

International Arachis Newsletter (IAN)

Co-publishers



Peanut CRSP Peanut Collaborative Research Support Program (www.griffin.peachnet.edu/pnutcrsp.html)



About Peanut CRSP

The Peanut Collaborative Research Support Program is an international program supported by USAID Grant LAG-G-00-96-00013-00 to The University of Georgia. The research supported seeks environmentally sound, sustainable agriculture production and food delivery systems for peanut. The program has five thrusts addressing priority constraints to the global peanut industry (aflatoxin, production efficiency, socioeconomic forces, postharvest processing, and utilization). Peanut CRSP also works to foster human resource development and the communication of research results.

The Peanut CRSP provides support for collaborative research, training, and exchange of information through grants to 14 universities in USA linked to 15 host countries in the developing world. Both host countries and USA are expected to benefit from the activities of Peanut CRSP. Peanut CRSP actively collaborates with other organizations with interest in advancing development through the application of science and technology.

About ICRISAT

The International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) is a nonprofit, non-political organization that does innovative agricultural research and capacity building for sustainable development with a wide array of partners across the globe. ICRISAT's mission is to help empower 600 million poor people to overcome hunger, poverty and a degraded environment in the dry tropics through better agriculture. ICRISAT belongs to the Alliance of Centers of the Consultative Group on International Agricultural Research (CGIAR).

IAN Scientific Editor

SN Nigam

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Contents

From the Editor	 iii
News and Views	
News from West Africa	 1
CLAN Steering Committee Meeting Held in the Philippines	 1
The Peanut CRSP	 3
Current ICRISAT Groundnut Research and Integrated Projects	 7
Research Reports	
Genetic Resources and Enhancement	
Use of 2n Pollen in Generating Interspecific Derivatives of Groundnut Nalini Mallikarjuna and Sunil Kumar Tandra	 8
Arachis hoehnei, the Probable B Genome Donor of Arachis hypogaea Based on Crossability, Cytogenetical and Molecular Studies Nalini Mallikarjuna, Sunil Kumar Tandra and Deepak Jadhav	 10
Differences in Pod Characters Among Groundnut Cultivar L7-1 and its Chemical Mutants <i>Chuan Tang Wang, Xin Dao Yang, Jian Zhi Xu</i> and <i>Guang Zhen Liu</i>	 12
Seed Releases	
A High-yielding Drought-tolerant Groundnut Variety Abhaya RP Vasanthi, PV Reddy, V Jayalakshmi, P Sudhakar, M Asalatha, P Sudhakar Reddy, P Harinatha Naidu, T Muralikrishna, O Venkateswarlu, K John, MS Basu, SN Nigam, RC Nageswara Rao and GC Wright	 15
New High-yielding Groundnut Varieties GG 8 and GG 16 VK Poshiya, LK Dhaduk and RB Bhuva	 17
Performance of Cultivar ICGV 93468 During Summer Season in Uttar Pradesh, India <i>RA Singh</i>	 19
Release of Groundnut Variety Huayu 23 in Shandong Province in China Yu Shanlin, CaoYuliang, Min Ping, Jiao Kun and Yang Qingli	 21

Registration of Foliar Disease Resistant and High-yielding Groundnut Varieties ICGV 92099 and ICGV 90084	 22
Adams Frimpong, Francis Kwame Padi and James Kombiok	
BRS Havana: A New Early-maturing Groundnut Variety for the Northeast Region in Brazil RC dos Santos, RMM Freire and SR de Farias	 24
Pathology	
Collar Rot of Groundnut Caused by Lasiodiplodia theobromae in North Vietnam Chi Mai Thi Nguyen, Thanh Thi Vu Dang, Hong Xuan Nguyen, Chinh Thi Nguyen, Thuy Thu Thi Le and Paul Holford	 25
Entomology	
White Grub Species Attacking Groundnut in the Saurashtra Region in Gujarat, India MN Kapadia, PG Butani and NN Beria	 28
Occurrence of White Grubs in Groundnut Crop in Uplands of South Vietnam: A New Report GV Ranga Rao, Ngo Thi Lam Giang, Phan Lieu and Nguyen Thi Hoai Tram	 29
Assessment of Integrated Pest Management Modules in Groundnut on Farmers' Fields DA Shambharkar, PK Dharne, TM Bahale, Anjali Deshmukh, RT Surywanshi and RB Jadhav	 31
Agronomy/Physiology	
Companion Cropping of Spring Sugarcane and Summer Groundnut – A New Cropping System for Uttar Pradesh, India <i>RA Singh</i>	 34
An Expert System for Cultivation and Management of Groundnut Kun Zhang, Yongshan Wan and Fengzhen Liu	 35
Utilization	
Quality Attributes of Peanut Butter Prepared from Some Indian Groundnut Cultivars JB Misra, Veena Girdhar, Vimal K Jain and Narinder K Dhamsaniya	 38

Publications

Publications from ICRISAT	•••••	41
SATSource Listing	•••••	42

From the Editor

Dear Readers

ICRISAT has launched an electronic journal, SAT Agricultural Research, which can be accessed at http://www.icrisat.org/journal/. Its first issue came out in 2005. The journal accepts direct contributions besides publishing selected articles from the three newsletters, International *Arachis* Newsletter (IAN), International Chickpea and Pigeonpea Newsletter (ICPN) and International Sorghum and Millets Newsletter (ISMN) co-published by ICRISAT.

Although IAN accepted in the past only short articles based on preliminary results, it is now ready to accept a few high quality full length papers in each issue. Short review articles on emerging sciences/tools will also be welcome. The readers are encouraged to contribute to News and Views section of the IAN their interesting observations, happenings, episodes and stories, recipes and people associated with groundnut in their region. It is important to widen the scope of IAN to sustain the interest of groundnut community in the newsletter. The authors are advised not to submit articles based on the results of varietal evaluation alone and inconclusive trials/experiments. The articles submitted to IAN are peer reviewed. When needed, I will be requesting some of you to review the manuscript submitted to IAN. Your contribution will be duly acknowledged in IAN. Your help in reviewing and improving the manuscript will go a long way in enhancing the quality of articles appearing in IAN and also it would provide guidance to young researchers in research paper writing.

Last year we sent 1700 copies of IAN 25 to members and libraries (as per the existing mailing list of 2005) with a request to state their interest to receive future issues of the newsletter. So far we have received only 406 responses, some of them from libraries. Therefore we have decided to send IAN 26 only to these respondents. We once again request libraries and other readers to indicate their willingness at <u>newsletter@cgiar.org</u> if they would like to receive future issues of IAN. It will help us to minimize the cost of printing and mailing.

I would like to acknowledge R Aruna, Jayashree Balaji, BR Ntare, Piara Singh, GV Ranga Rao, KL Sahrawat, RP Thakur, V Vadez (ICRISAT); MS Basu and T Radhakrishnan [National Research Centre for Groundnut (NRCG), Junagadh, Gujarat, India]; and GV Subbaratnam [Acharya NG Ranga Agricultural University (ANGRAU), Hyderabad, India] who reviewed IAN articles and ICRISAT library for compiling SATSource listing.

The festive season in India continues. We celebrated Deepavali and Ramazan in October and now are awaiting X-mas in December. I wish all the readers merry X-mas and a very happy new year.

SN Nigam

News and Views

News from West Africa

CFC-ICRISAT-FAO groundnut seed project gets into fourth year

The CFC-ICRISAT-FAO project on Development of Sustainable Groundnut Seed Systems in West Africa, commonly known as groudnnut seed project (GSP), has successfully completed the third year of operation. This project aims at promoting utilization and uptake of improved groundnut varieties responding to market requirements through the development of sustainable community-based seed systems; promoting measures to minimize aflatoxin contamination; improving skills of farmers and other entrepreneurs in seed production, delivery, processing, marketing and small-scale seed enterprise management; and improving the flow of information between farmers and market intermediaries.

The project partners from Mali, Niger, Nigeria, Senegal, ICRISAT, FAO and CFC met for the annual project coordination and planning meeting at Bamako during 25–27 April 2006. BR Ntare (Country Representative and Project Manager), F Waliyar (PEA representative), Peter Thoenes and Robert Guei (FAO representatives) and Sieste van der Werff (CFC representative) gave opening remarks of welcome. The Deputy Director General of Institut d'Economie Rurale (IER) inaugurated the meeting and hailed the progress made by making available improved groundnut varieties, the efforts made to minimize aflatoxin contamination to improve quality and marketability, the efforts made to enhance skills of farmers and other entrepreneurs, and the initiation of community-based seed production and distribution systems in pilot areas.

ICRISAT technical support is helping farmers overcome the problem of access to improved varieties and also providing more suitable varieties. It is also providing socioeconomic support through targeted studies. The project partners reviewed progress achieved, the constraints encountered and strategies for sustaining the achievements. After an in-depth review of the various reports, together with ICRISAT, the partners prepared a comprehensive work plan and budget considering the sustainability of the achievements after the end of the project in 2007.

New groundnut varieties empowering women farmers in Mali

A 3-year long participatory research in the selection and evaluation of improved groundnut varieties has yielded positive results. Women groundnut farmers in the village of Wakoro in Mali have selected the varieties ICGV 86124. Fleur 11 and JL 24, which produce high quality seed. They are high yielding, mature early and are a sure way of increasing income and food security in Wakoro. The program which started with only 5 women farmers in this district has inspired 195 women farmers who are growing the three varieties in plots ranging from 0.25 to 2.0 ha, an indication of adoption process of groundnut varieties. The women are organized into a groundnut farmers' association and have taken up groundnut growing as a business. Similar progress is being made in other pilot areas where farmer groups, especially women are being empowered to grow quality groundnut seed as one of the strategies to increase availability of quality seed countrywide.

> *Contributed by:* BR Ntare ICRISAT Bamako, Mali

CLAN Steering Committee Meeting Held in the Philippines

The Eighth Cereals and Legumes Asia Network (CLAN) Steering Committee Meeting was held at the Central Luzon State University (CLSU), Science City of Munoz, Nueva Ecija, Philippines during 4–6 November 2005. It was co-sponsored by ICRISAT, ICARDA, AVRDC and APAARI, and co-hosted by Philippine institutions, namely, CLSU, Philippine Council for Agriculture, Forestry and Natural Resources Research and Development (PCARRD), and the Bureau of Agricultural Research (BAR), Philippines. All CLAN Country Coordinators (except India and Yemen) participated along with representatives of AVRDC, ICARDA, ILRI, IRRI, ICRISAT and APAARI. In addition, there were 20 observers from Philippines national program. JE Eusebio, Director – Crops Research Division, PCARRD, Philippines was elected as Chairperson of CLAN Steering Committee for 2006–07. SH Sabaghpour (Iran) was elected Deputy Chair.

The Steering Committee reviewed the 2004–05 accomplishments of the network in the areas of germplasm exchange, varieties released, training, exchange of scientists, and adoption and impact of technology. The meeting noted substantial progress in the number of germplasm samples, breeding lines and sets of trials/nurseries on CLAN mandated crops supplied by ICRISAT (sorghum, pearl millet, chickpea, pigeonpea and groundnut), AVRDC (mung bean) and ICARDA (lentil) to member countries.

Considering the role of crop-livestock systems for sustainable agriculture in Asia, the Steering Committee requested ILRI to join the network as one of the cofacilitators along with ICRISAT, ICARDA and AVRDC. CLAN membership consists of 13 countries in Asia, namely, Bangladesh, China, India, Iran, Indonesia, Myanmar, Nepal, Pakistan, the Philippines, Sri Lanka, Thailand, Vietnam and Yemen. ICRISAT, AVRDC, ICARDA and other regional and international institutes in the Asia-Pacific region are a part of the network, providing genetic material, technology and research information, and training input.

The expanded CLAN is now co-facilitated by ICRISAT, ICARDA and AVRDC. The coordination unit is located at and supported by ICRISAT-Patancheru. APAARI has committed support to help sustain the network activities.

The participants reviewed the ICRISAT's vision and strategy to 2015, and offered feedback, comments and suggestions to enhance the document. A concept note on "Crop diversification with food legumes for improving income and nutrition of rural poor, and sustainable productivity of cereal-based cropping systems in South and Central Asia" was discussed and endorsed for submission to the International Fund for Agricultural Development (IFAD) for funding.

Varieties release	ed in CLAN count	tries during 2005–06.		
Crop	Country	ICRISAT name	Release name	Year
Sorghum (1)	India (1)	NSSH 104	NSSH 104	2005
Pearl millet (4)	India (4)	HHB 67-2 (improved) Sagar 205 PHB 2168 GICV 98771	HHB 67-2 Sagar 205 PHB 2168 JBV 4	2005 2005 2006 2006
Chickpea (4)	India (4)	ICCV 88202 ICCV 96329 (LBeG-7) ICCV 95332 ICCV 93952	Pratap Chana 1 Lam Senaga JGK 2 (Jawahar Gram Kabuli 2) JAKI 9218	2005 2006 2006 2006
Pigeonpea (4)	India (4)	ICPL 88039 ICPL 99050 ICP 7035 ICPL 13092	JK Champion (JKPL 2) JK Sania (JKPL 3) JK Sweety (JKPL 5) JK Sixer (JKPL 6)	2005 2005 2005 2005
Groundnut (4)	India (2)	ICGV 92195 ICGV 91114	Pratap Mungphali-2 ICGV 91114	2005 2006
	Nepal (2)	ICGV 86300 ICGV 90173	Rajarshi Baidehi	2005 2005

Other CLAN-related activities

Contributed by: CLL Gowda ICRISAT Patancheru, India

The Peanut CRSP

The Peanut Collaborative Research Support Program (CRSP) was established in 1982, and the present grantperiod will end in 2007. The goal of the program is to link US agricultural universities with research institutions in developing countries to enhance the role of peanut (groundnut) in food production and economic development. The CRSPs implement a portion of Title XII of the US Congress and are funded through USAID by grants to US universities. The Peanut CRSP is managed by the University of Georgia and involves a number of US universities and host/developing countries. Linkages exist between other institutions such as ICRISAT to assure development goals are met. In late 2004 and early 2005 an External Evaluation Panel reviewed the Peanut CRSP and concluded that it had been highly effective in developing technologies through research in the five thematic/cluster areas, across the whole value chain; and the technologies transferred to farmers, entrepreneurs and key stakeholders have resulted in significant impacts in the host countries and the US. Long- and short-term training, institutional capacity development and information dissemination has been accomplished. Notable is the development and use of a Web-based program and fiscal management system that has been efficient and cost-effective in the timely management of a worldwide program. Impacts, achievements and mechanisms for technology transfer within the five thematic/cluster areas were identified and evaluated for their importance in the host countries and USA, and for the benefits to women who are often peanut farmers and village-level or small-scale peanut-food processors. These impacts and achievements are summarized as follows.

Market driven development

The Peanut CRSP has over the past increased the emphasis on enhancing demand for peanuts in the market place. Too often we have seen that a production technology is initially adopted, farmers do well for a while until the production increases significantly and then the prices collapse leaving discouraged farmers. Sustainable development requires a balanced development of consumption to provide a market pull and crop technologies to respond to this situation.

A significant achievement in the Philippines has been the development of strong partnership with the food industry. The partnership was described as the Peanut Industry Incubator Model (PIIM) and identifies and solves peanut

industry problems, and transfers relevant Peanut CRSP technologies to the user. This model requires that the research institution and private food industry partners agree on the projects to be developed early through intensive interactions, and allows the food industry to access the public research capacity and technologies while cost-sharing where resources allow. The model has now been applied in other developing countries with some modification to consider country-specific situations. Among the first successes of the PIIM was the co-development of vitamin A-fortified peanut butter. Successful marketing of the product resulted in a 37% increase in peanut butter production by the partner company in the Metro-Manila area; this result was established through impact studies conducted in the past phase of the CRSP. Children who are most a risk of vitamin A deficiency were the highest consumers.

The PIIM resulted in the adoption of hand-sorting technology by a company to assure aflatoxin-free peanuts for production of a peanut sauce ("Kare-kare") and led to the company entering the export market, with significant economic returns from increasing export volumes. The PIIM trained women's cooperatives in the central Philippines to improve the quality and packaging of a peanut candy product and obtain significant increase in their income because of expanding their market from bus stops to access to the Manila supermarkets. Similar results were obtained in Thailand among village-level peanut processors. In Thailand, villages generally concentrate on processing of one product for the market. Market pull was a key factor in the successful transfer of the technologies developed by research. In Europe a honey-coated roasted peanut product developed by a PIIM effort should be on the market in Bulgaria in 2006.

Production driven/market sustained development

Improved cropping practices and increased areas have doubled yields and caused five-fold increase in peanut production in Guyana. Before market forces depressed prices and farmers became discouraged, the local collaborator (Beacon Foundation, an NGO), the US scientists and the local government worked together in the Rupununi region to develop a school lunch program. A pilot program based on seven villages which produced peanut butter and cassava bread for their schools was found to be highly successful and is now being expanded to a much larger number of schools. This PIIM-like market development effort which connects the producer to markets is helping to sustain profitability to the growers.

Health induced market development

Nutrition research showed that peanut consumption is associated with improved blood-lipid profiles and reduced cardiovascular disease risk and has provided critical evidence that peanuts have a satiety factor that offsets the high-energy content making the food neutral for obesity. The initial Peanut CRSP research inspired other research and the pool of information contributed to the US Food and Drug Administration (FDA) awarding a "heart healthy" claim for peanuts (including some tree nuts). Peanut CRSP nutrition research contributed to the reversal of an 18% reduction in peanut consumption in USA during the 1990s, and current sales of peanut products are increasing more than 10% annually. The value of industries promoting the health benefits of peanut as a food is something that can help develop the peanut industry on a global scale.

Preventing human aflatoxicosis – food safety and development

Peanut is well recognized as having significant aflatoxin problem. Indeed this problem is recognized as a major barrier to the trade in peanut; but very little has been done to protect the people living in countries where foods cannot be exported because of the levels of contamination. The Peanut CRSP focused on preventing human aflatoxicosis, which requires an integrated approach to preventing contamination in the field, at harvest and during storage. Decontamination and protection of consumers are also viable strategies in the management of the problem.

