, we have evaluated tanks with dimensions of 1.5 m (5 feet) width, 4.5 m (15 feet) length and 0.9 m (3 feet) height. The commercial biodigester contains a partition wall with small holes to facilitate easy movement of earthworms from one tank to the other.

Cement rings

Vermicompost can also be prepared above the ground by using cement rings (ICRISAT and APRLP 2003). The size of the cement ring should be 90 cm in diameter and 30 cm in height. The details of preparing vermicompost by this method have been described in a later section.

Commercial model

The commercial model for vermicomposting developed by ICRISAT consists of four chambers enclosed by a wall (1.5 m width, 4.5 m length and 0.9 m height) (Fig. 2). The walls are made up of different materials such as normal bricks, hollow bricks, shabaz stones, asbestos sheets and locally available rocks. This model contains partition walls with small holes to facilitate easy movement of earthworms from one chamber to another. Providing an outlet at one corner of each chamber with a slight slope facilitates collection of excess water, which is reused later or used as earthworm leachate on crop. The outline of the commercial model is given in Figure 3.

The four components of a tank are filled with plant residues one after another. The first chamber is filled layer by layer along with cow dung and then earthworms are released. Then the second chamber is filled layer by layer. Once the contents in the first chamber are processed the earthworms move to chamber 2, which is already filled and ready for earthworms. This facilitates harvesting of decomposed material from the first chamber and also saves labor for harvesting and introducing earthworms. This technology reduces labor cost and saves water as well as time.



Figure 2. Commercial model for vermicomposting at ICRISAT.

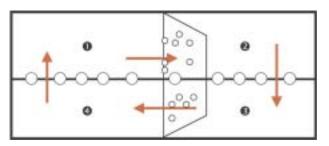


Figure 3. Diagrammatic representation of the commercial model with four chambers for vermicomposting.

Materials Required for Vermicomposting

A range of agricultural residues, all dry wastes, for example, sorghum straw and rice straw (after feeding cattle), dry leaves of crops and trees, pigeonpea (*Cajanus cajan*) stalks, groundnut (*Arachis hypogaea*) husk, soybean residues, vegetable wastes, weed (*Parthenium*) plants before flowering, fiber from coconut (*Cocos nucifera*) trees and sugarcane (*Saccharum officinarum*) trash can be converted into vermicompost. In addition, animal manures, dairy and poultry wastes, food industry wastes, municipal solid wastes, biogas sludge and bagasse from sugarcane factories also serve as good raw materials for vermicomposting.

The quantity of raw materials required using a cement ring of 90 cm in diameter and 30 cm in height or a pit or tank measuring $1.5 \text{ m} \times 1 \text{ m} \times 1 \text{ m}$ is given below:

Dry organic wastes (DOW) 50 kg
Dung slurry (DS) 15 kg
Rock phosphate (RP) 2 kg
Earthworms (EW) 500–700

Water (W) 5 L every three days

The various ingredients are used in the ratio of 5:1.5:0.2:50–75:0.5 of DOW:DS:RP:EW:W. In the tank or pit system 100 kg of raw material and 15–20 kg of cow dung are needed for each cubic meter of the bed.

Vermicompost Preparation

Steps in the process

Vermicomposting involves the following steps which are depicted in Figure 4(a-k):

- Cover the bottom of the cement ring with a layer of tiles or coconut husk or polythene sheet (Fig. 4a).
- Spread 15–20 cm layer of organic waste material on the polythene sheet (Fig. 4b). Sprinkle rock phosphate powder if available (it helps in improving nutritional quality of compost) on the waste material and then sprinkle cow dung slurry (Fig. 4c and d). Fill the ring completely in layers as described. Paste the top of the ring with soil or cow dung (Fig. 4e). Allow the material to decompose for 15 to 20 days.
- When the heat evolved during the decomposition of the materials has subsided (15–20 days after heaping), release selected earthworms (500 to 700) through the cracks developed (Fig. 4f).
- Cover the ring with wire mesh or gunny bag to prevent birds from picking the earthworms. Sprinkle water every three days to maintain adequate moisture and body temperature of the earthworms (Fig. 4g).