Our plan focused on determining the levels of exposure in developing countries, in determining what the health consequences of that exposure would be, and in the risks of different interventions to prevent exposure. Exposure is very widespread; in our and other studies everyone had biomarkers for exposure.

Our studies of interventions using a toxin binding additive have just been completed. These studies are based on evaluating a specialized clay (NovaSil-TM) that was found in earlier Peanut CRSP research and which is highly adsorptive of aflatoxins in the digestive tract of the animal. NovaSil as a feed additive (0.5% of the feed) binds aflatoxin and prevents adsorption, metabolism and subsequent aflatoxicosis in animals and is being adopted for livestock worldwide. Research to transfer this to human application was added in 2001. A study showed that lifetime exposure to NovaSil was harmless to rats. Based on these results, a human study was conducted in USA that showed no adverse nutritional effects from consuming the clay. Although aflatoxins have been shown to be immune-suppressive agents in animals, the potential role of aflatoxins in modifying the distribution and function of leukoctye subsets in humans has never been assessed. The cellular immune status of a group of Ghanaians was examined in relation to levels of aflatoxin B_1 -albumin adducts in plasma, and the alterations shown in immunological parameters in participants with high aflatoxin B_1 levels could result in impairments in cellular immunity that could decrease host resistance to infections. Results are near completion from a subsequent study in Ghana to measure the effect of NovaSil to reduce the ingestion/metabolism of aflatoxin in humans.

Gender studies in Uganda show that women farmers and housewives have no knowledge of aflatoxin, showing the need for extensive outreach and education. The socioeconomics project in Ghana has trained 900 professionals in three workshops to increase awareness of the aflatoxin problems. Producers, consumers and processors (male and female) will be familiar with the prevalence and health effects of aflatoxins, and available interventions to manage contamination.

Production driven development

The experience in Bolivia shows that for adoption of peanut production technologies by farmers, research has to be complemented by a strong technology transfer effort and a seed production program, which connects multiple sectors of the whole value chain. Participation of technology transfer/extension institutions and farmers' associations with the research efforts has facilitated the access of farmers to new peanut varieties, management practices and information. It also generated the interest of the Bolivian government that increased the priority of peanut in its agricultural development plan. Local consumption and exports are presently providing adequate markets to use the double yields and expanded production areas that are a result of the new technologies adopted by the farmers.

In Ghana, farmers (50% are women) who adopted environmentally-friendly integrated pest management (IPM) practices have increased yields two-fold. The IPM technology has been transferred through farmer field schools, television, radio and other extension means. In Ghana and Benin, crop models have identified major constraints to production and showed that yield increase and reduced cost of production are attainable. Rosette virus has been a devastating disease of peanuts for several years in Africa. Resistant varieties have been long season and not adapted to all the affected production systems. In Malawi, the national agricultural research programs released rosette-resistant, shortseason cultivars bred by ICRISAT and tested by Peanut CRSP participating scientists, extending the use of resistant varieties to short-season environments. CRSP/ Uganda collaboration transferred the varieties to Uganda where it was estimated that the new varieties, when fully adopted by farmers, could contribute about US\$47 million annually to the economy.

The collaborative research has altered oil quality and increased product shelf life. New cultivars with disease resistance, seed dormancy, and oil quality (high oleic acid to linoleic acid ratio) that increases shelf life are near release in Senegal and Burkina Faso. A 'high oleic' variety was released in Texas, which through reduced rancidity is benefiting processors and consumers by extending the shelf life of peanut products. US processors have said that this is one of the most important technologies made available to the processing industry in many years.

Other variety development contributions have been realized. Impact studies in North Carolina show continued benefit from cylindrocladium black rot (CBR) disease resistant varieties developed earlier with Peanut CRSP support. Cultivars introduced from Bulgaria yield 5–20% more than the local Valencia types in New Mexico, and should have short-term impact on variety development. Germplasm exchanges between Bolivia and Georgia/Florida have resulted in the development of higher yielding, disease resistant varieties in Bolivia and the US. In Florida, genetic marker research is identifying genes for drought tolerance and other traits that have potential to decrease the time and cost for developing new varieties.

In North Carolina, farmers who adopted the tomato spotted wilt virus (TSWV) index reduced virus incidence by 50% in a single year. Since its adoption, there has only been one year of significant virus incidence. Also, the adoption of the southern corn rootworm (SCR) advisory index by farmers and extension agents has reduced pest damage by 50% per year. The SCR treatment of preventive applications of insecticides has been reduced to only "high risk" areas. These environmentally-friendly advisory programs utilized Peanut CRSP research outputs.

Socioeconomic, gender and policy research

Significant socioeconomic impacts that related to gender concerns and aflatoxin awareness were observed. Economic impact studies have documented the impacts of variety adoption and IPM practices in North Carolina, Thailand, Malawi, Uganda, Senegal and the Philippines. Peanut CRSP also documented the economic and health benefits of vitamin A-fortified peanut butter and aflatoxin-free peanut sauces in the Philippines. Poverty could be reduced by 1.3% through the full adoption of the rosette resistant varieties released in Uganda.

In Senegal, impact studies have shown a 25% yield increase when farmers adopted new varieties developed earlier by the Peanut CRSP. Socioeconomic data in Senegal resulted in an increased number of publications on the impact of new varieties, pricing and marketing of peanuts, optimizing farm planning to reduce poverty, and peanut production and processing. The capacity of the host country institution was greatly increased in socioeconomic research and in the desire to publish information.

Postharvest and utilization researchdevelopment potentials

In USA, a patent is pending for 2005 for peanut enhanced with resveratrol. Peanut processors, globally, will benefit from the introduction of resveratrol-enhanced peanuts for use in many products. Consumers will benefit from the nutraceutical peanuts with their anti-cancer and anticardiovascular disease properties.

Production of new high-protein food products and other nutraceuticals from peanut processing by-products resulting from Peanut CRSP research has potential to add value to the peanut industry worldwide. It could also meet the fast growing demand for meat substitutes in vegetarian diets. The meat analog industry in USA is growing rapidly providing an opportunity to increase peanut market demand. Peanut-derived nutraceuticals will also tap into the large and growing nutraceuticals and functional food market, currently estimated at US\$17 billion per year.

Information, training and technology transfer

In the CRSP model every project has a role in capacity development and technology transfer. In addition to this we have had a number of projects dedicated to these objectives.

A Peanut CRSP developed Web-based World Geography of Peanut (http://lanra.anthro.uga.edu/ peanut/knowledgebase) is a significant repository for worldwide peanut publications and information. It includes data on the status of peanut production and industry in many countries, with potential use in policy making.

A technology transfer project in Thailand continues the partnership with this USAID "graduate country" in regional training efforts, resulting in Thailand being a center of excellence for training of trainers. Frequent workshops, largely attended by women, focus on product development and food safety practices. The program is also reaching many villages in Thailand and assisting women entrepreneurs in improving the production and marketing of peanut food products. Most villages follow the "one village-one product" scheme of processing, fostered by the Thai Princess' development program for poor villages, utilizing Peanut CRSP-developed technologies. Capacity development results from projects providing fiscal support for equipment and supplies and by training activities. Long-term, degree training of host country personnel is usually done at the collaborating US university. Short-term training has been done in the US and at ICRISAT, as well as the regional efforts such as cited above for Thailand.

The importance of multiple-institutional involvement in technology transfer was seen in the host country institution/CRSP/ICRISAT effort in developing and introducing to farmers the new, rosette resistant peanut varieties in Malawi and Uganda. Similar cooperation led to the success in technology transfer in Bolivia and Guyana, and the success of the PIIM in the Philippines, Thailand and Bulgaria. The close relationship between the research and extension programs and the farmers contribute to the success in the reduction of incidence of TSWV and SCR in North Carolina.

> *Contributed by:* JH Williams Peanut CRSP, 1109 Experiment Street Griffin, GA 30223-1797, USA

		Project	Grant	
Investor	Project title	coordinator	(US\$ '000)	Duration
Australia/ACIAR	Regional Workshop on Minimizing Aflatoxin Risk in Peanuts	SN Nigam	22	2006
Common Fund for Commodities	Development of sustainable groundnut seed systems in West Africa	F Waliyar B Ntare	2,103	Apr 2003– Mar 2007
Canada/CIDA	An aflatoxin risk early warning system to improve nutrition, health and income in West African smallholder farms	PCS Traore	188	Apr 2006– Mar 2009
CGIAR/ICARDA/CAC	Research activities on groundnut and on management of drought in chickpea, targeted to the Central Asia and the Caucasus (CAC) region	SN Nigam	34	2001–2006
CGIAR Global Challenge Program – HarvestPlus	Genetic engineering of groundnut for enhanced β -carotene production to combat vitamin A deficiency in the semi-arid tropics	KK Sharma	170	Nov 2003– Dec 2006
GCIAR – Generation Challenge Program – CIMMYT/EMBRAPA	Unlocking the genetic diversity in peanut's wild relatives with genomic and genetic tools	V Vadez	276	Jul 2005– Dec 2007
CGIAR/IFAR	Identification and field-testing of salinity tolerant groundnut in saline areas of India	Namita Srivastava V Vadez	11	2006
ndia/MoA&C	Development and popularization of 'Model' seed system(s) for quality seed production of major legumes to ensure seed-sufficiency at the village level	SN Nigam	1,000	2006–2010
ndia/ISOPOM/DoA&C	Farmers' participatory groundnut improvement in rainfed cropping system	SN Nigam	109	2005-2008
ndia/Govt. of AP	Establishment of aflatoxin testing laboratory at Anantapur	F Waliyar	49	2005-2006
nternational Fund for Agricultural Development IFAD)	Farmer-participatory improvement of grain legumes in rainfed Asia	SN Nigam	1,300	Sep 2001– Dec 2006
McKnight Foundation	ALIVE and nutritious cropping systems: A participatory approach to legume intensification and variety enhancement	E Weltzien-Rattund R Tabo	le 251	Mar 2006– Feb 2010
AcKnight Foundation	Developing short and medium duration groundnut varieties with improved yield performance, acceptable market traits and resistance to foliar diseases	EM Monyo	194	2006–2010
Norway/Development Fund	Enhancing groundnut production in the non-traditional and dry-land areas of Malawi for improved nutrition and poverty reduction	M Siambi	180	Jan 2004– Dec 2006
Philippines	Enhancing adoption of ICRISAT legume varieties and technologies in the Philippines	CLL Gowda	50	Jul 2004– Jun 2007
	Introduction, promotion and efficient seed support system of ICRISAT 'Asha' peanut variety in Region 2, Philippines	SN Nigam	55	Apr 2005– Apr 2007
JSA/Univ of Georgia Peanut CRSP)	Peanut CRSP support for regional workshop and publications	F Waliyar	141	2000-2006
JSAID/US Univ Linkages – Jniv of Georgia	Quantifying yield gaps and abiotic stresses in soybean- and groundnut-based production systems	P Pathak	90	2001-2006
JSANID/US Univ Linkages - Jniv of Georgia	- Management of aflatoxin in peanut through the use of atoxigenic strains of <i>Aspergillus flavus</i>	RB Jones	60	Jan 2005– Dec 2006
JSAID/US Univ Linkages – Jniv of Wisconsin-Madison	Elucidation of the peanut/Aspergillus interaction	F Waliyar	60	Jan 2005– Dec 2006
USAID/ABSP II (Sathguru Consultants)	Development of tobacco streak virus resistant sunflower and groundnut	KK Sharma	50	Apr 2005– Sep 2007

Current ICRISAT Groundnut Research and Integrated Projects

Research Reports

Genetic Resources and Enhancement

Use of 2n Pollen in Generating Interspecific Derivatives of Groundnut

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Numerically unreduced gamete called 2n pollen is a product of meiosis that bears sporophytic rather than the gametophytic chromosome number. Abnormalities in the division during meiosis or during spore wall formation result in 2n pollen. Often, such pollen are fertile (Christopher 1971).

The presence of dyads and triads at the microspore tetrad stage indicates the presence of 2n gametes. One of the main reasons for 2n pollen formation is meiotic nuclear restitution, which was first proposed by Rosenberg (1927). It is defined as the formation of a single nucleus with unreduced chromosome number, and the failure of the first or the second meiotic division. In the first division restitution, abnormal meiosis takes place with the formation of many univalents, and according to Wagennar (1968), it is a cellular mechanism for terminating the prolonged first division. Nevertheless the resultant restitution forms unreduced pollen. Restitution following

the second meiotic division in pollen formation also yields 2n pollen. In some plant species there can be double restitution, although rarely, resulting in the formation of giant pollen. Triad formation occurs as a result of second division restitution (Sosa and Hernandez de Sosa 1971). Here, one group of chromosomes resulting from first meiotic division undergoes normal second meiotic division whereas in the other group, there is restitution nucleus.

During the development of interspecific hybrids in groundnut (Arachis hypogaea), cytological-tetrad analysis of F, hybrids revealed the presence of dyads, triads and tetrads. Detailed cytological analysis revealed the restitution of second division. This meant that the first meiotic division was normal, but the cytokinesis in the second division was impaired, resulting in the formation of dyads and triads. Formation of 2n restitution nucleus or the 2n pollen was observed in crosses with wild species from section Arachis, to which cultivated groundnut belongs (Singh and Moss 1984). Formation of 2n pollen in F₁ hybrids from crosses A. hypogaea \times A. chiquitana (Figs. 1a and 1b), A. hypogaea × A. kretschmeri (section *Procumbentes*), and *A. duranensis* × *A. glabrata* (section Rhizomatosae) is a new finding. The 2n pollen from the cross A. hypogaea \times A. chiquitana and A. hypogaea $\times A$. kretschmeri were used to cross with A. hypogaea and develop tetraploid hybrids without going through the hexaploid route of backcross.

Use of 2n pollen in *Arachis* crossing program requires a large number of pollinations, but the process amply compensates by the development of tetraploids in one step, without the need to double the chromosome number

Cross	No. of dyads formed	No. of triads formed	No. of tetrads formed	Pollen stainability
A. hypogaea × A. hoehnei	48 (5) ¹	123 (13)	773 (82)	28
A. hypogaea \times A. cardenasii	53 (5)	205 (20)	759 (75)	26
A. hypogaea $\times A$. chiquitana	16(1)	150 (12)	1091 (87)	15
A. hypogaea $\times A$. kretschmeri	10 (2)	53 (12)	366 (85)	10
A. diogoi $\times A$. glabrata	15 (6)	23 (9)	209 (85)	30
A. duranensis $\times A$. glabrata	30 (32)	8 (9)	56 (60)	38
A. hypogaea $\times A$. glabrata	9 (3)	16 (5)	299 (92)	26

Table 1. Formation of dyads, triads and tetrads in interspecific derivatives of groundnut (<i>Arachis hypogaea</i>).	ecific derivatives of groundnut (<i>Arachis hypogaea</i>).
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of triploids and the laborious backcrossing program of the hexaploids to generate tetraploids. By this method it was possible to develop interspecific tetraploid derivatives from the crosses A. hypogaea $\times A$. chiquitana and A. hypogaea $\times A$. kretschmeri.

Dyads were observed as a result of restitution of both the groups of chromosomes at anaphase II. The number of dyads formed was low compared to the total number of pollen grains (Table 1), but the advantage of dyads is that they are fertile, which is evident from the acetocarmine stainability and in vivo pollen germination studies. Crosses using triploid pollen (*A. hypogaea* \times *A. cardenasii*) (Table 1) gave rise to a few pegs and pods, which is a further confirmation that some of the triploid

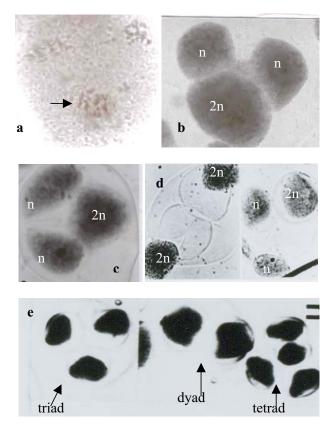


Figure 1. Formation of dyads and triads in *Arachis* interspecific hybrids: (a & b) Telophase with a normal separation of chromosomes and a restitution nucleus, leading to the formation of a triad in *A. hypogaea* \times *A. chiquitana* (Note: Arrow points towards restitution nucleus); (c) Triad formation in *A. hypogaea* \times *A. cardenasii;* (d) Dyads and triads in *A. duranensis* \times *A. glabrata*; and (e) Dyads, triads and tetrads in the hybrid *A. hypogaea* \times *A. glabrata.*

pollen are fertile. Reciprocal crosses using the triploid (*A. hypogaea* × *A. cardenasii*) as the female parent and *A. hypogaea* as the pollen donor, gave rise to 6% peg formation as a result of 500 pollinations, resulting in 7 pods. The resultant hybrids, obtained using *A. hypogaea* × *A. cardenasii* as the female parent and *A. hypogaea* as pollen donor, were tetraploids, which was confirmed by pollen diameter analysis. It is fairly simple to observe dyads and triads in tetrad analysis, which may not be the case with megasporogenesis, as the eggs are embedded deep in the ovular tissues.

Singh and Moss (1984) reported the formation of pegs and pods in triploid interspecific derivatives from the crosses A. hypogaea \times A. chacoense and A. hypogaea \times A. cardenasii, which were obtained as cuttings from the University of Reading, Reading, UK. Interestingly in Reading, the triploids were sterile, but some of the plants grown at the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, India were partially fertile with the formation of a few pegs and pods. Cytological analysis of triploids showed that there were meiotic irregularities and the formation of restitution nucleus. Second division restitution was observed in the interspecific hybrids which were developed at ICRISAT (2000-05) between A. diogoi and A. glabrata, A. hypogaea and A. hoehnei, A. duranensis and A. glabrata, and A. hypogaea and A. cardenasii (Table 1). The action of restitution nucleus was evident by the presence of dyads and triads in pollen tetrad analysis (Figs. 1c and 1d). Dyads and triads have been observed in the tetraploid cross A. hypogaea $\times A$. glabrata (Fig. 1e), which may not be of use in the improvement of A. hypogaea.