- The vermicompost is ready in about 2 months if agricultural waste is used and about 4 weeks if sericulture waste is used as substrate (Fig. 4h).
- The processed vermicompost is black, light in weight and free from bad odor.
- When the compost is ready, do not water for 2–3 days to make compost easy for sifting. Pile the compost in small heaps and leave under ambient conditions for a couple of hours when all the worms move down the heap in the bed (Fig. 4i). Separate upper portion of the manure and sieve the lower portion to separate the earthworms from the manure (Fig. 4j). The culture in the bed contains different stages of the earthworm's life cycle, namely, cocoons, juveniles and adults. Transfer this culture to fresh half decomposed feed material. The excess as well as big earthworms can be used for feeding fish or poultry. Pack the compost in bags and store the bags in a cool place (Fig. 4k).
- Prepare another pile about 20 days before removing the compost and repeat the process by following the same procedure as described above.

Precautions during the process

The following precautions should be taken during vermicomposting:

- The African species of earthworms, *Eisenia fetida* and *Eudrilus eugenae* are ideal for the preparation of vermicompost. Most Indian species are not suitable for the purpose.
- Only plant-based materials such as grass, leaves or vegetable peelings should be utilized in preparing vermicompost.
- Materials of animal origin such as eggshells, meat, bone, chicken droppings, etc are not suitable for preparing vermicompost.
- Gliricidia loppings and tobacco leaves are not suitable for rearing earthworms.
- The earthworms should be protected against birds, termites, ants and rats.
- Adequate moisture should be maintained during the process. Either stagnant water or lack of moisture could kill the earthworms.
- After completion of the process, the vermicompost should be removed from the bed at regular intervals and replaced by fresh waste materials.

How to Use Vermicompost?

- Vermicompost can be used for all crops: agricultural, horticultural, ornamental and vegetables at any stage of the crop.
- For general field crops: Around 2–3 t ha⁻¹ vermicompost is used by mixing with seed at the time of sowing or by row application when the seedlings are 12–15 cm in height. Normal irrigation is followed.
- For fruit trees: The amount of vermicompost ranges from 5 to 10 kg per tree depending on the age of the plant. For efficient application, a ring (15–18 cm deep) is made around the plant. A thin layer of dry cow dung and bone meal is spread along with 2–5 kg of vermicompost and water is sprayed on the surface after covering with soil.
- For vegetables: For raising seedlings to be transplanted, vermicompost at 1 t ha⁻¹ is applied in the nursery bed. This results in healthy and vigorous seedlings. But for transplants, vermicompost at the rate of 400–500 g per plant is applied initially at the time of planting and 45 days after planting (before irrigation).
- For flowers: Vermicompost is applied at 750–1000 kg ha⁻¹.
- For vegetable and flower crops vermicompost is applied around the base of the plant. It is then covered with soil and watered regularly.



Plastic sheet placed below the ring



Layer of raw material placed on polythene sheet



Rock phosphate powder sprinkled on organic material



Cow dung slurry



Cement ring sealed with cow dung

f

Earthworms are released near cracks

Figure 4(a–k). Vermicomposting process.



Cement ring covered with gunny bag



Processed vermicompost



Heaping of vermicompost



Compost sieved

Bag filled with vermicompost

Biodiversity in Vermicompost

In the present study, vermicompost samples were collected and analyzed for microbial diversity and population studies. The vermicompost samples were collected in sterile containers from the rings before harvesting the compost. To compare microbial diversity, samples from the partially decomposed dry organic waste material, ready for the release of the earthworms, were also collected and checked for diversity and population counts.

Total mircobial populations of bacteria, fungi and actinomycetes from the substrates were determined by using dilution plate techniques with suitable media (Nutrient Agar, Potato Dextrose Agar, Actinomycetes Isolation Agar-HI Media). The number of colony forming units (CFU) was expressed as CFU g⁻¹.

Several authors have noted that the earthworms play a major role in affecting populations of soil organisms, especially in causing changes in the soil microbial community (Coleman 1985, Parmelee 1998). The present work recorded higher microbial populations in the partially decomposed dry organic waste material for vermicompost than the vermicompost (Table 6). This may be due to the existing temperatures and pH in the partially decomposed raw material. But compared to conventional thermophilic composts, vermicompost is much richer in microbial diversity, populations and activities (Subler et al. 1998).