The report by Singh and Moss (1984) shows that environment may have a role to play in the formation of restitution nucleus in *Arachis* interspecific hybrids obtained as a result of crossing wild *Arachis* with cultivated groundnut. The results from our study show that 2n pollen can be effectively used to obtain tetraploids interspecific derivatives, without the use of colchicine to double the chromosome number of triploids and avoid the laborious hexaploid route to obtain tetraploids.

References

Christopher J. 1971. Asynapsis in *Paspalum* Linn. Nucleus 14:116–118.

Rosenberg O. 1927. Die semiheterotypische teilung und ihre bedeutung fur die entstehung verdoppelter chromozomenzahien. Hereditas 8:305–358.

Singh AK and **Moss JP.** 1984. Utilization of wild relatives in genetic improvement of *Arachis hypogaea* L. VI. Fertility in triploids: Cytological basis and breeding implications. Peanut Science 11:17–21.

Sosa RM and **Hernandez de Sosa.** 1971. Use of dihaploids in the breeding of *Solanum tuberosum* L. I. Cytological considerations. Hereditas 69:83–100.

Wagennar EB. 1968. Meiotic restitution and the origin of polyploidy. II. Prolonged duration of metaphase I as casual factor of restitution induction. Canadian Journal of Genetics and Cytology 10:844–852.

Arachis hoehnei, the Probable B Genome Donor of *Arachis hypogaea* Based on Crossability, Cytogenetical and Molecular Studies

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Cultivated groundnut (Arachis hypogaea) is made up of two genomes, A and B. It is presumed that polyploidization event between diploid A and B genome species gave rise to cultivated tetraploid groundnut some 3500 years ago (Singh and Simpson 1994). There is no ambiguity regarding A. duranensis as the A genome donor of A. hypogaea (Gregory and Gregory 1979, Singh 1988, Kochert et al. 1991, Paik-Ro et al. 1992, Stalker 1992). Different species from the B genome pool have been proposed as the B genome donor. According to Singh (1998), A. batizocoi is the B genome donor. Based on RFLP (restriction fragment length polymorphism) studies, Kochert et al. (1991) have suggested A. ipaensis as the B genome donor. According to Paik-Ro et al. (1992), A. batizocoi is not closely related to A. hypogaea and hence cannot be the B genome donor. Karyotype studies of Fernandez and Krapovickas (1994) support A. duranensis and A. ipaensis as the A and B genome donors of A. hypogaea.

We studied the crossability relationship between *A. hypogaea* and six B genome species. Cultivated groundnut was crossed with *A. hoehnei*, *A. benensis*, *A. valida*, *A. magna*, *A. batizocoi* and *A. ipaensis*. *Arachis hoehnei* when crossed with *A. hypogaea* set bold seeds without

the application of growth regulators. Majority of the seeds germinated in vitro and hybrid plants were obtained and a few (5%) mature seeds were obtained. Fertility in the hybrids ranged from 14 to 21%, whereas A. benensis, A. valida, A. magna and A. ipaensis set immature seeds, when crossed with A. hypogaea. The seeds were less than 3 mm in size. This indicated that the hybrid embryos aborted early. Embryo rescue technique was necessary to obtain hybrid plants if A. benensis, A. valida, A. magna and A. ipaensis were used as pollen donor. Arachis *batizocoi* set mature seeds with A. hypogaea, but pollen fertility was low (7%). Singh and Moss (1984) reported that in the crosses involving A. batizocoi and diploid A genome wild species from section Arachis, mean bivalents ranged from 3.2 to 6.9 with pollen fertility ranging between 3 and 7%. When A. hypogaea was crossed with A. batizocoi, the survival of the seedlings was poor.

Crosses were also carried out between *A. duranensis* and *A. hoehnei* (Fig. 1a). Large number of seeds (15%) was obtained. Cytogenetical study of the hybrid between *A. duranensis* and *A. hoehnei* showed 10 bivalent formation in 30% of the pollen mother cells analyzed (Fig. 1b). Amongst the bivalents, 4–6 were ring bivalents. The formation of large number of bivalents and in ring formation shows that there is homeology between the genomes of *A. duranensis* and *A. hoehnei*. For a hybrid between A and B genomes to survive in nature, greater degree of homeology between the genomes would be a contributory factor and would play a major role in the perpetuation of the hybrid. Such a hybrid could have doubled its chromosome number to give rise to the amphidiploid groundnut.

Simple sequence repeat (SSR) analysis of A. duranensis, A. hoehnei and the hybrid between A. duranensis and A. hoehnei was carried out. The SSR 4F07 profile of A. duranensis was different from that of A. hoehnei. The hybrid had the DNA profile with bands from both the parents. The interesting feature of the hybrid DNA profile was that it resembled the DNA profile of A. hypogaea with some differences (Fig. 1c). This shows that the hybrid, which has the genomes of both A. duranensis and A. hoehnei, has close resemblance to the genome of A. hypogaea. The difference between the hybrid A. duranensis $\times A$. hoehnei and A. hypogaea may be due to ploidy difference and the synthesis of A. hypogaea which took place some 3500 years ago.

Based on crossability between *A. duranensis* and *A. hoehnei*, cytogenetical data and molecular analysis of the hybrid between *A. duranensis* and *A. hoehnei*, we propose *A. hoehnei* as the probable B genome donor of cultivated groundnut.

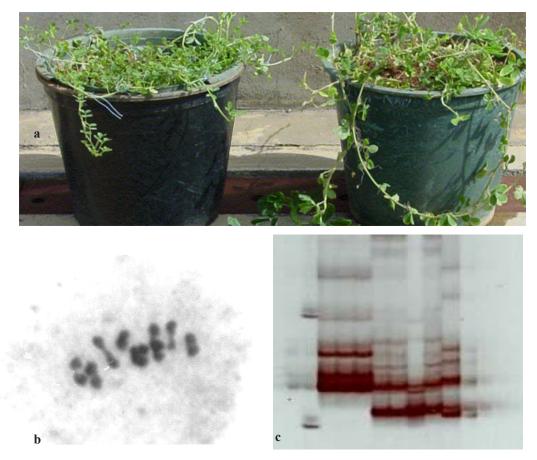


Figure 1. Arachis hoehnei as the B genome donor of cultivated groundnut *A. hypogaea*: (a) Arachis duranensis (left) and *A. hoehnei* (right); (b) metaphase plate of *A. duranensis* × *A. hoehnei* (note the presence of 10 bivalents); and (c) SSR marker 4F07 profile: Lane 1- 100 base pair ladder, Lanes 2–4 - *A. duranensis*, Lanes 5, 6 & 8 - hybrid between *A. duranensis* and *A. hoehnei*, Lane 7 - *A. hoehnei*, Lane 9 - *A. hypogaea*.

References

Fernandez A and Krapovickas A. 1994. Chromosomas y evolution en *Arachis (Leguminosae)*. Bonplandia 8:187–220.

Gregory MP and **Gregory WC.** 1979. Exotic gremplasm of *Arachis* L. interspecific hybrids. Journal of Heredity 70:185–193.

Kochert G, Halward T, Branch WD and Simpson CE. 1991. RFLP variability in peanut (*Arachis hypogaea* L.) cultivars and wild species. Theoretical and Applied Genetics 81:565–570.

Paik-Ro OG, Smith RL and **Knauft DW.** 1992. Restriction fragment polymorphism evaluation of six peanut varieties within the *Arachis* section. Theoretical and Applied Genetics 84:201–205.

Singh AK. 1988. Putative genome donors of *Arachis hypogaea* (Fabaceae), evidence from crosses with synthetic amphidiploids. Plant Systematics and Evolution 160:143–151.

Singh AK and **Moss JP.** 1984. Utilization of wild relatives in genetic improvement of *Arachis hypogaea* L. 5. Genome analysis in section Arachis and its implication in gene transfer. Theoretical and Applied Genetics 68:350–364.

Singh AK and **Simpson CE.** 1994. Biosystematics and genetic resources. Pages 96–137 *in* The groundnut crop: A scientific basis for improvement (Smartt J, ed.). London, UK: Chapman & Hall.

Stalker ST. 1992. Utilizing *Arachis* germplasm resources. Pages 281–295 *in* Groundnut – a global perspective: proceedings of an international workshop, 25–29 Nov 1991, ICRISAT Center, India (Nigam SN, ed.). Patancheru 502 324, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics.

Differences in Pod Characters Among Groundnut Cultivar L7-1 and its Chemical Mutants

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Early studies on mutation breeding in groundnut (*Arachis hypogaea*) were conducted in USA by Gregory (1955) using X-rays, in Africa by Tuchlenski (1958) using γ -rays, in Israel by Ashri and Goldin (1965) and Ashri (1970) using diethyl sulfate (DES), and in India by Patil (1968) and Lin (1960) in China by using X-rays. Most of the recent efforts in groundnut mutation breeding have been made through irradiation in India and China (Branch 2002).

Until now, over 30 groundnut varieties have been released worldwide with the help of induced mutations (Knauft and Ozias-Akins 1995). In China, for example, Luhua 12, a cultivar of *Hsuji*TM (Spanish) market type, was developed after treatment of hybrid pegs with EMS (ethane methyl sulfonate). Previous attempts at chemicalinduced mutation in groundnut reported alterations in external characters, but did not mention if there were changes in internal quality traits (Wan 2005). Considering the narrow gene base of the cultivated groundnut, the potential of chemical-induced mutation in groundnut breeding deserves further evaluation. This is extremely important for Shandong province, the leading groundnut producer of China, where groundnut breeding for yield and quality has remained stagnant for more than 8 years (Wang et al. 2006).

We have obtained large-podded and small-podded groundnut mutants following sodium azide (NaN_3) treatment of groundnut cultivar L7-1 (Wang et al. 2002). The pod size and seed size of these mutants were stable based on the observations over 6 years. The objective of this study was to examine the differences, if any, in both external and internal quality traits among the large- and small-podded mutants and L7-1.

The cultivar L7-1 is a hybrid derivative of *A. hypogaea* cv Shilihong and *A. glabrata* with bold, pink, elliptical oblong seeds desirable for export. L7-1 and its two NaN₃-induced mutant lines with either large or small pods were planted in adjacent rows for sampling.

The seed was sown under polythene (with Acetochlor) mulch, with a population of 124,995 plants ha⁻¹ at Qingdao Agricultural Academy Experimental Region, Shandong

Table 1. Difference in external	and internal quality traits
among groundnut cultivar L7-1	and its mutants.

Trait	Type ¹	Mean	2	SE±
Pod length	LP	52.31	А	0.57
	SP	33.40	В	0.31
	L7-1	40.77	С	0.38
Pod width	LP	18.66	А	0.23
	SP	13.80	В	0.11
	L7-1	17.32	С	0.20
Pod thickness	LP	17.33	А	0.18
	SP	12.61	В	0.10
	L7-1	15.46	С	0.16
Pod mass	LP	3.97	А	0.09
	SP	1.86	В	0.04
	L7-1	3.11	С	0.06
Apical seed length	LP	25.46	А	0.42
	SP	17.04	В	0.18
	L7-1	21.69	С	0.22
Apical seed width	LP	8.96	a	0.11
	SP	8.61	ab	0.09
	L7-1	8.31	bc	0.23
Apical seed thickness	LP	12.02	A	0.21
	SP		B	0.12
A ¹ 1 1	L7-1	11.16	C	0.14
Apical seed mass	LP	1.46	A	0.05
	SP	0.71	B	0.02
Decel and low eth	L7-1	1.13	C	0.03
Basal seed length	LP SP	23.77	A B	0.43
	SP L7-1	16.08 20.63	С	0.16 0.25
Basal seed width	L7-1 LP	9.73	A	0.23
Dasal seeu wiutii	SP	9.73	B	0.13
	L7-1	10.21	C	0.09
Basal seed thickness	LP	12.97	A	0.09
Dasai seeu ullekliess	SP	9.44	B	0.09
	L7-1	11.77	C	0.10
Basal seed mass	LP	1.59	A	0.04
Busur seed muss	SP	0.72	В	0.04
	L7-1	1.28	C	0.02
Protein (%)	LP	27.40	ab	0.00
	SP	27.36	a	0.00
	L7-1	27.81	b	0.00
Oil (%)	LP	42.80	AC	0.00
	SP		В	0.00
	L7-1		BC	0.00
Oleic acid (O) (%)	LP	48.29	А	0.01
	SP	36.21	В	0.02
	L7-1	45.33	А	0.01
Linoleic acid (L) (%)	LP	34.74	А	0.01
	SP	44.62	В	0.01
	L7-1	36.71	А	0.01
Palmitic acid (%)	LP	11.83	А	0.00
	SP	14.35	В	0.00
	L7-1	12.06	А	0.00
O/L ratio	LP	1.39	А	0.05
	SP	0.82	В	0.06
	L7-1	1.24	А	0.03

1. LP = Large-podded mutant; SP = Small-podded mutant.

2. Means of specific trait within the same column followed by the same letter are not significantly different, for lower case letters at P = 0.05 and for upper case letters at P = 0.01.

on 1 May 2005. Weeds were pulled out by hand, and pesticide was sprayed a week before wheat (Triticum aestivum) harvest (Wan 2003). During the whole plant growth period, no irrigation was needed due to plenty of rainfall and adequate soil moisture. Groundnut was harvested on 10 September 2005. After drying, 48 representative pods from each entry were randomly selected for measurement of length, width and thickness of pod, apical seed, and basal seed using vernier calipers. The protein, oil, and oleic, linoleic and palmitic acids contents (percentage of total fatty acids) were determined in 18–20 bulked seed samples (5 groups for each entry) by near infrared reflectance spectroscopy (NIRS) (Yu et al. 2003a, 2003b). Each sample was measured 4 times, and the average was used in subsequent statistical analysis. For external quality traits, where equality of error variances was not assumed according to Levene's test, robust tests of equality of means by Welch and Brown-Forsythe methods and multiple comparisons of differences by Games-Howell test were conducted (Quinn and Keough 2002). For internal quality characters, multivariate analysis of variance (MANOVA) using Pillai's trace/Wilks' lambda, and multiple comparisons of differences by Tukey's Honest Significant Difference (HSD) test were exploited (Quinn and Keough 2002).

Significant differences between the mutants and the control for pod and seed length and mass were detected by visual inspection (Fig. 1). Robust tests of equality of means of external quality traits in the mutants and L7-1 indicated that length, thickness and mass of pod/seed of the 3 types differed significantly (P = 0.01), and apical seed

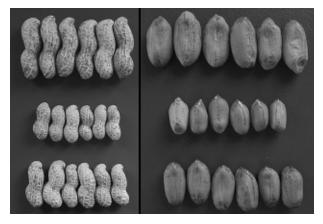


Figure 1. Pods (left) and seeds (right) of groundnut cultivar L7-1 (bottom), and its derived large-podded (top) and small-podded (middle) mutants.

width differed at 0.05 (P=0.05) level. Multiple comparisons further showed that the large-podded and the smallpodded mutants exhibited a drastic change in pod and seed length, thickness and mass as compared with the control L7-1, whereas the apical seed width of the smallpodded mutants was not significantly different from that of L7-1 (Table 1).

MANOVA of internal quality traits in the mutants and L7-1 revealed that internal quality characters inclusive of protein, oil, and oleic, linoleic and palmitic acids contents differed significantly (P=0.01) among the 3 types. Multiple comparisons showed that for protein content, significant difference (P = 0.05) existed only between the smallpodded mutant and L7-1; for oil content, significant difference (P = 0.01) existed only between the largepodded mutant and the small-podded mutant. In contrast to the large-podded mutant, the small-podded mutant showed a reverse tendency in fatty acid composition. The oleic acid (O) of the small-podded mutant was lower, and linoleic acid (L) and palmitic acid contents were higher than those of L7-1, and the differences were significant at P = 0.01. The O/L ratio of the small-podded mutant was much lower than the large-podded mutant and L7-1 (significant at P = 0.01). The O/L ratio of the large-podded mutant, however, was not statistically different from L7-1 (Table 1).

The change in internal quality in the "wrong" direction in this study does not necessarily mean the same case as in additional mutants. Conversely, alterations in traits of the two mutant lines from this study indicated the necessity for large-scale evaluation of the groundnut mutant bank of EMS, DES and NaN, for broad internal quality variation range. In conclusion, this study demonstrated the possible utility of chemical-induced mutation in groundnut germplasm enhancement not only for external quality traits but for internal quality traits as well. Mutants with much larger and smaller seed size, even with altered fatty acids composition, may be produced following chemical mutagen treatment. This is especially important for the cultivated groundnut, whose genetic base is narrow, as a result of genetic bottleneck where ploidy difference and pre-/post-fertilization barriers make gene exchange difficult with its wild relatives; this situation is further aggravated in cultivars by the limited number of core parents exploited by breeders. Chemical induced mutation techniques, when used in TILLING (Targeting Induced Local Lesions IN Genomes), may facilitate creation and identification of mutations in DNA regions of interest, and when combined with NIRS, may speed up the process of groundnut quality improvement.

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References

Ashri A. 1970. A dominant mutation with variable penetrance and expressivity induced by diethyl sulfate in peanuts, *Arachis hypogaea* L. Mutation Research 9:473–480.

Ashri A and Goldin E. 1965. The mutagenic activity of diethyl sulfate in peanuts. Radiation Botany 5:431–441.

Branch WD. 2002. Variability among advanced gammairradiation induced large-seeded mutant breeding lines in the 'Georgia Browne' peanut cultivar. Plant Breeding 121:275–277.

Gregory WC. 1955. X-ray breeding of peanuts (*Arachis hypogaea* L.). Agronomy Journal 47:396–399.