Table 6. Microbial populations from the samples of vermicompost.

	Bacteria (CFU g ⁻¹)	Fungi (CFU g ⁻¹)	Actinomycetes (CFU g ⁻¹)
Vermicompost	54×10^{6}	8×10^{4}	1×10^4
Partially decomposed dry organic waste	69×10^{6}	11×10^{4}	2×10^{4}
material for vermicompost			

The fungal isolates from the samples were identified upto species level (Table 7). Much diversity was observed between the two samples collected. Aspergillus, Fusarium, Mucor, Cladosporium, and Trichoderma were the common genera observed in both the samples. Genera like Absidia, and Stachbotrys were recorded in vermicompost. Genera like Alternaria, penicillium, and Thermomyces were isolated from partially decomposed dry organic waste material for vermicompost. This clearly indicates that the fungal diversity is more in the decomposed material than in the vermicompost. The digestive epithelium of the simple straight tubular gut of worms is known to secrete cellulase, amylase, invertase, protease, phosphatase (Ranganathan and Vinotha 1998). Earthworms inevitably consume the soil microbes during the ingestion of litter and soil. It has been recently estimated that earthworms necessarily have to feed on microbes, particularly fungi for their protein/nitrogen requirement (Ranganathan and Parthasarathi 2000). This may be the reason for the less diversity of fungi and microbial counts seen in the vermicompost collected.

In both the samples percentage of *Aspergillus* was more when compared with other genera. *Tricoderma* and *Penicillium* have antibiotic activities and can also be used as biological control on soil borne pathogens. Only a few studies have investigated that the suppression of soil borne plant pathogens by vermicompost (Szczech et al. 1993), or disease suppression in the presence of earthworms (Stephens and Davoren 1997, Stephens et al. 1994). Disease suppression by compost has been attributed to the activities of competitive or antagonistic microorganisms as well as the antibiotic compounds present in the vermicompost.

Table 7. List of fungi isolated from partially decomposed dry organic waste for vermicompost and vermicompost.

Partially decomposed dry organic waste	
for vermicompost	Vermicompost
Alternaria citri	Absidia cylindrospora
Aspergillus fumigatus	Aspergillus fumigatus
Aspergillus niger	Aspergillus niger
Aspergillus cervinus	Aspergillus clavoto nanicus
Aspergillus terreus	Aspergillus terreus
Aspergillus sydowii	Aspergillus sydowii
Aspergillus niveus	Aspergillus nidulans
Aspergillus sclerotiorum	Cladosporium herbarum
Cladosporium cladosporioides	Fusarium oxysporum
Cladosporium herbarum	Fusarium semitactum
Fusarium samucinum	Fusarium nivale
Fusarium dimerum	Mucor circinelloides
Mucor racemosus	Stachbotrys chartarum
Penicillium chrysogenum	Trichoderma viride
Penicillium thomii	
Penicillium citrinum	
Trichoderma viride	
Thermomyces lanuginous	

Vermicomposting: A Livelihood Micro-enterprise for Rural Women

ICRISAT with support from the Asian Development Bank (ADB), Philippines, District Water Management Agency (DWMA), Government of Andhra Pradesh and Tata-ICRISAT-ICAR project in northeastern regions of India was keen to promote the vermiculture technology. The primary objective of this project was to help women from rural areas to set up micro-enterprises based on vermiculture technology and also to improve crop productivity by increasing soil fertility through ecological methods of farming (Wani 2002).

The training program conducted by ICRISAT for DWACRA (Development of Women and Child in Rural Area) group of women and other women self-help groups (SHGs) covered technical aspects of multiplying earthworms, managing and collection of organic wastes, application of vermicompost for various crops, accounting and marketing. At the same time a noxious weed, *Parthenium hysterophorus* (locally referred as *vayyari bhama* or congress weed), was found abundantly in the fields as well as on field bunds, which inhibited crop growth and caused environmental pollution. Hence, the women have come forward to utilize this weed as raw material for vermicomposting, which is a safe weed disposal mechanism and an opportunity to convert into valuable compost.

Case Studies

Adarsha watershed, Kothapally

Ms Lakshmamma and four other women have set up a vermicomposting enterprise in a common place under one roof. Having begun with a population of 2,000 earthworms of three epigeic species, they regularly harvest around 400 kg of vermicompost every month collectively. Their work in making vermicompost is shared collectively and the unique marketing strategy involves meeting potential customers. Sometimes, they even get customers from distant places. They earn a net income of around Rs 500 each month. By becoming an earning member of the family, they are involved in the decision-making process in the family. This has also raised their status in the society.

APRLP watershed village

Ms Padmamma living in Sripuram, one of the thousand non-descript villages of Mahbubnagar district in Andhra Pradesh, leads a routine life and has never dreamt of a different life. She joined the women's SHG at the begining of the Andhra Pradesh Rural Livelihoods Programme (APRLP) project. Though reluctant during the initial stage, she started taking active part in the weekly meetings and showed interest in the discussions about raising income through small initiatives like adopting the vermicompost scheme. This scheme was introduced to enhance crop productivity in the fields and enable the farmers to get more per-hectare yield. Ms Padmamma is able to get higher yield from different crops such as maize and vegetables with the application of vermicompost in her own field. She now proudly displays the vermiculture beds to any visitor who comes to meet her.

Tata-ICRISAT-ICAR project

The farmers of Bundi nucleus watershed in Rajasthan, India have shown lot of interest in vermicomposting. Two farmers have built a multiple compartment system (commercial model) of vermicomposting while many are following the regular vermicomposting. In Guna nucleus watershed in Madhya Pradesh, nearly 35 farmers from all the three microwatersheds are practicing vermicomposting. Most of them are producing vermicompost on a large scale and are applying to their own fields for vegetable crops and getting higher yields with low-cost technology. A few farmers have already started selling their extra produce of vermicompost at the nearby market at the rate of Rs 5–7 per kg.

Conclusions

The production of degradable organic waste and its safe disposal becomes the current global problem. Meanwhile the rejuvenation of degraded soils by protecting topsoil and sustainability of productive soils is a major concern at the international level. Provision of a sustainable environment in the soil by amending with good quality organic soil additives enhances the water holding capacity and nutrient supplying capacity of soil and also the development of resistance in plants to pests and diseases. By reducing the time of humification process and by evolving the methods to minimize the loss of nutrients during the course of decomposition, the fantasy becomes fact. Earthworms can serve as tools to facilitate these functions. They serve as "nature's plowman" and form nature's gift to produce good humus, which is the most precious material to fulfill the nutritional needs of crops. The utilization of vermicompost results in several benefits to farmers, industries, environment and overall national economy.

To farmers:

- Less reliance on purchased inputs of nutrients leading to lower cost of production
- Increased soil productivity through improved soil quality
- Better quantity and quality of crops
- For landless people provides additional source of income generation

To industries:

Cost-effective pollution abatement technology

To environment:

• Wastes create no pollution, as they become valuable raw materials for enhancing soil fertility

To national economy:

- Boost to rural economy
- Savings in purchased inputs
- Less wasteland formation

References

Aveyard Jim. 1988. Land degradation: Changing attitudes - why? Journal of Soil Conservation, New South Wales 44:46–51.

Bhadauria T and **Ramakrishnan PS.** 1996. Role of earthworms in nitrogen cycle during the cropping phase of shifting agriculture (jhum) in northeast India. Biology and Fertility of Soils 22:350–354.

Bhiday MR. 1994. Earthworms in agriculture. Indian Farming 43(12):31–34.

Coleman D C. 1985. Through a red darkly: an ecological assessment of root soil microbial faunal interactions. Pages 1–21 *in* Ecological interaction in Soil (Fitter AH, Atkinson D, Read DJ and Usher MB, eds.). London, UK: Blackwell Scientific Publications.

Desai VR, Sabale RN and **Raundal PV.** 1999. Integrated nitrogen management in wheat-coriander cropping system. Journal of Maharasthra Agricultural Universities 24(3):273–275.

Devi D and **Agarwal SK.** 1998. Performance of sunflower hybrids as influenced by organic manure and fertilizer. Journal of Oilseeds Research 15(2):272–279.

Devi D, Agarwal SK and Dayal D. 1998. Response of sunflower [Helianthus annuus (L.)] to organic manures and fertilizers. Indian Journal of Agronomy 43(3):469–473.