Knauft DA and **Ozias-Akins P.** 1995. Recent methodologies for germplasm enhancement and breeding. Pages 54–94 *in* Advances in peanut sciences (Pattee HE and Stalker HT, eds.). Stillwater, Oklahoma, USA: American Peanut Research and Education Society Inc.

Lin H. 1960. The influences of thermal neutrons and X-ray irradiation on peanut in the first generation. Journal of Agricultural Association of China New Ser. No. 32:27–37.

Patil SH. 1968. Cytogenetics of X-ray induced aneuploids in *Arachis hypogaea* L. Canadian Journal of Genetics and Cytology 10:545–550.

Quinn GP and **Keough MJ.** 2002. Experimental design and data analysis for biologists. UK: Cambridge University Press. 562 pp.

Tuchlenski H. 1958. Groundnut breeding with special reference to production of mutations. Proceedings of the First Congress of South Africa Genetic Society 07:107–109.

Wan SB. 2003. Peanut cultivation science. Shanghai, China: Shanghai Science and Technology Press. pp. 340–429.

Wan SB. 2005. Peanut quality science. Beijing, China: China Agricultural Science and Technology Press. pp. 156–159.

Wang CB, Wan SB, Zheng YP, Cheng B, Wu ZF and **Gao XH.** 2006. The present major problem, cause and development measure of peanut produce in Shandong province. Journal of Peanut Science 35(1):25–28.

Wang CT, Yang XD, Chen DX, Zhang JC, Xu JZ and **Yang WQ.** 2002. Production of extra large-podded and small-podded peanut mutants following chemical mutagen treatment. Journal of Peanut Science 31(4):5–8.

Yu SL, Zhu YJ, Min P, Liu H and **Cao YL.** 2003a. NIRS to determine protein and oil contents in peanut kernels. Journal of Peanut Science 32 (Supplement):138–143.

Yu SL, Zhu YJ, Min P, Liu H, Cao YL, Wang CT, Zhang CS, Liu X and Zhou XQ. 2003b. NIRS to determine major fatty acids contents in peanut kernels. Pages 344–349 *in* High quality and high output peanuts: Principal and techniques for production (Wan SB, ed.). Beijing, China: China Agricultural Science and Technology Press.

Seed Releases

A High-yielding Drought-tolerant Groundnut Variety Abhaya

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Groundnut (Arachis hypogaea), an important oilseed and food crop of Andhra Pradesh, India is grown largely as a rainfed crop during the rainy season. Drought is the major abiotic stress affecting yield and quality of rainfed groundnut in the state. Yield losses due to drought are highly variable depending on its timing, intensity and duration coupled with other location specific environmental factors such as irradiance and temperature (Nigam et al. 2001). Thus the groundnut productivity in rainy season in the state ranges between 500 kg ha⁻¹ and 1200 kg ha⁻¹ (Reddy et al. 2003). To stabilize yield under rainfed conditions, it is necessary to develop varieties that tolerate moisture stress at different stages of crop growth. To achieve this objective, research was initiated to identify donor parents for drought tolerance traits such as low specific leaf area (SLA), high SPAD chlorophyll meter reading (SCMR) and high harvest index (HI) that confer advantage under drought conditions. Through principal component analysis, ICGV 86031, CSMG 84-1, ICGS 76 and TAG 24 were identified as genotypes with most of the useful traits for drought tolerance (Nageswara Rao and Wright 2003). Hybridization was effected in 1998 involving these genotypes as male parents. From K 134 \times TAG 24 cross, TPT 25 was developed through modified pedigree method with focus on drought tolerance traits in segregating generations. It belongs to supspecies *fastigiata* and variety *vulgaris*. TPT 25 is a short-statured, drought-tolerant, high-yielding Spanish bunch groundnut variety (Fig. 1). Its special attributes are: plant height 27–34 cm, sequential branching pattern, short internodes, narrow dark green leaflets, 4–6 primary branches, decumbent plant type, slender pods without beak, higher frequency of three-seeded pods, thin shell, higher shelling outturn, and high oil content of 52% (Table 1). It matures 105–110 days in the rainy season.

TPT 25 was tested in yield trials at Regional Agricultural Research Station (RARS), Tirupati, in different All India Coordinated Research Project (AICRP) centers and on farmers' holdings in Chittoor, Kadapa and Anantapur districts extensively (Table 2). It was also tested in state



Figure 1. A mature plant of groundnut variety TPT 25.

Variety	Plant height	Time to maturity		SLA	RWC	Shelling outturn	SMS	Oil content
	(cm)	(days)	SCMR	(cm ² g ⁻¹)	(%)	(%)	(%)	(%)
TPT 25	30.7	105.6	42.5	167	83.3	72.2	84.5	52.3
Narayani (check)	52.5	90.0	40.9	217	77.9	70.6	82.0	48.3

SCMR = SPAD chlorophyll meter reading; SLA = Specific leaf area; RWC = Relative water content in leaf; SMS = Sound mature seed.

							AICRP trials	als	4	Minikit trials	als		
		Sta	Station trials			(ea	(early season stress)	stress)	(mi	(mid-season stress)	stress)	Σ	Mean
Variety	2003	2004	Mean	.004 Mean 2004-05 2005	2005	2004	2005	Mean	2004	2005	2005 Mean	2004 2005	2005
TPT 25	2741	1945	2343	3756	2303	1235	1203	1219	1134	1545	1545 1340	1191	890
Narayani/TMV 2/Local (check) ¹	2257	1302	1780	3444	1787	1065	932	666	860	1353	1107	810	710
Increase over control (%)	17	49	29	10	29	16	29	22	32	14	21	47	14
CD at 50%	580	204						251			261		
CV (%)	15	21											

multilocational varietal trials at different research stations of Acharya NG Ranga Agricultural University (ANGRAU) for two years covering different agroclimatic situations of Andhra Pradesh. It outperformed the existing varieties JL 24 and TMV 2 at many locations with additional attributes of tolerance to drought and late leaf spot. Based on these results, the Andhra Pradesh State Varietal Release Committee released TPT 25 as Abhaya in June 2006 for general cultivation in the state. It is recommended for both rainy and postrainy season cultivation throughout Andhra Pradesh. Due to its compact nature, TPT 25 is also suitable for high rainfall areas where excess vegetative growth in the existing varieties leads to drastic reduction in yield and the quality of the produce during the rainy season.

In trials at RARS, Tirupati during rainy season 2003 and 2004, TPT 25 produced an average pod yield of 2343 kg ha⁻¹ that was 29% higher than Narayani and 13% higher than Vemana, the two recently released varieties in the state. Its seed yield was 1608 kg ha-1, which was 34% higher than Narayani and 15% higher than Vemana. It was tested at AICRP centers identified for their drought pattern - early season drought stress (Tirupati, Anantapur and Vriddhachalam) and mid-season drought stress (Jalgaon, Chintamani and Raichur). The average pod yield of TPT 25 under early season drought stress was 1219 kg ha⁻¹ (mean of rainy season 2004 and 2005) with an overall increase of 22% over the check variety TMV 2. In the mid-season drought stress situation, the average pod yield of TPT 25 was 1340 kg ha-1 which was 21% higher than the check variety TMV 2 (Table 2). In endof-season drought stress situation, the pod yield of TPT 25 was limited to that of check variety (data not given).

References

Nageswara Rao RC and **Wright GC.** 2003. The physiological basis for selection of peanut genotypes as parents in breeding for improved drought resistance. Pages 10–14 *in* Breeding for drought-resistant peanuts. ACIAR Proceedings No. 112. Canberra, Australia: Australian Centre for International Agricultural Research.

Nigam SN, Nageswara Rao RC and Wright GC. 2001. Breeding for increased water-use efficiency in groundnut. Pages 1–2 *in* Abstracts, New Millenium International Groundnut Workshop, 4–7 September 2001, Shandong, China. Qingdao, China: Shandong Peanut Research Institute.

Reddy PV, Asalatha M, Vasanthi RP, Sujatha D and **Jayalakshmi V.** 2003. Evaluation of trait-based and empirical selections for drought resistance at Tirupati, Andhra Pradesh, India. Pages 37–42 *in* Breeding for drought-resistant peanuts. ACIAR Proceedings No. 112. Canberra, Australia: Australian Centre for International Agricultural Research.

New High-yielding Groundnut Varieties GG 8 and GG 16

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Groundnut (*Arachis hypogaea*) is the major oilseed crop of India. It is grown in 11 states in the country in an area of 7.6 million ha with a production of 7.8 million t of pods per annum. The average productivity of groundnut in India is about 1000 kg ha⁻¹ and is stagnating for the last several years. To overcome such stagnation in production and productivity of groundnut, efforts on varietal improvement with emphasis on high yield and resistance/ tolerance to biotic and abiotic stresses, and development of low-cost crop management practices are needed.

In this direction, the Main Oilseeds Research Station, Junagadh Agricultural University (JAU), Junagadh, Gujarat, India has developed two new high-yielding groundnut varieties GG 8 and GG 16 for Zone III and Zone V, respectively. Gujarat Groundnut 8 (GG 8), a Spanish bunch variety (*A. hypogaea* subsp *fastigiata* var *vulgaris*) and Gujarat Groundnut 16 (GG 16), a Virginia runner variety (*A. hypogaea* subsp *hypogaea* var *hypogaea*), were developed from the crosses $27-5-1 \times$ JL 24 and JSP $14 \times$ JSSP 4, respectively following pedigree method of selection. After preliminary evaluation at the Main Oilseeds Research Station as J 53 (GG 8) and JSP 39 (GG 16), they were proposed in rainy season 2002 for evaluation in the All India Coordinated Varietal Trials, IVT-SB-I and IVT-VG-I, respectively. Then they were tested in IVT-II and AVT during rainy season 2002–04.

For J 53 (GG 8) the trials were conducted at 11 locations in Akola, Khargone, Jalgaon and Raipur centers, while for JSP 39 at 27 locations in Aliyarnagar, Vriddhachalam, Jagatial, Kadiri, Raichur, Dharwad, Chintamani, Kayumkulam, Digraj and Latur centers. In three years of testing, J 53 produced a mean dry pod yield of 1716 kg ha⁻¹ as compared to 1493 kg ha⁻¹ of JL 24 (national check) and 1608 kg ha⁻¹ of TAG 24 (zonal check). The yield

Table 1. Comparative dry pod yield (kg ha⁻¹) and seed yield (kg ha⁻¹) of groundnut variety J 53 (GG 8) in the All India Coordinated Varietal Trials (AICVTs) conducted in Zone III, India during rainy season 2002–04.

		J 53	J 53 (GG 8)		nal check)	TAG 24 (zo	onal check)
Year	Number of locations	Dry pod yield	Seed yield	Dry pod yield	Seed yield	Dry pod yield	Seed yield
2002	4	1205	836	1269	865	1888	1244
2003	3	2567	1812	2112	1403	1859	1172
2004	4	1375	930	1108	760	1076	725
Mean		1716	1193	1493	1009	1608	1047
Increase over check (%)				14.9	18.2	6.7	13.9

Table 2. Reaction of groundnut variety J 53 (GG 8) to major diseases and pests.

		Mean values (2002–04)			
Diseases/insect pests	J 53 (GG 8)	JL 24 (national check)	TAG 24 (zonal check)		
Late leaf spot ¹	6.1	5.3	5.2		
Rust ¹	4.7	3.9	1.5		
Stem rot incidence (%)	6.3	5.2	2.6		
Bud necrosis intensity (%)	1.5	3.3	NA^2		
Collar rot intensity (%)	0.5	6.0	NA		
Root rot intensity (%)	2.7	4.3	4.4		
Thrips damage (%)	30.3	41.4	41.0		
Jassids demage (%)	23.3	23.3	NA		
Prodenia damage (%)	2.9	4.3	2.0		
No. of leaf miners plant ⁻¹	0.5	0.4	NA		

1. Scored on a 0–9 rating scale, where 0 = no disease, $1 = \le 1\%$, 3 = 1-5%, 5 = 6-20%, 7 = 21-50%, $9 = \ge 51\%$ disease damage.

2. NA = Data not available.

Table 3. Comparative dry pod yield (kg ha⁻¹) and seed yield (kg ha⁻¹) of JSP 39 (GG 16) in All India Coordinated Varietal Trials (AICVTs) conducted in Zone V, India during rainy season 2002–04.

		JSP 39 (GG 16)		M 335 (national check)		ICGV 86325 (zonal check)	
Year	Number of locations	Dry pod yield	Seed yield	Dry pod yield	Seed yield	Dry pod yield	Seed yield
2002	9	2079	1526	867	616	1466	1121
2003	8	1978	1227	1447	1031	1383	1113
2004	10	1979	1262	1645	1122	1472	1069
Mean		1992	1338	1373	923	1459	1024
Increase over check (%)				45.1	45.0	27.0	18.6

Diseases/insect pests	Mean values (2002–04)				
	JSP 39 (GG 16)	M 335 (national check)	ICGV 86325 (zonal check)		
Late leaf spot ¹	4.6	3.8	3.9		
Rust ¹	4.1	3.1	2.7		
Stem rot intensity (%)	3.9	4.2	5.7		
Bud necrosis intensity (%)	1.8	3.1	4.8		
Collar rot intensity (%)	3.0	2.3	4.1		
Root rot intensity (%)	4.8	8.6	15.0		
Thrips damage (%)	30.2	32.0	19.9		
Jassids demage (%)	19.2	16.7	29.8		
Prodenia damage (%)	3.0	9.0	4.7		
No. of leaf miners plant ⁻¹	1.8	0.1	1.7		

1. Scored on a 0–9 rating scale, where 0 = no disease, $1 \le 1\%$, 3 = 1-5%, 5 = 6-20%, 7 = 21-50%, $9 = \ge 51\%$ disease damage.

advantage over national check was 14.9% and over zonal check 6.7%. The seed yield of J 53 (1193 kg ha⁻¹) was 18.2% higher than JL 24 (1009 kg ha⁻¹) and 13.9% higher than TAG 24 (1047 kg ha⁻¹) (Table 1). This variety also showed higher shelling outturn (69%) than JL 24 (68%) and TAG 24 (66%). It exhibited slightly higher incidence of late leaf spot, rust and stem rot diseases but lower incidence of bud necrosis, collar rot and root rot diseases as compared to both the checks. This variety was similar to the check varieties in jassids and leaf miner reactions. Thrips and prodenia damage was low on J 53 compared to JL 24 (Table 2).

GG 8 (J 53) is erect in growth habit and takes 106 days to mature. The leaves are medium green and oblong in shape. Pods are two-seeded with slight reticulation and constriction, and pod beak is absent. Seeds are medium, round and rose in color. The oil content in GG 8 (46%) is less than JL 24 (47%) and TAG 24 (49%).

During the three-year testing, JSP 39 (GG 16) recorded dry pod yield of 1992 kg ha⁻¹ as compared to 1373 kg ha⁻¹ of M 335 (national check) and 1459 kg ha⁻¹ of ICGV 86325 (zonal check). The yield advantage in GG 16 was 45.1% over M 335 and 27.0% over ICGV 86325. The seed yield of this variety was 1338 kg ha⁻¹, which was 45.0% and 18.6% higher than M 335 (923 kg ha⁻¹) and ICGV 86325 (1024 kg ha⁻¹), respectively (Table 3).

GG 16 (JSP 39) showed similar reaction to rust, late leaf spot, stem rot and collar rot diseases as that of check varieties, while it was superior in bud necrosis and root rot diseases reaction. Leaf damage by thrips and jassids was similar in GG 16 and the check varieties under field condition (Table 4). GG 16 is a spreading type with profuse branching and takes 119 days to mature. The leaves are green and elliptical. Pods are big and two-seeded with moderate constriction and reticulation, and have slight beak. Seeds are medium, elongated and rose in color. The 100-seed mass is 43 g and the shelling outturn is 63%. The oil content in this genotype is 46%.

GG 8 and GG 16 were identified for Zone III (northern Maharashtra and Madhya Pradesh) and Zone V (Tamil Nadu, Andhra Pradesh, Karnataka, Kerala and Southern Maharashtra), respectively by the Variety Identification Committee meeting of the All India Coordinated Research Project (AICRP) on Groundnut held at the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, India during 2–4 October 2005.

Performance of Groundnut Cultivar ICGV 93468 During Summer Season in Uttar Pradesh, India

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Groundnut (Arachis hypogaea) is an important oilseed crop of Uttar Pradesh, India primarily grown during the rainy season. In early 1980s, groundnut was grown in Uttar Pradesh on 0.3 million ha during the rainy season with a production of 0.19 million t. Since then, both area and production have shown a steady decline due to biotic and abiotic stresses. During 1997-98, groundnut area declined from 0.3 million ha to 0.13 million ha and production from 0.19 million t to 0.12 million t. Efforts to arrest this decline in area and production did not succeed due to various administrative and economic reasons. A strong need was felt to develop a suitable technology to make groundnut cultivation more profitable in Uttar Pradesh. The main function of the National Agricultural Research Project (NARP), Mainpuri, Uttar Pradesh was to lead research on groundnut. Therefore, scientists of NARP deliberated on this important issue with SN Nigam at the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Andhra Pradesh, India and planned to introduce summer cultivation of groundnut in the state targeting the area vacant in summer season after harvesting potato (Solanum tuberosum)

Table 1. Performance of ICGV 93468 in comparison to the check varieties under state varietal trials conducted at different RATDSs of Uttar Pradesh, India during summer season 2005.