Gandhi M, Sangwan V, Kapoor KK and **Dilbaghi N.** 1997. Composting of household wastes with and without earthworms. Environment and Ecology 15(2):432–434.

ICRISAT and **APRLP.** 2003. Vermicomposting: Conversion of organic wastes into valuable manure. Andhra Pradesh, India: ICRISAT and APRLP. 4 pp.

Jadhav AD, Talashilkar SC and Pawar AG. 1997. Influence of the conjunctive use of FYM, vermicompost and urea on growth and nutrient uptake in rice. Journal of Maharashtra Agricultural Universities 22(2):249–250.

Karmegam N, Alagermalai K and **Daniel T.** 1999. Effect of vermicompost on the growth and yield of greengram (*Phaseolus aureus* Rob.). Tropical Agriculture 76(2):143–146.

Karmegam N and **Daniel T**. 2000. Effect of biodigested slurry and vermicompost on the growth and yield of cowpea [*Vigna unguiculata* (L.)]. Environment and Ecology 18(2):367–370.

Maheswarappa HP, Nanjappa HV and Hegde MR. 1999. Influence of organic manures on yield of arrowroot, soil physico-chemical and biological properties when grown as intercrop in coconut garden. Annals of Agricultural Research 20(3):318–323.

Manna MC, Singh M, Kundu S, Tripathi AK and Takkar PN. 1997. Growth and reproduction of the vermicomposting earthworm *Perionyx excavatus* as influenced by food materials. Biology and Fertility of Soils 24(1):129–132.

Marinari S, Masciandaro G, Ceccanti B and Grego S. 2000. Influence of organic and mineral fertilisers on soil biological and physical properties. Bioresource Technology 72(1):9–17.

Mitchell A and Edwards CA. 1997. The production of vermicompost using *Eisenia fetida* from cattle manure. Soil Biology and Biochemistry 29:3–4.

Nethra NN, Jayaprasad KV and Kale RD. 1999. China aster [Callistephus chinensis (L)] cultivation using vermicompost as organic amendment. Crop Research, Hisar 17(2): 209–215.

Nogales R, Melgar R, Guerrero A, Lozada G, Beniteze E, Thompson R, Gomez M and Garvin MH. 1999. Growth and reproduction of *Eisenia andrei* in dry olive cake mixed with other organic wastes. Pedobiologia 43(6):744–752.

Parmelee RW, Bohlen PJ and Blair JM. 1998. Earthworms and nutrient cycling processes: intergrating across the ecological hierarchy. Pages 123–143 in Earthworm Ecology (Edwards CA, ed.). New York, USA: St Lucie Press.

Patil SL and Sheelavantar MN. 2000. Effect of moisture conservation practices, organic sources and nitrogen levels on yield, water use and root development of rabi sorghum [Sorghum bicolor (L.)] in the vertisols of semi-arid tropics. Annals of Agricultural Research 21(21):32–36.

Ranganathan LS and Parthasarathi K. 2000. Enhanced phosphatase activity in earthworm casts is more of microbial origin. Current Science 79: 1158–1159.

Ranganathan LS and Vinotha SP. 1998. Influence of pressmud on the enzymatic variations in the different reproductive stages of *Eudrilus eugeniae*. Current Science 74: 634–635.

Reddy R, Reddy MAN, Reddy YTN, Reddy NS, Anjanappa N and Reddy R. 1998. Effect of organic and inorganic sources of NPK on growth and yield of pea [Pisum sativum(L)]. Legume Research 21(1):57–60.

Sreenivas C, Muralidhar S and **Rao MS.** 2000. Vermicompost, a viable component of IPNSS in nitrogen nutrition of ridge gourd. Annals of Agricultural Research 21(1):108–113.

Sunitha ND, Giraddi RS, Kulkarni KA and Lingappa S. 1997. Evaluation methods of vermicomposting under open field conditions. Karnataka Journal of Agricultural Sciences 10(4): 987–990.

Stephens PM, Davoren CW, Ryder MH, Doube BM and **Correll RL.** 1994. Field evidence for reduced severity of *Rhizoctonia* bare-patch disease of wheat, due to the presence of the earthworms *Aporrectodea rosea* and *Aporrectodea trapezoides*. Soil Biology and Biochemistry 26(11): 1495–1500.