Genotype	Hardoi	Mathura	Bareilly ¹	Average
ICGV 93468	2.91	2.46	1.13	2.17
ICGS 44	2.64	2.26	0.89	1.93
ICGS 1	2.55	1.84	0.90	1.77
ICGV 86590	2.02	1.72	1.05	1.60
G 201 (local check)	1.30	1.75	0.74	1.26
SEm±	0.11	0.05	0.06	
CD at 5%	0.36	0.17	0.19	
CV (%)	10.41	12.80	13.43	

1. The crop was sown one-month late on 4 April 2005 at Belva farm of RATDS, Bareilly.

(0.38 million ha), mustard (*Brassica* sp) (0.53 million ha) and field pea (*Pisum sativum*) (0.23 million ha). The seed of different genotypes supplied by ICRISAT was tested during summer season of 1998 at Mainpuri. Among the ICRISAT genotypes ICGV 93468 performed well during summer season. Due to high yield potential, low incidence of insect pests and diseases, better survival under water stress condition, thermo-tolerance and early maturity (90–95 days), ICGV 93468 was considered to be most suitable for the groundnut farmers of Uttar Pradesh.

In 2001, the farmers of Mainpuri district followed for summer season groundnut cultivation with genotype ICGV 93468. Summer season groundnut cultivation in traditional and non-traditional areas of Mainpuri, Firozabad, Etawah, Auraiya, Kanpur, Kannauj, Farrukhabad, Aligarh, Hathras, Etah, Unnao, Hardoi, Fatehpur and Shahjahanpur districts slowly spread on an area of about 27,500 ha. The Department of Agriculture, Lucknow later on organized a meeting of the officers of State Agriculture Department and scientists of different agricultural universities of the state and decided to evaluate the variety ICGV 93468 further at Regional Agriculture Testing and Demonstration Stations (RATDSs) located in the different regions of Uttar Pradesh.

A varietal trial with improved ICRISAT varieties and a local check G 201 (Kaushal) was laid out during summer season of 2005 at RATDSs, Hardoi, Mathura (at Raya) and Bareilly (at Belva). The crop was sown on 9 March 2005 at Hardoi, 19 March 2005 at Mathura and 4 April 2005 at Bareilly. The crop was harvested on 21 June 2005 at Hardoi, 5 July 2005 at Mathura and 30 July 2005 at Bareilly. Sowing was done in rows 30 cm apart with 10 cm plant spacing. Recommended dose of 20 kg nitrogen ha⁻¹ + 30 kg P_2O_5 ha⁻¹ + 45 kg K₂O ha⁻¹ was applied at the time of planting of groundnut seed. Gypsum was applied at 300 kg ha⁻¹ with 50% quantity applied at sowing and the remaining 50% top dressed between

Table 2. Maturity duration of different groundnut varieties at different RATDSs of Uttar Pradesh, India during summer season 2005.

	Time to maturity (days)				
Genotype	Hardoi	Mathura	Bareilly	Average	
ICGV 93468	90	103	99	97	
ICGS 44	97	105	101	101	
ICGS 1	97	106	102	102	
ICGV 86590	95	107	102	101	
G 201 (local check)	93	108	103	101	



Figure 1. A bumper crop of groundnut variety ICGV 93468 (Avtar) in Uttar Pradesh, India.

flowering and pegging stage to ensure supply of calcium and sulfur to developing pods.

The genotype ICGV 93468 gave higher average pod yield (2.17 t ha⁻¹) compared to ICGS 1, ICGS 44, ICGV 86590 and G 201 (local check) (Table 1). ICGV 93468 gave 71.15% higher pod yield than local check G 201. The maximum yield of ICGV 93468 was 2.91 t ha⁻¹, harvested in Central Plain Zone of Uttar Pradesh at RATDS, Hardoi closely followed by 2.46 t ha⁻¹ in South Western Semi Arid Zone at RATDS, Mathura. The minimum yield obtained was 1.13 t ha⁻¹ in Middle Western Plain

Zone at RATDS, Bareilly. The yield variation in ICGV 93468 was not due to agroclimatic zones of Uttar Pradesh but due to time of sowing. In Hardoi and Mathura, sowing was done under recommended time while at Bareilly it was planted one-month late.

The variety ICGV 93468 (Avtar) matured at 97 days after planting (Fig. 1; Table 2). This maturity time was found conducive to the production of groundnut during summer season where the crop with longer duration is usually caught by rains at maturity/harvest.

Release of Groundnut Variety Huayu 23 in Shandong Province in China

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The groundnut (*Arachis hypogaea*) variety Huayu 23 is derived from a cross of two advanced breeding lines, ICGS 37 and R1(8124-19-1). ICGS 37, developed at the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, is a released cultivar in India; R1(8124-19-1) is a breeding line developed at Shandong Peanut Research Institute, China. Huayu 23 was released by the Shandong Crop Variety Approval Committee in 2004 for cultivation in Shandong province in China.

In Shandong provincial trials, Huayu 23 ranked first during 2002/03. The pod yield averaged 4.69 t ha⁻¹, 13.5% more than Luhua 12 (control) over 22 locations, and the seed yield averaged 3.51 t ha⁻¹, 16% more than the control. In national test, Huayu 23 produced 4.11 t ha⁻¹ pod yield, 26.71% more than Luhua 12 (control).

Huayu 23 matures in 125 days in the spring season. It has an erect growth habit, sequential flowering and dark green leaves. The main stem height of the plant is 37 cm, and the average length of branches is 43 cm. Huayu 23 has 9 primary branches. The pod has moderate to prominent reticulation with slight to moderate pod beak (Fig. 1). Pod constriction is medium. Seed coat is pale red. One-seeded and two-seeded pods account for 10% and 72% of all pods, respectively. The 100-pod mass is



Figure 1. Mature plants, pods and seeds of groundnut variety Huayu 23 in China.

154 g and the 100-seed mass is 64 g, with a shelling outturn of 75%. The seed contains 53.1% oil and 22.9% protein. The oleic acid/linoleic acid ratio is 1.55, making Huayu 23 a breakthrough in quality breeding for export of small-seeded groundnut after Luhua 15 in China (Table 1). It has good resistance to cercospora and phaeoisariopsis leaf spots.

Huayu 23 grows well on sandy soil with good drainage. The seed rate should be around 165,000 hills ha⁻¹ with two seeds in each hill for spring sowing.

Table 1. Seed quality traits of groundnut varieties Huayu 23 and Luhua 15 in China.					
Variety	Protein	Oil	Oleic acid (O)	Linoleic acid (L)	O/L
	(%)	(%)	(%)	(%)	ratio
Huayu 23	22.9	53.1	49.3	31.9	1.55
Luhua 15	28.6	50.9	44.7	34.1	1.31

Registration of Foliar Disease Resistant and High-yielding Groundnut Varieties **ICGV 92099 and ICGV 90084**

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Groundnut (Arachis hypogaea) is an important crop for small-scale farmers in Ghana. Although the crop is produced in all agro-ecologies of the country, the bulk of production occurs in the northern region, which spans the Guinea and Sudan savannah ecologies lying within 8-11° N. The crop is produced mainly for oil, although a significant proportion of the seed is consumed in confectionery products or soups.

In Ghana, the major constraint to groundnut production is disease incidence, mainly early leaf spot (Cercospora arachidicola) and late leaf spot (Cercosporidium *personatum*), although rosette, rust (*Puccinia arachidis*) and Aspergillus flavus incidence may be severe depending on year and location. The effects of drought are particularly important in the northeast corner of the country where varieties that mature after 110 days are particularly unsuitable (Marfo and Padi 2000). Seed yield loss from leaf spot alone occurs in more than 40% of yield potential of the crop in northern Ghana (Tsigbey 1996). Bavistin (carbendazim) and Topsin-M (thiophanate methyl) are recommended for control of leaf spot; however, cost and availability of these fungicides have restricted their widespread use. The effects of foliar diseases and their interaction with moisture availability during the cropping season on groundnut performance has restricted earlymaturing varieties that are susceptible to the major foliar

diseases in the Sudan savannah ecology whereas latematuring varieties with resistance to foliar diseases are more preferable in the wetter Guinea savannah ecology (Marfo and Padi 1999).

Host plant resistance to the major diseases, and tolerance to drought will maintain yield stability of the crop and increase the profitability of production. To meet these objectives the Savanna Agricultural Research Institute (SARI), Ghana has been evaluating a number of advanced breeding lines of groundnut developed by the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, India for high seed vield and stability of vield at benchmark sites. Lines identified as high yielding are further evaluated for oil content. A number of lines that have high seed yield and high oil content have been identified over the years and were further tested in farmers' fields (Marfo 1997).

The National Varietal Release Committee of Ghana has released the groundnut varieties ICGV 92099 and ICGV 90084 on 9 October 2005 as Gusie-Balin and Kpanieli, respectively, for the northern sector of Ghana. ICGV 92099 is early in maturity (100 days) with high seed yield and resistance to early and late leaf spots. ICGV 90084 is a late-maturing variety (120 days) and is resistant to early and late leaf spots with high seed and oil yields. On a scale of 1 to 9 (where 1 = no leaf spot and 9 = complete defoliation due to leaf spot), scores for reaction to leaf spots for ICGV 92099 are consistently lower (4 to 5) than that of Chinese (7 to 9), the most important commercial cultivar in northern Ghana. Similarly, ICGV 90084 shows better resistance to leaf spots (score of 3 to 4) compared to Chinese or Manipintar (score of 4 to 5) and has similar reaction as F-mix. In advanced yield trials involving 17 lines tested at four sites across northern Ghana, ICGV 92099 produced seed yields

Variety	1996	1997	2001	2002	2003	2004
Early maturity group						
ICGV 92099			0.98	1.18	0.73	1.11
Chinese (check)			0.56	0.85	0.64	0.90
Sinkarzie (check)			0.70	0.98	0.91	1.10
Trial mean			0.69	0.96	0.75	1.13
LSD at 5%			0.23	0.20	0.15	0.27
Late maturity group						
ICGV 90084	1.16	1.22	1.05	1.51	1.18	2.12
F-mix (check)	0.89	1.21	0.88	1.06	1.25	1.92
Manipintar (check)	0.71	0.82	0.82	1.31	1.27	1.74
Trial mean	0.66	1.06	0.75	1.09	1.00	1.52
LSD at 5%	0.22	0.29	0.27	0.34	0.21	0.41

Table 1. Average seed yield (t ha-1) of groundnut varieties ICGV 92099 and ICGV 90084 across northern Ghana.
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Plant character	ICGV 92099	ICGV 90084
Branching pattern	Alternate	Alternate
Height of main stem ¹ (cm)	33	35
Plant spread ¹ (cm)	42	32
Stem pigmentation	Absent	Present
Peg pigmentation	Present	Present
Type of inflorescence	Compound	Compound
Standard petal color	Yellow	Yellow
Leaf color	Green	Light green
Leaflet length (cm)	4.2	5.0
Leaflet width (cm)	2.0	2.0
Leaflet shape	Wide-elliptic	Oblong-elliptic
Seed color	Brown	Red
Pod constriction	None	Very deep
Pod beak	Slight	Moderate
Pod length (cm)	3.2	3.2
Pod width (cm)	1.3	1.2
Seeds pod ⁻¹	2	2
Seed length (cm)	1.7	1.5
Seed width (cm)	1.1	1.0
100-seed mass (g)	70	67
Time to 50% germination (days)	5	5
Time to 50% flowering (days)	29	27
Time to maturity (days)	100	120
Potential seed yield (t ha ⁻¹)	2.0	2.5
Potential haulm yield ² (t ha ⁻¹)	4.0	5.0
Shelling outturn (%)	66	70
Oil content (%)	46	51

Table 2. Some characteristics of groundnut varieties ICGV 92099 and ICGV 90084 in northern Ghana.

2. After 4 days of continuous sun drying.

similar to, or better than the commercial varieties Chinese and Sinkarzie (Table 1). Also, among the late maturity group in which 14 advanced breeding lines were tested, ICGV 90084 produced seed yields similar to or higher than the late-maturing commercial cultivars Manipintar and F-mix. In 30 farmer-managed trials conducted between 2003 and 2004, ICGV 92099 and ICGV 90084 performed on average better than farmers' current varieties.

ICGV 92099 has alternate branching pattern, and the pods are typically two-seeded, slightly beaked, with no constriction between the seeds (Table 2). The oil content of 46% in ICGV 92099 is similar to that of Chinese. The large seed size of ICGV 92099 makes it attractive for developing confectionery-type products. ICGV 90084 also has alternate branching pattern, two-seeded pods that are moderately beaked with a deep constriction between the seeds (Table 2). ICGV 90084 has high oil content (51%) similar to that of F-mix (50%).

These new varieties provide opportunities for integrated management of leaf spot in northern Ghana, as the levels

of resistance are high. ICGV 92099 being early maturing will provide greater flexibility in planting time to obtain maximum yields in the Guinea savannah ecology, and proper utilization of available rainfall with reduced risks of terminal drought in the Sudan savannah ecology. ICGV 90084 is recommended for the Guinea savannah ecology alone because of its longer maturity period.

References

Marfo KO. 1997. The performance and association among some important groundnut yield traits in northern Ghana. Pages 133–140 *in* Improvement of cropping systems in the savanna zone: the challenges ahead (Mercer-Quarshie H, Marfo KO, Langyintuo AS and Owusu RK, eds.). Proceedings of the third conference on improving farming systems in the savannah zone of Ghana, 11–14 March 1993, Nyankpala Agricultural Experiment Station, Nyankpala, Tamale, Ghana. Nyankpala, Ghana: The Crop Research Institute (CRI)/ Nyankpala Agricultural Experiment Station (NAES) and Deutsche Gesellschaft für Technische Zusammenarbeit (GTZ) Joint Project.

Marfo KO and **Padi FK.** 1999. Yield stability of some groundnut accessions in northern Ghana. Ghana Journal of Agricultural Science 32:137–144.

Marfo KO and **Padi FK.** 2000. Evaluating groundnut (*Arachis hypogaea* L.) seed yield determinants in northern Ghana: a breeding perspective. Ghana Journal of Agricultural Science 33:23–28.

Tsighey FK. 1996. Integrated disease management in groundnuts: effects of neem seed extract, Bavistin and Topsin-M on foliar diseases of groundnut. Pages 126–130 *in* SARI Annual Report (Marfo KO and Owusu RK, eds.). Nyankpala, Tamale, Ghana: Savanna Agricultural Research Institute.

BRS Havana: A New Early-maturing Groundnut Variety for the Northeast Region in Brazil

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Groundnut (*Arachis hypogaea*) is one of the major oilseeds and food legumes in the world. It is an excellent food crop to reduce malnutrition due to rich nutritional properties of its oil and protein. As a short-season, annual tropical legume, it can be adopted in environments with low rainfall availability and distribution.

In Brazil, specially in the Northeast region, where malnutrition is a very serious problem, the consumption of groundnut derivatives represents a way to minimize this dietary deficiency, considering the low consumption of protein from animal origin.

The Brazilian Agricultural Research Corporation (Empresa Brasileira de Pesquisa Agropecuária – EMBRAPA) has researched groundnut crop for more than 20 years aiming to obtain high-yield potential and short-cycle groundnut cultivars, adapted to the semi-arid conditions and improvements in its seed quality, attending to Brazilian *in natura* market demands (low oil content, 3–4 seeds pod⁻¹ and red testa color seeds). Recently, confectionery market has increased in Brazil occupying 30% groundnut market. To cater to this demand, EMBRAPA

Table 1. Agronomical traits of groundnut cultivar BRS Havana.

Traits	Description
Pod beak, constriction and reticulation	Slight
Seeds pod ⁻¹	3–4
Flowering (days after plant emergence)	23
Maturity (days after plant emergence)	90
Pods plant ⁻¹	35-55
"Pops" (%)	<10
100-seed mass (g)	44-48
Pod yield (t ha ⁻¹) (rainy season)	1.9
Shelling outturn (%)	70-72
Oil (%)	43
Protein (%)	28
Oleic acid/linoleic acid ratio	1

released in 2005 BRS Havana, an early-maturing, tantesta color and drought-tolerant bunch type cultivar, recommended for confectionery market segments.

Origin and development

BRS Havana is a Valencia bunch type derived from CNPA 75 AM, a Brazilian accession belonging to Germplasm Collection of EMBRAPA and originated from Southeast region in Brazil. This accession was submitted to several selection cycles to shorten cycle (earliness), low oil content and adaptation to semi-arid environmental conditions. The breeding process lasted four years and was carried out in semi-arid region in five northeastern states.

Agronomic performance

In 30 yield trials, where evaluation was done under different ecological conditions in Northeast region, including nine breeding lines and two control cultivars, BR 1 (high yield) and BRS 151 L7 (early maturing), BRS Havana showed high pod yield and tolerance to drought. In rainy season, the pod yield was 1.9 tha⁻¹ and shelling outturn was about 71% (Table 1). BRS Havana has medium seed with 3–4 seeds pod⁻¹ and is early maturing.

Nutritional aspect

BRS Havana has tan testa color and is the lowest in crude oil (43%) among other Brazilian cultivars. The seed is composed mainly of linoleic acid (L) and oleic acid (O), which together make up 88% of the total unsaturated fatty acids. The O/L ratio is 1. BRS Havana contains 28% protein.

Pathology

Collar Rot of Groundnut Caused by Lasiodiplodia theobromae in North Vietnam

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Collar rot of groundnut (Arachis hypogaea), caused by Lasiodiplodia theobromae (Pat.) Griffon & Maubl. (syn Botryodiplodia theobromae Pat., Diplodia natalensis Pole-Evans and Diplodia gossypina Cooke), the anamorph of Botryosphaeria rhodina (Berk. & M.A. Curtis) Arx, was first recorded in the early part of the 20th century by Miller and Harvey (1932). In USA, L. theobromae caused severe damage in North Carolina in 1947 (McGuire and Cooper 1965) and some other sites. After these instances, there were no further reports of collar rot in USA until 1998 when the disease caused severe losses on groundnut in Virginia (Phipps and Porter 1998). Lasiodiplodia theobromae has now been isolated from stems, shells and seeds of groundnut from a number of countries including Australia (Bell et al. 2003), Chad (Sougnabe and Foko 2003), Egypt (El Habbaa et al. 2002), Gabon (Ndzoumba et al. 1990), India (Ramakrishna and Kolte 1984, Rao and Pande 1992), Indonesia (Dharmaputra and Retnowati 1996), Ivory Coast (Savary 1987) and Nigeria (Osuinde and Daibo 1999).

In Vietnam, collar rot was first reported on groundnut by Dan et al. (2000) and was especially severe in spring 2003, when the disease was found on more than 20% of plants at Dong Anh, Hanoi. Instances of this disease seem to be increasing especially in areas that have sandy soil and two crops of groundnut during one season. In this article, we present a description of the pathogen, the symptoms that it causes on groundnut in Vietnam and media for its culture.

Materials and methods

Seedlings and mature plants of groundnut were collected from fields at Tuliem and Dong Anh, Hanoi in 2002 and were incubated in a humid chamber for 3–8 days at 28°C. At the end of the season seeds were also collected, sterilized in 1% NaHClO for 1 min and rinsed with sterilized distilled water. The seeds were then placed on moistened filter paper in petri dishes and incubated under 12-h lightdark cycle at 26±2°C for 3–10 days. *Lasiodiplodia theobromae* on infected plants and seeds was identified by examination of spore-producing structures.

Isolate #77 obtained from groundnut stems collected from Tuliem, Hanoi in 2002 was used to determine mycelial growth on four different media: potato dextrose agar (PDA), Czapek Dox agar (CZA), carrot agar (CA) and white bean agar (WBA). These media were prepared according to the methods described by Dhingara and Sinclair (1995). Discs of 5-mm diameter from 2-day-old



Figure 1. A young groundnut plant infected by *Lasiodiplodia theobromae*.

mycelium of isolate #77 cultured on PDA at 30°C in the dark were placed on each medium and the plates incubated at 28°C for 3 days. Colony diameter was measured everyday for 3 days. There were 5 replicates per treatment.

After autoclaving, aliquots of PDA were adjusted to pH 4.0–7.5 using 0.1M HCl or 0.1M NaOH. These media were inoculated with 5-mm diameter discs of mycelium of *L. theobromae* #77 as described above. Mycelial growth was assessed 2 days after incubation at 28°C. There were 3 replicates per treatment.

Analyses of variance were performed using SAS statistical analysis software (SAS Institute Inc., Cary, North Carolina, USA) or STATISTICA software release 6.0 (StatSoft Inc., Tulsa, Oklahoma, USA, 2001). Treatment means were compared by Tukey's Honest Significant Difference (HSD) test at the 5% significance level.

Results and discussion

Although collar rot was first identified by Miller and Harvey in 1932, there have been few reports of this disease until relatively recently. Only one previous observation of collar rot has been made in Vietnam (Dan et al. 2000); however, the authors did not describe the symptoms. We have isolated *L. theobromae* from stems, seeds and necrotic tissues of infected groundnut and the symptoms were similar to those described by McGuire and Cooper (1965) and Subrahmanyam et al. (1992).

In Vietnam, infection with *L. theobromae* caused preand post-emergence damping-off or wilting of seedlings (Fig. 1). The leaflets and stems remained green until the seedlings died. Many black pycnidia were found on the collar of the seedlings at soil level. On mature plants, infection occurred on the collar region of the plant. The first symptom observed was chlorosis on leaflets on lateral branches followed by wilting and dehydration of single or several branches or the whole plant within a few days. Many black pycnidia were produced at the collar near soil surface and on stems, petioles and other necrotic tissues (Fig. 2). Infected seeds were covered with white to dark gray mycelium and caused soft rot. Black pycnidia were produced 3–8 days after incubation at 28°C.

Lasiodiplodia theobromae produced white, gray or black mycelium on PDA. On plant tissues, the pathogen produced black, nearly round pycnidia consisting of either single or multiple chambers. Single-chambered pycnidia ranged from 160 to 260 mm in diameter. The pycnidia and pycnidiospores from the isolate of *L. theobromae* used in this study were similar to the descriptions of Roger (1953), McGuire and Cooper (1965), Punithalingam (1976) and Phipps and Porter (1998). Immature conidia were singlecelled and hyaline. Mature conidia were black, 2-celled, thick walled and elliptical in shape measuring 10–16 µm × 17–28 µm. This size is within the range reported by Phipps and Porter (1998) (10–18 µm × 17–34 µm), Roger (1953) (11–15 µm × 20–30 µm) and Punithalingam (1976) (10– 15 µm × 18–30 µm).

Some authors studied various features of the culture of *L. theobromae* isolated from yam (*Dioscorea* sp), pineapple (*Ananas comosus*) and citrus (*Citrus* sp). However, there are no reports on the nutritional requirements of *L. theobromae* isolated from groundnut. In our study, *L. theobromae* grew well at pH 4 to 7 (Fig. 3); outside this range, growth was reduced. The mycelial growth of *L. theobromae* differed significantly (P < 0.05) on the four media tested. Growth was greatest on CZA and PDA, followed by WBA and CA resulting in colony

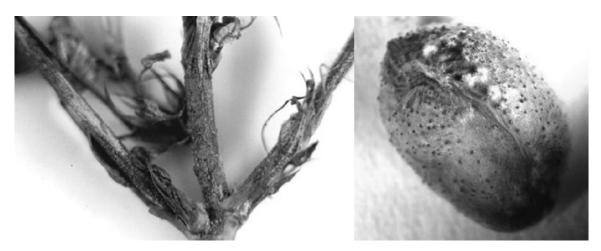


Figure 2. Black pycnidia of Lasiodiplodia theobromae produced on collar region of plant (left) and on seed (right) of groundnut.

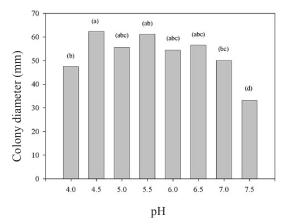


Figure 3. Mycelial growth of *Lasiodiplodia theobromae* on potato dextrose agar adjusted to different pH values. (Note: Means with the same letter are not significantly different at P = 0.05 according to Tukey's HSD test.)

diameter of 86, 81, 57.8 and 49.2 mm, respectively, after 3 days. On all media, colonies were lighter when young, and darker when old with the color changing from grayish to dark gray to black. However, further work is required to optimize the conditions for spore production as significant interactions occur among temperature, nutrition and irradiation in terms of both number of spores produced and the virulence on their host plant (Ghajar et al., in press).

Collar rot may occur more widely in Vietnam but may not be recognized clearly as some symptoms are similar to those caused by *Aspergillus niger*. Therefore, further investigations are necessary to help build a comprehensive picture of *L. theobromae* in Vietnam. Given that *L. theobromae* is a weak pathogen, work will be needed to determine why it is becoming more prevalent and to propose suitable methods for its control.

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References

Bell M, Harch G, Tatnell J and **Middleton K.** 2003. The impact of crop rotation on peanut productivity in rainfed cropping systems. Pages 1–4 *in* Solutions for a better environment. Proceedings of the 11th Australian Agronomy Conference, Geelong, Victoria, Australia, 2–6 February 2003.

Dan NT, Hong NX, Dung DT, Chinh NT, Dao VT, Toan PV, Long TD and Gowda CLL. 2000. Technologies to achieve high groundnut yields in Vietnam. Hanoi, Vietnam: Agriculture Publishing House. 73 pp.

Dharmaputra OS and **Retnowati I.** 1996. Fungi isolated from groundnuts in some locations of West Java. Biotropia 9:15–25.

Dhingara DO and **Sinclair BJ.** 1995. Basic plant pathology methods. Boca Raton, Florida, USA: CRC Press Inc. 434 pp.

El-Habbaa GM, Felaifel MS, Zahra AM and **Abdel-Ghany RE.** 2002. *In vitro* evaluation of some fungicides, commercial biocontrol formulations and natural plant extracts on peanut rootrot pathogens. Egyptian Journal of Agricultural Research 80:1017–1030.

Ghajar F, Holford P, Alhussaen K, Beattie A and **Cother E.** In press. Optimising sporulation and virulence in *Drechslera avenacea*. Biocontrol Science and Technology.

McGuire JM and **Cooper WE.** 1965. Interaction of heat injury and *Diplodia gossypina* and other etiological aspects of collar rot of peanut. Phytopathology 55:231–236.

Miller JH and **Harvey HW.** 1932. Peanut wilt in Georgia. Phytopathology 22:371–383.

Ndzoumba B, Conca G and Porta-Puglia A. 1990. Observations on the mycoflora of seeds produced in Gabon. FAO Plant Protection Bulletin 38:203–212.

Osuinde MI and **Daibo OO.** 1999. Wilt disease of *Arachis hypogaea* L. in Ekpoma, Edo State, Nigeria. Australian Journal of Experimental Agriculture 39:39–42.

Phipps PM and **Porter DM.** 1998. Collar rot of peanut caused by *Lasiodiplodia theobromae*. Plant Disease 82:1205–1209.

Punithalingam E. 1976. *Botryodiplodia theobromeae*. CMI descriptions of pathogenic fungi and bacteria. UK: Commonweath Agricultural Bureaux.

Ramakrishna N and **Kolte SJ.** 1984. Collar rot of groundnut. Indian Phytopathology 37:737–738.

Rao VG and **Pande A**. 1992. New host records of fungi from Maharashtra. Indian Phytopathology 45:136.

Roger L. 1953. Phytopathologie de pays chauds. Pages 1758–1766 *in* Chamgignons Inparfaits (Deuteromycetes ou Adelomycetes).

Savary S. 1987. A survey of fungal diseases of groundnut (*Arachis hypogaea*) in the Ivory Coast. I. Survey methods and descriptive study: cropping techniques and the main diseases. Netherlands Journal of Plant Pathology 93:167–188.

Sougnabe SP and **Foko J.** 2003. Knowledge contribution to groundnut (*Arachis hypogeae* L.) spermoflora parasitic fungus in the Mayo-Kebbi Basin of Chad. Pages 5–27 *in* Savanes africaines: des espaces en mutation, des acteurs face a de nouveaux defis, Actes du colloque, Garoua, Cameroun, 27–31 Mai 2002.

Subrahmanyam P, Wongkaew S, Reddy DVR, Demski JW, McDonald D, Sharma SB and Smith DH. 1992. Field diagnosis of groundnut diseases. Information Bulletin no. 36. Patancheru 502 324, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics. p. 34.

Entomology

White Grub Species Attacking Groundnut in the Saurashtra Region in Gujarat, India

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Groundnut (Arachis hypogaea) is extensively grown in the Saurashtra region of Gujarat state in India. Since 2003, white grub infestation has been frequently encountered in farmers' fields in the southern part of the Saurashtra region. The white grub damage varies with soil type. The lighter soils favor more activity of the pest. In heavily infested fields, 80% plant damage and 50% pod damage by white grubs, particularly Apogonia rauca, have been observed. An intensive survey and collection of white grubs was carried out during the rainy season in 2005 in the problematic areas of Visavadar taluka (Pindakhay, Sanosara, Kalsari and Jaliya villages) and Keshod taluka (Sergadh and Anida villages) of South Saurashtra. The grubs and adults were collected in August 2005 from the soil and also adults from host trees such as neem (Azadirachta indica), and ber (Zizyphus spp) and babul (Acacia spp), growing near the fields. The collected grubs were reared in the laboratory until adult emergence. Fourteen species of white grubs were identified in groundnut fields (Table 1).

Among these, *Holotrichia consanguinea* is a well known soil pest of groundnut. However, in our study, *A. rauca* was found predominant. The proportion of various white grub species population was: 80% *A. rauca*, 12% *H. consanguinea* and the remaining 8% all the other species. Some observations on biology and ecology of *A. rauca* were made. Its huge adult population was attracted to the host trees, particularly *babul* and was observed feeding upon them during early to midnight. Food preference of *babul* was confirmed in laboratory also. They were also attracted to light. As many as six grubs were seen near the plant root zone. They did not cut the root but fed on the nodules, rootlets and immature pods (Fig. 1). The adults also fed on the leaves of groundnut. The infested plants did not die but remained stunted with

weak growth. Thus the feeding habit of *A. rauca* apparently differed from the feeding habit of *H. consanguinea*. The full grown *A. rauca* grub measured 18 to 20 mm in length and 4 to 9 mm in width. The grub period lasted for 60 to 75 days. It pupated in earthen cocoon. The pupal stage lasted for 7 to 10 days. The freshly emerged beetle was reddish and then turned blackish the next day. It measured 10 mm in length and 5 mm in width (Fig. 2). It might have 2 to 3 overlapping generations during the groundnut season.

The infestation of *A. rauca* was noticed throughout the groundnut season (July to October). Severely infested fields failed to yield the healthy pods. Yadav (1987) listed predominant white grub species recorded in different parts of India whereas Nandagopal and Prasad (2004) compiled the world list of white grub species attacking groundnut. All the white grub species recorded in our study have previously been reported damaging groundnut except *Schizonycha ruficollis*, *Phyllognathus* sp and *Adoretus bicolor*. The investigation on seasonal activity and habitats of *A. rauca* and its management in groundnut has been initiated.

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Figure 1. White grub damage in groundnut: (a) damaged plants; (b) healthy plants; (c) damaged pods; and (d) healthy pods.

Table 1. White grub species recorded on groundnut inSaurashtra region in Gujarat, India.

White grub species	Coleopteran family		
Phyllognathus sp	Dynastinae		
Apogonia rauca Fabr.	Melolonthinae		
Holotrichia consanguinea Blanch.	Melolonthinae		
Holotrichia fissa Br.	Melolonthinae		
Holotrichia serrata Hope.	Melolonthinae		
Maladera sp	Melolonthinae		
Schizonycha ruficollis F.	Melolonthinae		
Adoretus bicolor Br.	Rutelinae		
Adoretus deccanus Ohaus	Rutelinae		
Adoretus versutus Harold	Rutelinae		
Adoretus sp	Rutelinae		
Anomala bengalensis Blanch.	Rutelinae		
Anomala dorsalis Fabr.	Rutelinae		
Anomala varicolor Gyll.	Rutelinae		



Figure 2. Different growth stages in white grub.

Sciences, GKVK, Bangalore, India for identification of the white grub species.

References

Nandagopal V and **Prasad TV.** 2004. World list of insect and non insect pests of groundnut and their natural enemies. Junagadh, Gujarat, India: National Research Centre for Groundnut. pp. 10–14.

Yadav CPS. 1987. Whitegrub – A national pest and strategies of its management. Pages 1–20 *in* Recent advances in entomology (Mathur YK, Bhattacharya AK, Pandey ND, Upadhyaya KD and Srivastava JP, eds.). Kanpur, Uttar Pradesh, India: New Gopal Printing Press.

Occurrence of White Grubs in Groundnut Crop in Uplands of South Vietnam: A New Report

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Groundnut (*Arachis hypogaea*) is an important crop in South Vietnam covering more than 125,000 ha under different cropping systems. Crop surveys, and the onfarm research organized in Trang Bang, Cuchi, Duc Hoa and Go Dau during the past (until 2000), brought out the importance of the foliage feeding insect pests (*Spodoptera, Helicoverpa*) as economically important in farmers' fields (Ranga Rao 1995). Field visits during the last week of May 2004 and interactions with the farmers in Tra Vinh province, villages around Cau Ngang town revealed the occurrence and importance of white grubs in this region. This soil-inhabiting pest is a menace in this area, which is in the heart of Mekong delta mostly covered by irrigated rice (*Oryza sativa*) cultivation with multiple cropping system.

During field visits, white grub adults were found feeding on nearby trees. Discussions with the farmers of My Thap village (Mai Van Tiep and colleagues) clearly brought out the importance of white grubs in their groundnut crops. According to farmers, these grubs infest crops such as groundnut, sugarcane (*Saccharum officinarum*), cowpea (*Vigna unguiculata*), cassava (*Manihot esculenta*) and maize (*Zea mays*) in uplands. The adults cause foliar damage in orchards particularly mango (*Mangifera indica*), cashew (*Anacardium occidentale*), litchi (*Litchi chinensis*), guava (*Psidium guajava*), etc. Among the various crops, groundnut and sugarcane were most severely affected. Though the adults were active during the nights, search for few hours in nearby mango and cashew orchards during daytime may yield several hundred adults.

Population dynamics of white grubs in Tra Vinh province

Based on the field observations and the farmers' experience, it was concluded that adults emerge soon after the summer rains (April–May) from their pupation sites (soil). The adults feed and mate at their feeding sites (cashew and mango trees). After feeding and mating, the adults return to their ovipositon sites (groundnut or any other upland crops). The young grubs are seen during June– July while weeding the groundnut crop. Generally groundnut is sown in these villages in the last week of May, which coincides with the adult emergence. Since the adult feeding sites are nearby the groundnut crops, it is easy for adults to locate the oviposition sites. After the harvest of May-sown crops, farmers takeup another groundnut crop in October. Thus two groundnut crops are grown in a year in the same field.

The adults are dull brown in color, measure about 25 mm in width and 40 mm in length with white markings on the posterior end of the elytra. The adults are identified as *Lepidiota signata* (Fig. 1). According to the farmers, the grub damage to May-sown groundnut crop was not severe, probably because the crop would be harvested before the grubs reach considerable size to inflict damage. The October-sown crops are affected severely because the crop is sown directly into grub-infested fields

Table 1. Calendar of even	nts in	white	grub	biology	in	Tra
Vinh province, Vietnam.						

Stage of the insect	Month of activity	
Adults	May–June	
Young grubs	June–July	
Well-grown grubs	September-October	
Grub developmental period	June-November	
Pupae	November-April	



Figure 1. Lepidiota signata adult.

and the well-grown grubs kill groundnut plants. Farmers observed grubs until November. Hence it is clear that the grub period extends from June to November (Table 1). However, detailed studies are required to define the developmental biology of this species in this region. According to farmers, total loss due to severe infestation on groundnut was not uncommon.

Though information is available on the importance of white grubs in North Vietnam pertaining to groundnut crop (Tran Huy Tho et al. 2001), the occurrence and the importance of white grubs in South Vietnam was not known. In view of the importance of Mekong delta for agricultural productivity and stability, the information pertaining to this pest is of immense value for sustaining the agricultural productivity in the upland areas of this region.