Subler S, Edwards C A and Metzger J. 1998. Comparing vermicomposts and composts. BioCycle 39: 63–66.

Szczech M, Rondomanski W, Brzeski MW, Smolinska U and Kotowski J. 1993. Suppressive effect of commercial earthworm compost on some root infecting pathogens of cabbage and tomato. Biological Agriculture and Horticulture 10(1): 47–52.

Tara Crescent. 2003. Vermicomposting. Development Alternatives (DA) Sustainable Livelihoods. (http://www.dainet.org/livelihoods/default.htm)

Thompson RB and Nogales R. 1999. Nitrogen and carbon mineralization in soil of vermicomposted and unprocessed dry olive cake ('Orujo seco') produced from two stage centrifugation for olive oil extraction. Journal of Environmental Science and Health, Part B, Pesticides, Food Contaminants and Agricultural Wastes 34(5):917–928.

Vadiraj BA, Siddagangaiah D and Potty SN. 1998. Response of coriander (*Coriandrum sativum L.*) cultivars to graded levels of vermicompost. Journal of Spices and Aromatic Crops 7(2):141–143.

Vermi Co. 2001. Vermicomposting technology for waste management and agriculture: an executive summary. (http://www.vermico.com/summary.htm) PO Box 2334, Grants Pass, OR 97528, USA: Vermi Co.

Wani SP. 2002. Improving the livelihoods: New partnerships for win-win solutions for natural resource management. Paper submitted in the 2nd International Agronomy Congress held at New Delhi, India during 26–30 November 2002.

Wani SP and Lee KK. 1992. Biofertilizers role in upland crops production. Pages 91–112 *in* Fertilizers, organic manures, recyclable wastes and biofertilisers (Tandon HLS, ed.). New Delhi, India: Fertilizer Development and Consultation Organisation.

Wani SP, Rupela OP and Lee KK. 1995. Sustainable agriculture in the semi-arid tropics through biological nitrogen fixation in grain legumes. Plant and Soil 174:29–49.

Zajonc I and **Sidor V**. 1990. Use of some wastes for vermicompost preparation and their influence on growth and reproduction of the earthworm *Eisenia fetida*. Pol'nohospodars-tvo (CSFR) 36(8):742–752.



ICRISAT-Bamako

Tel +223 2223375

Fax +223 2228683 icrisat-w-mali@cgiar.org

Bamako, Mali

BP 320

About ICRISAT



The International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) is a non-profit, non-political, international organization for science-based agricultural development. ICRISAT conducts research on sorghum, pearl millet, chickpea, pigeonpea and groundnut – crops that support the livelihoods of the poorest of the poor in the semi-arid tropics encompassing 48 countries. ICRISAT also shares information and knowledge through capacity building, publications and ICTs. Established in 1972, it is one of 15 Centers supported by the Consultative Group on International Agricultural Research (CGIAR).

ICRISAT-Nairobi

(Regional hub ESA)

Tel +254 20 524555 Fax +254 20 524001

icrisat-nairobi@cgiar.org

Contact information:

PO Box 39063, Nairobi, Kenya

ICRISAT-Patancheru (Headquarters)

Patancheru 502 324 Andhra Pradesh, India Tel +91 40 23296161 Fax +91 40 23241239 icrisat@cgiar.org

ICRISAT-Bulawayo

Matopos Research Station PO Box 776, Bulawayo, Zimbabwe Tel +263 83 8311-15 Fax +263 83 8253/8307 icrisatzw@cgiar.org

ICRISAT-Lilongwe

Chitedze Agricultural Research Station
PO Box 1096
Lilongwe, Malawi
Tel +265-1-707297/071/067/057
Fax +265-1-707298

icrisat-malawi@cgiar.org

ICRISAT-Maputo

ICRISAT-Niamey

icrisatsc@cgiar.org

BP 12404

(Regional hub WCA)

Niamey, Niger (Via Paris)

Tel +227 722529, 722725 Fax +227 734329

> c/o INIA, Av. das FPLM No 2698 Caixa Postal 1906 Maputo, Mozambique Tel +258-1-461657 Fax +258-1-461581 icrisatmoz@panintra.com

Visit us at www.icrisat.org