Control

Generally farmers apply basudin 10H at 10 kg ha⁻¹ as basal application in groundnut to manage this pest. Some farmers are also aware that soil application of carbofuran (furadan) granules 3 G at 1 kg ai ha⁻¹ controls the pest. However, the farmers are not clear about the efficient management of this pest.

Conclusions

- White grubs occur in upland areas of Mekong delta.
- *Lepidiota signata* causes loss to groundnut crops in Tra Vinh province of South Vietnam.
- Adults emerge in April–May soon after the summer rains.
- White grubs cause severe plant mortality in groundnut crop sown in October than in the crop sown in May.
- Basal application of basudin at the time of sowing gave satisfactory control.
- Several dryland crops such as sugarcane, cassava and maize are also infested by white grubs.
- Studies on the detailed biology, crop loss assessment, taxonomy and potential management strategies of white grub species are of high priority.
- We suggest to have a nation-wide white grub research project for effective control.
- Since the grubs pupate by November, delaying groundnut sowing to December wherever possible can help to overcome this menace.

References

Ranga Rao GV. 1995. Groundnut entomological work during spring 1995–96. Summary of sabbatical work on insect pests associated with peanut crop in Vietnam. ICRISAT Project Report. Patancheru 502 324, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics. 109 pp.

Tran Huy Tho, Pham Thi Vuong, Nguyen Thi Mao, Nguyen Chuc Quynh and **Pham Chi Hoa.** 2001. Some results of research on whitegrubs in upland crops and their management. Plant Protection Research and Extension Scientific Report. Vietnam: Oil Plant Institute of Vietnam. pp. 27–29.

Assessment of Integrated Pest Management Modules in Groundnut on Farmers' Fields

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Currently India is facing shortage of edible oils which is being met through large-scale imports. To meet the growing requirement of oil and to ensure nutritional security to the population of over one billion in the country, groundnut (Arachis hypogaea) has to play a pivotal role. Groundnut is a principal oilseed crop that suffers severe yield losses due to insect pests and diseases at different stages of crop growth. The defoliating caterpillars Spodoptera litura, Helicoverpa armigera and Amsacta albistriga, groundnut leaf miner (GLM) Aproaerema modicella, sucking pest Aphis craccivora, jassids and thrips attack the crop and cause economic loss. Early and late leaf spots, rust and blight are serious foliar diseases. Seedling crown rot, collar rot, stem and pod rot, and dry root rot are important soilborne diseases. Bud necrosis disease is one of the serious viral diseases of groundnut. When groundnut is grown in poor soil under inadequate growing condition, the crop becomes highly affected by these pests and diseases. Since groundnut is raised predominantly under rainfed conditions by resource-poor farmers who cannot afford the expensive agrochemicals (Rabindra 2004), intensive use of chemical pesticides as practiced during the era of green revolution is not a sustainable practice.

Though chemical pesticides have played an important role in increasing groundnut production, their indiscriminate use for the control of pests has led to several environmental problems such as development of resistance in pests to pesticides, pesticide residues and the destruction of beneficial parasites and predators of pests. Thus, a holistic, integrated pest management (IPM) program was developed in groundnut based on six years of independent research on entomological, pathological and weed management aspects conducted by the scientists of the National Research Centre for Groundnut (NRCG), Junagadh, Gujarat and All India Coordinated Research Project (AICRP) on groundnut at various centers in India. This IPM technology gave control of major insect pests ranging from 24 to 46% and diseases from 28 to 48% with an average increase in yield by 19% (Ghewande et al. 2002).

Integrated pest management options include diseasefree seeds of resistant/tolerant varieties, cultural practices [viz, use of castor (*Ricinus communis*) as a trap crop and intercropping system], usage of pheromones as monitoring tool, biocontrol agents, biopesticides and economic threshold level (ETL)-based chemical pesticides application. The IPM modules for groundnut based on production system can lead to higher crop production and conservation of biotic fauna. In shifting from chemical control to management of pests and diseases, IPM has to play a crucial role (Amerika Singh et al. 2004).

To attain high production level with minimum risk of pesticides contamination and risk of crop failure, this attempt has been made to assess and demonstrate the profitable viable intercropping system of groundnut and soybean (*Glycine max*) followed by utilization of IPM modules in comparison with farmers' practice.

Materials and methods

Field demonstrations were conducted through frontline demonstrations in non-replicated trial on farmers' fields in Jalgaon and Dhule districts of Maharashtra, India for three years in 2003, 2004 and 2005 during rainy season (July–November). Groundnut variety JL 286 (Phule Unap) and soybean variety JS 335 were sown in 0.2 ha area with a spacing of 30 cm \times 10 cm. A total of fifteen demonstrations (five in each year) was conducted including one at Oilseeds Research Station, Mahatma Phule Krishi Vidyapeeth (MPKV), Jalgaon Maharashtra.

The sowing was completed in 2^{nd} week of July and the trials were harvested in 2^{nd} week of November in each season. Two rows of castor were sown around the plot as

Item	IPM plot	Farmers' practice plot	
Cultivation			
Date of sowing	2 nd week of July	2 nd week of July	
Seed treatment	<i>Trichoderma</i> at 4 g kg ⁻¹ seed	_	
Type of sowing	Hand dibbling	Drilling	
Cropping system	Soybean and groundnut intercrop (4:1)	Sole crop (groundnut)	
Soil amendment	Castor cake at 500 kg ha ⁻¹	_	
Trap crop	Castor	_	
Pheromone trap	10 traps ha ⁻¹	_	
Plant protection ²	5% NSKE at 30 DAS and 50 DAS	Spray of Dimethoate at 0.03% at 35 DAS and Endosulfan at 0.07% at 60 DAS	
Pest incidence			
Aphids	1–5 plant ⁻¹	5–10 plant ⁻¹	
Thrips	15–25% at 30 DAS	20–50% at 30 DAS	
Leafhopper	10-20% at 45 DAS	20-25% at 45 DAS	
Spodoptera	10-30% at 60 DAS	35-50% at 60 DAS	
Groundnut leaf miner	2–15% at 80 DAS	55–70 % at 80 DAS	
Parasites/predators	Lady bird beetle,	Lady bird beetle,	
	Crysoperla, Bracon sp,	Crysoperla, Bracon sp,	
	syrphid fly, parasitization 5–10%	parasitization 5-10%	
Disease incidence (one week before harvest)			
Collar rot ³	2-5%	3-18%	
Stem rot	2-7%	5-10%	
Rhizoctonia root rot	<1%	1-2%	
Bud necrosis	2-5%	5-15%	
Early leaf spot	5-20%	10-20%	
Late leaf spot	20-30%	30-80%	
Rust	2-10%	20-25%	
Economics			
Yield (kg ha ⁻¹)			
Groundnut	1149	1076	
Soybean	516		
Gross income (Rs)	28558	20938	
Cost of cultivation (Rs)	20213	16226	
Net returns (Rs)	8345	4994	
Increase in net returns over farmers' practice (%)	42.3	_	
Benefit-cost ratio	1.43	1.31	

Table 1. Evaluation of IPM modules on farmers' fields in Maharashtra, India during rainy season 2003–05¹.

1. Data are means of three years of evaluation on fifteen farmers' fields.

2. NSKE = Neem seed kernel extract; DAS = Days after sowing.

3. At 30 DAS.

trap crop. Basal application of castor cake at 500 kg ha⁻¹ and seed treatment with *Trichoderma* spp at 4 g kg⁻¹ seed were carried out. Groundnut was intercropped with soybean at 4:1 ratio. Bird perches at 50 ha⁻¹ and pheromone traps at 10 ha⁻¹ (*Spodolure* and GLM) were fixed in each demonstration plot. Need-based sprays of neem seed kernel extract (NSKE) at 5% and spodo nucleo polyhederosis virus (SNPV) at 1.5×10^{13} ha⁻¹ were applied in each demonstration plot.

The demonstrations conducted at different locations were visited frequently and the incidence of pests and diseases was recorded periodically [starting from 30 days after sowing (DAS) to 80 DAS with 15 days interval] in both IPM as well as farmers' practice plots. To protect the groundnut crop from drought, two life saving irrigations were given. Dry pod yield of groundnut and grain yield of soybean were recorded after harvesting the crop. Economics of IPM and farmers' practice plots were worked out.

Results and discussion

The infestation of thrips and leaf hopper was severe at 30 to 45 DAS. The average damage by thrips was 15–25% in IPM plots as against 20–50% in farmers' practice plots (Table 1). The defoliators *Spodoptera* and GLM were observed at 60 and 80 DAS, respectively. The damage was reduced by 5–25% when spraying of NSKE at 5% and SNPV at 1.5×10^{13} ha⁻¹ was done in IPM plots. Intercropping of soybean in groundnut reduced the infestation of GLM (2–15%) in IPM plots as compared to farmers' practice plots (55–70%) (Ghewande et al. 1993). Significant differences in natural parasitization were not observed in both the practices. The population of lady bird beetle was meager in plots sprayed with insecticide.

The incidence of soilborne diseases was reduced by 20–50% in IPM plots as compared to farmers' fields. The foliar disease intensity, particularly late leaf spot was up to 30% in IPM plots whereas it was up to 80% in farmers' fields (Table 1).

There was considerable reduction of pest and disease incidence after adoption of IPM modules which realized high net returns of Rs 8345 ha⁻¹. Increase in income of 42.3% over farmers' practice was realized in IPM plots. Higher benefit-cost ratio of 1.43 was found in IPM plots compared to farmers' practice plots (1.31).

Conclusion

On the basis of demonstrations conducted on farmers' fields and analysis of results of three years revealed that the application of various components in IPM modules, viz, basal soil application of castor cake at 500 kg ha⁻¹, groundnut and soybean intercroping (4:1), seed treatment with *Trichoderma* spp at 4 g kg⁻¹, spraying of NSKE at 5% and SNPV $1.5\% \times 10^{13}$ ha⁻¹, 50 ha⁻¹ bird perches and 10 ha⁻¹ pheromone traps gave high net returns and an increase in income of 42.3% was incurred over farmers' practice.

References

Amerika Singh, Saroj Singh and Surender Kumar. 2004. Pest management in groundnut – An integrated approach. Page 62 *in* Souvenir of the National Symposium on Enhancing Productivity of Groundnut for Sustaining Food and Nutritional Security, 11–13 October 2004, Junagadh, India. Junagadh, Gujarat, India: National Research Centre for Groundnut.

Ghewande MP, Nandagopal V, Desai S and **Basu MS.** 2002. Integrated pest management in groundnut. Technical Bulletin. Junagadh, Gujarat, India: National Research Centre for Groundnut.

Ghewande MP, Prem Narayan and **Ingale AP.** 1993. Integrated management of foliar diseases of groundnut *Arachis hypogaea* L. in India. International Journal of Pest Management 39:375–378.

Rabindra R. 2004. Prospects for biological control of pests and diseases of groundnut. Pages 60–61 *in* Souvenir of the National Symposium on Enhancing Productivity of Groundnut for Sustaining Food and Nutritional Security,11–13 October 2004, Junagadh, India. Junagadh, Gujarat, India: National Research Centre for Groundnut.

Agronomy/Physiology

Companion Cropping of Spring Sugarcane and Summer Groundnut – A New Cropping System for Uttar Pradesh, India

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Among the cash crops grown in Uttar Pradesh, India, sugarcane (Saccharum officinarum) occupies a premier place and is grown on 2.1 million ha. It is a long-duration crop grown in widely spaced rows at 90 cm apart during spring season (15 February to March). From March to June, sugarcane planted during spring season attains around 30 cm height and the canopy does not cover the land adequately necessitating frequent weeding. If this period and the wide interrow spacing could be effectively used, not only weed infestation would be reduced but the farmers would get good return from the land early in the season. Earlier workers recommended intercropping of onion (Allium cepa), muskmelon (Cucumis melo), black gram (Vigna mungo), green gram (Vigna radiata), okra (Abelmoschus esculentus), etc with spring-planted sugarcane. No attempt was made on intercropping of summer groundnut (Arachis hypogaea) with spring-planted

Table 1. Yield of groundnut in spring sugarcane and summer
groundnut intercropping system.

Genotype	Pod yield of groundnut (t ha-1)	Increase over local check (%)
Dh 86	2.63	82.6
ICGS 1	1.87	29.9
ICGS 44	1.75	21.5
ICGV 86590	1.73	20.1
ICGV 93468	2.61	81.3
G 201 (local check)	1.44	_

sugarcane because summer groundnut has been cultivated in Uttar Pradesh only since 2001. In Uttar Pradesh, since 1982 to date the area under rainy season groundnut has declined from 0.3 million ha to 0.09 million ha and total production declined from 0.19 to 0.07 million t during 2004–05 over 1982–83. With the introduction and diffusion of groundnut cultivars ICGV 93468 and Dh 86 for cultivation during summer, the area under groundnut crop has increased from nil in 2001 to 27,500 ha in 2005 and 63,710 ha in 2006. This unprecedented success led to further research on summer groundnut.

An innovative adaptive experiment was planned on sandy loam riverine soils of Central Plain Zone V of Uttar Pradesh to increase the area under summer groundnut through utilization of vacant wider space of sugarcane rows from March to June. The operational area is situated in village Bhadauna of Unnao district between Bithor and Jankikund-Pariyar in the river Ganga catchment area. Sugarcane was planted at 90 cm row spacing after harvest of winter season vegetables on 25 February 2006. After partial completion of cane germination, the summer groundnut was sown on 20 March 2006 at row spacing of 25 cm. Three rows of summer groundnut were sown in between two rows of spring sugarcane and, thus, 100% plant stand of both crops was adjusted. The distance between sugarcane rows and groundnut rows was maintained at 20 cm from both the sides of sugarcane rows for easy intercultural operations. Six genotypes of groundnut, ie, Dh 86, ICGS 1, ICGS 44, ICGV 86590, ICGV 93468 and G 201 were tested in the intercropping system of spring sugarcane and summer groundnut. The recommended package of practices was followed for both the main crop and intercrop. The harvesting of summer groundnut was started 95 days after planting from 23 June 2006 and completed at 100 days after planting on 28 June 2006.

The groundnut cultivars Dh 86 (2.63 t ha⁻¹) and ICV 93468 (2.61 t ha⁻¹) registered significantly higher pod yield in the intercropping system of spring sugarcane and summer groundnut. Cultivar G 201 gave lowest yield of 1.44 t ha⁻¹. Therefore, the order of varietal performance was Dh 86 and ICGV 93468, followed by ICGS 1, ICGS 44, ICGV 86590 and G 201 in companion cropping of spring sugarcane and summer groundnut without any adverse effect of sugarcane on summer groundnut and vice versa (Table 1).

An Expert System for Cultivation and Management of Groundnut

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Expert system for crops such as wheat (*Triticum aestivum*), rice (*Oryza sativa*), maize (*Zea mays*), cotton (*Gossypium* spp) and rape (*Brassica napus*) have been developed successfully, but nothing is reported in the literature on the important oilseed crop groundnut (*Arachis hypogaea*). This article reports the development of an expert system for the cultivation and management of groundnut, which can not only improve the crop production but also modernize and standardize it.

Introduction of the system

Principle of the system. The basic design principle of the system is to combine cultivation technique with crop simulation, climate and soil condition. Because groundnut production system depends upon environmental factors and cultivation technique, the development of the system must consider these aspects (Shu-bo Wan 2003). The environmental factors refer to such factors as climate and soil. The technological factors are varieties chosen, sowing date, seed density, fertilizer use, seed treatment, prevention of waterlogging and drought, etc.

Structure and characteristics of the system. This system comprises databases, knowledge bases, simulation models, and data and knowledge acquisition system. It has a wide range of new knowledge and its practicability is strong. It is easy and simple to handle, easy to learn and use and is good at interaction. It includes multimedia and many kinds of accessory systems. This system also possesses edification, transparency and flexibility.

Functions of the system. The system mainly has three functions. First, make decisions on groundnut cultivation and management techniques. After getting the basic information, the system decides how to plant groundnut according to the user's field condition (Fig. 1). Second, dynamically simulate and regulate the growth of groundnut. In this part, the system can dynamically simulate groundnut growth, and also can judge whether there is a need to do some management. Third, retrieve and consult information

about groundnut and its cultivation. In this part users will be given any technique or information they want to know about groundnut and its cultivation.

Agronomy management decision module prior to sowing

This module is designed to first consider what aspects are related to the planting process. Secondly, it considers the influence factors of each aspect in the first step, then looks for the relationships between these influence factors, and in the end it solves these aspects according to the influence factors and relationships. Based on this thought, we set up seven sub-modules as follows: ascertaining cultivation method, optimizing fertilizer management, selection of variety, optimizing seed density, optimizing sowing date, optimizing seed treatment and consulation. These seven sub-modules were linked up effectively.

Optimizing landform and management. Choice of cropping system is important in groundnut cultivation to obtain high yields. There is a range of cropping systems in China, eg, spring-sown groundnut, summer-sown groundnut, and wheat and groundnut intercropping system. Planting is done on flat ground or on ridge. In most of the area, plastic mulching is adopted.

Users can follow the landform as required and then choose suitable cropping system and cultivation technique in this module. The system will teach the user these cultivation techniques through text and picture, and will provide guidance for better production.

Optimizing fertilizer management. In this module, the system carries out a method of nutrient balance to confirm the dosage of fertilizer. This method improves fertilizer-use efficiency that contributes to high income. The system considers the contents of soil nutrients, the contents of fertilizer and the dosage of the fertilizer (Zhen-wen Yu and Yong-shan Wan 1995). First, the system judges the soil fertility level according to the contents of soil nutrients, and then gives the objective vield according to the different soil fertility levels. If the user is satisfied with the objective yield, he/she can choose the fertilizer from the chemical fertilizer database. and the system will give the user the quantity of fertilizer to be applied. If the user does not agree with the objective yield, he/she can change it and then choose the fertilizer, then the system will calculate the quantity to be applied.

Selection of variety. For information on varieties, the system operates on the database directly. First, the system searches for useful groundnut varieties in the variety databases according to the user's demand, inserts them in a temporary table and expresses them to the user. To save space, after the user chooses the variety, the system will delete the table. It is easy to write the program codes and improve the efficiency of the system (Ai-hong Tong and Tai-ping Hou 2004).

Optimizing seed density. Confirming seed density should mainly consider three aspects. These are variety characters, soil fertility level and cropping system. First, we give every variety a suitable density according to its characters (Zhen-wen Yu and Yong-shan Wan 1995) and then judge the other two aspects. For example, if the soil fertility level is high, the system will give a lower density

compared with the suitable density, and if the soil fertility level is low, the system will give a higher density. Because it is difficult to give an exact number of the density, we let each level of the soil fertility match a coefficient, and we let the coefficient multiply the suitable density and get different density in different soil fertility levels.

Optimizing sowing date. This system confirms the sowing date according to various factors. These factors are period of duration, accumulated temperature, air temperature, ground temperature, previous crop and following crop, soil moisture, planting method, etc. Considering all these factors the proper sowing date is confirmed to ensure that the crop is grown in the most suitable condition. By doing this, one can fully utilize the favorable climatic conditions, avoid the influence of the unfavorable climatic conditions, and get good harvest.

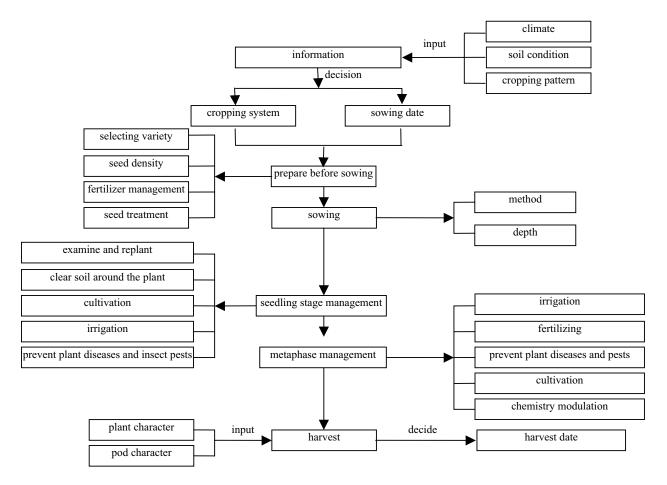


Figure 1. Flow diagram of the decision-making process of the expert system for groundnut cultivation.

Optimizing seed treatment. Seed treatment method is confirmed according to such factors as soil humidity, disease situation and soil nutrient condition. The method of seed treatment, the treatment step, the suitable chemical and the quantity will be given to the user through text, picture, video or kinescope before sowing.

Review of decisions. This module offers the decision scheme of individual event technology to the consultant before sowing. Users get advice about concrete questions when preparing before sowing. In addition, this module also offers information about groundnut variety resource, agriculture chemicals, chemical fertilizers, diseases, pests and weeds (Shu-bo Wan 2003).

Results and discussion

The advice of the system is based on the truthfulness of the user's response to the queries made by the system. The result can be saved in file, which contains record of the whole information across the process. We have now accomplished the program work and will start much more information related to groundnut cultivation, especially some simulation modules about groundnut growth. To make our expert system highly beneficial and acceptable, it will be based on multimedia technique, ie, the possibility of communication by text, pictures and sound. And then we will let it be examined in practice. We hope it will be applied to the production of groundnut widely in the future.

References

Ai-hong Tong and **Tai-ping Hou.** 2004. Visual basic database programming. Beijing, China: Tsinghua University. pp. 182–216.

Shu-bo Wan. 2003. Peanut cultivation in China. Shanghai, China: Shanghai Science and Technology. pp. 303–315 & 463–538.

Zhen-wen Yu and **Yong-shan Wan.** 1995. Crop culture studying. Beijing, China: Chinese Agriculture. pp. 179–209.

Utilization

Quality Attributes of Peanut Butter Prepared from Some Indian Groundnut Cultivars

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Peanut (*Arachis hypogaea*), also known as groundnut, is a major oilseed crop of India. However, unlike other oilseeds, groundnut can be consumed directly as food. With the growing awareness among people about the importance of balanced diet, demand for low calorie-high protein foods is increasing as people tend to avoid consumption of high-fat foods lest it should cause obesity and associated health problems.

The dairy butter is almost 100% fat and does not contain any protein while the peanut butter besides 50% fat, contains about 25% protein and all the other nutrients that are naturally present in groundnut. Hence consumption of groundnut in the form of peanut butter is more beneficial on the basis of economic and health aspects. It is already quite popular in USA and other European countries. In India, however, this product is available commercially only in the metropolitan cities. In times to come the demand for peanut butter in India is likely to grow owing to its nutritional value. For expulsion of oil at the oil mills, groundnut shell is added to the seed as a crushing aid. Thus the groundnut protein, which is obtained almost entirely in the form of groundnut cake, is no more useful for human consumption as it contains several extraneous substances, crushed shells, dust particles, insects and microorganisms. However, when groundnuts are processed for preparing peanut butter, no portion, except the red skin, is lost and hence the seeds are utilized rather in a wholesome manner as all the nutrients become available for human consumption. Thus popularization of peanut butter can go a long way in combating the problems of malnutrition. Consumers/ vendors would prefer the peanut butter to be easily spread and also have a long shelf life.

However, systematic information about the quality of peanut butter prepared from Indian cultivars is lacking. Therefore, it was of interest to study the quality attributes of peanut butter prepared from some of the Indian groundnut cultivars.

Materials and methods

Seven groundnut cultivars commonly grown in major groundnut producing states of India were selected for the study. The seeds of selected cultivars were obtained from the National Research Centre for Groundnut (NRCG), Junagadh, Gujarat, India. The peanut butter was prepared by following the procedure described by Tressler and Woodroof (1983). The seed lot of 150 g for each sample was spread over a petri dish of 177 cm² area, roasted at 130°C for 60 min using laboratory digital electrical oven (sensitivity 1°C). The seeds were cooled with forced air and then split to remove the skin and hearts. The weight of these roasted, blanched and split seeds devoid of hearts was recorded. Grinding was done in two steps. The blanched seeds were ground for 1 min at full speed in a domestic grinder and then the additives salt and sugar were added at the rate of 1% and 4%, respectively, of the weight of sample used for grinding. The mixture was again ground at full speed for 22 min. The butter samples were stored in glass jars with airtight plastic lids.

The butter samples so prepared were presented to a few persons for determining off taste, if any. A quantitative evaluation of organoleptic qualities by presenting the butter to panelists was not done owing to highly subjective nature of method.

Textural quality measurement. Textural quality was measured using Texture Analyser of Stable Micro Systems, UK (Model: TA-XT2i). The conical Perspex probe (code: P/45C) of 45° was penetrated into the sample by 14 mm with the pre-test, test and post-test speed as 2, 1 and 10 mm s⁻¹, respectively. The peanut butter samples were taken in a glass beaker (5.5 cm depth \times 2.5 cm diameter) and placed atop the load cell. The crosshead was set to move downward and penetrate the peanut butter sample for a distance of 14 mm. At the point of maximum penetration the crosshead direction of travel was automatically reversed and the probe was withdrawn at 10 mm s⁻¹ speed. Results were expressed as maximum force (g) required for cone penetration and withdrawal from peanut butter column. The adhesiveness measurements were selected according to the definition established

(Friedman, Whitney and Szezesniak 1963). The spreadability and firmness of butter was recorded in terms of maximum adhesive force required for cone penetration and withdrawal with the distance traveled by the probe. The probe used, represented the palate and the force required to remove the material from the probe complies with the definition of adhesiveness.

Proximate determination. For determination of moisture content, butter samples (10 g) were dried at 110°C for 10 hours in a hot air electrical oven. The oil and its fatty acid composition along with protein content of the butter prepared from selected groundnut cultivars were determined following the standard procedures. The oil content was determined gravimetrically by extracting the meal (10 g) with n-hexane in a Soxhlet extraction assembly for over 6 hours. The fatty acid composition of the oil was determined after converting the constituent fatty acids into their methyl esters, which were then separated on Nucon Gas Chromatograph (AIMIL, India) model 5700, fitted with a DEGS (polydiethylene glycol succinate) (2 mm internal diameter, 180 cm length) column. The temperature of the column was kept at 195°C while that of injection and flame ionization detector ports was kept at 250°C. The flow rates of carrier (nitrogen), fuel (hydrogen) and air were 40, 30 and 300 ml min⁻¹, respectively. The fatty acids were identified by comparison of their retention time with those of authentic samples. The area of a peak as fraction of the total area under all the peaks was expressed as per cent. The stability index (SI) was defined as the ratio of oleic acid (O) to linoleic acid (L) (Ahmed and Young 1982). The nitrogen content was determined by micro-Kjeldahl method using a Kjeltech auto nitrogen analyzer and the protein content was obtained by multiplying the nitrogen content of meal with a factor of 5.46 (St. Angelo and Mann 1973).

Color. The color code was assigned to butter preparations by visually comparing the color of butter preparation with those given in Methuen Handbook of Colour (Kornerup and Wanscher 1978).

Results and discussion

The moisture content of butter prepared from various cultivars was less than 1%. The butter of cultivar DRG 12 had the lowest moisture (0.54%) while that of cultivar JL 24 and BAU 13 had the maximum moisture content (0.74%).

No cultivar was found to produce any off flavor. Thus all the butter preparations were acceptable from the taste point of view.

As shown in Table 1, the butter prepared from GG 6 recorded the lowest adhesive force for cone penetration and withdrawal and this implied the ease in spreadability and firmness. This cultivar was followed by the butter prepared from ICGV 37 and Somnath. The maximum adhesive force for cone penetration and withdrawal was required for the peanut butter prepared from the seeds of BAU 13 followed by the butter prepared from DRG 12, JL 24 and ICGV 86325. The results revealed that among

Cultivar	Color ¹	Maximum adhesive		Moisture	Oil	Protein	Unsaturated fatty acids		
		force (dyne)	Oleic acid				Linoleic	O/L	
		Penetration	Withdrawal	(g kg ⁻¹)	(g kg ⁻¹)	(g kg ⁻¹)	(0) (%)	acid (L) (%)	ratio
ICGV 86325	AL	82.0	52.4	5.6	456	195	10.6	8.1	1.3
Somnath	GO	73.2	44.8	5.6	490	211	6.2	3.1	2.0
DRG 12	GO	94.3	56.6	5.4	494	197	4.6	3.4	1.4
GG 6	AY	63.3	38.9	6.8	511	232	4.1	3.1	1.3
JL 24	RG	90.8	81.2	7.4	504	242	10.0	7.9	1.3
ICGV 37	RG	66.9	45.6	5.8	501	216	16.0	11.7	1.4
BAU 13	AL	108.4	81.7	7.4	494	224	12.5	3.7	3.4
Maximum		108.4	81.7	7.4	511	242	16.0	11.7	3.4
Minimum		63.3	38.9	5.4	456	195	4.1	3.1	1.3
Mean		82.7	57.3	6.3	493	217	9.1	5.9	1.6

the selected groundnut cultivars GG 6 is the most appropriate followed by ICGV 37 and Somnath for the production of peanut butter. Thus it appears that firmness/spreading quality of peanut butter and oil content of the groundnut seed are closely related.

However, considering the peanut butter as an item of table-food both the nutritional quality as well as textural quality play an important role in the overall acceptability of the butter. The total oil and protein contents are important from nutritional point of view. The oil content of butter prepared from different groundnut cultivars varied between 45.6 and 51.1%. The lowest oil content was found in ICGV 86325 and highest in GG 6. Similarly, the protein content varied from 19.5 to 24.2%. Also, the shelf life of butter is determined by the SI, which is O/L ratio. The butter of BAU 13 cultivar exhibited the highest SI followed by cultivar Somnath. This implied that the butter prepared from cultivar BAU 13 had the highest shelf life followed by that from cultivar Somnath. The butter prepared from other cultivars recorded SI value less than 2.0 and thereby implying relatively a poor shelf life of the peanut butter.

Conclusion

The data indicated that the oil content in groundnut seed influenced the textural quality of peanut butter. Undesirable textural qualities might be due to low oil content. Therefore, on the basis of a combined textural and proximate evaluation, it could be recommended that amongst the cultivars evaluated, cultivar Somnath was the best suited for producing groundnut butter. While comparing the color attribute of the butter prepared from the selected cultivars, it was observed that those prepared from Somnath and DRG 12 reflected the most preferable grayish orange color. However, the differences among the colors of butter prepared from various cultivars evaluated were not significant.

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References

Ahmed EM and Young CT. 1982. Composition, quality and flavor of groundnut. Pages 655–688 *in* Peanut science and technology (Pattee HE and Young CT, eds.). Texas, USA: American Peanut Research and Education Society.

Friedman HJ, Whitney and **Szezesniak A.** 1963. The Texturometer, a new instrument for objective texture measurement. Journal of Food Science 28:396.

Kornerup A and **Wanscher JH.** 1978. Methuen Handbook of Colour. 3rd edition. London, UK: Eyre Ethuen Ltd. 252 pp.

St. Angelo AJ and **Mann GE.** 1973. Peanut proteins. Pages 559–592 *in* Peanut culture and uses. Raonoke, USA: Stone Printing Co.

Tressler DK and **Woodroof JG.** 1983. Food products formulary – Fruit, vegetable and nut products. Westport, Connecticut, USA: AVI Publishing Co. pp. 234–236.

Publications

Publications from ICRISAT

Pande S, Upadhyaya HD, Rao JN, Lakshmi Reddy P and **Parthasarathy Rao P.** 2005. Promotion of integrated disease management of ICGV 91114, a dual-purpose, early maturing groundnut variety for rainfed areas. Information Bulletin no. 68. Patancheru, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics. ISBN No. 92-9066-480-0. Order code IBE 068. 28 pp.

Groundnut (Arachis hypogaea L.) is an important oilseed crop that provides high quality oil for human consumption and fodder for ruminants. Its yields are very low in India due to several diseases as well as the non-availability of improved cultivars and their production technologies. Of these diseases, two foliar diseases, late leaf spot (Phaeoisariopsis personata) and rust (Puccinia arachidicola) are particularly destructive and together cause more than 70% of the losses in yield and quality. To add to this, about 80% of the area under groundnut in the Deccan Plateau is covered by the traditional cultivar TMV 2, which is highly susceptible to these foliar diseases. Considering various factors that limit groundnut production in the Deccan Plateau, scientists at ICRISAT emerged with an early-maturing, dual purpose cultivar, ICGV 91114 that was highly responsive to integrated disease management (IDM). The IDM package succeeded in consistently obtaining higher pod and fodder yields under farm conditions. The IDM technology comprised the improved early-maturing cultivar ICGV 91114, fungicide seed treatment with bavistin + thiram at 2.5 g kg⁻¹ seed, and one application of fungicide kavach at 65-70 days after sowing. The evaluation and promotion of ICGV 91114 and its IDM technology was carried out in three phases (1995-2004) in collaboration with ANGRAU, INGOs and NGOs. In all three phases, ICGV 91114 performed well, exhibiting lower severities of foliar diseases and higher pod and fodder yields. Moreover, in vitro tests at ICRISAT-Patancheru showed that the fodder from IDM-treated plots of ICGV 91114 had higher digestibility than TMV 2. During this period, the cultivar and its IDM technology spread to several villages in Andhra Pradesh, Karnataka and Tamil Nadu states in India.

Through all the years of testing, ICGV 91114 gave higher returns than the local cultivar. Participating farmers in all three states felt the new cultivar gave them higher quantities of pods and haulms as well as higher quality fodder that in turn translated to higher milk yields. The cultivar ICGV 91114, therefore, has rapidly become the favorite of several participating and non-participating farmers in the three states. Thanks to these advantages, ICGV 91114 and its associate IDM technology, which began with 11 farmers in 1995, spread to nearly 5000 farmers in 2002 and about 10000 farmers by 2005.

Pande S, Sreenivas B, Parthasarthy Rao P, Narayana Rao J and **Lakshmi Reddy P.** 2006. Farmers' participatory management of diseases for higher yield and nutritive value of crop residues of groundnut, Deccan Plateau, India. Patancheru 502 324, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics. ISBN 92-9066-485-1. Order code CPE 157. 168 pp.

Nigam SN, Aruna R, Giri DY, Ranga Rao GV and Reddy AGS. 2006. Obtaining sustainable higher groundnut yields: Principles and practices of cultivation. Information Bulletin no. 71. Patancheru 502 324, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics. ISBN 92-9066-484-3. Order code IBE 071. 48 pp.

With a few exceptions, groundnut productivity in most developing countries continues to be low. Although many high-yielding varieties have been released, their full potential is not realized in the absence of appropriate crop management practices. General agronomic recommendations are broad based and do not help much because of large variation in soil characteristics and nutrient status and other agroecological factors across groundnut fields. This bulletin discusses the underlying principles of various aspects of crop cultivation to encourage farmers to develop their own package of cultivation practices suitable to their fields and needs. It also provides information on groundnut cultivation under polythene mulch, which has resulted in 20–50% increase in groundnut productivity in China and on a seed production method to build self-reliance in the seed of improved groundnut varieties.

SATSource Listing

The following 2005 list of publications have been generated from ICRISAT's electronic bibliographic database SATSource – online database of the Semi-Arid Tropical Crops. Copies of entries can be obtained by writing to:

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Groundnut publications

Abd-Allah EF. 2005. Effect of a *Bacillus subtilis* isolate on southern blight (*Sclerotium rolfsii*) and lipid composition of peanut seeds. Phytoparasitica 33(5):460–466.

Adel-Patient K, Bernard H, Ah-Leung S, Créminon C and Wal JM. 2005. Peanut- and cow's milk-specific IgE, Th2 cells and local anaphylactic reaction are induced in Balb/c mice orally sensitized with cholera toxin. Allergy 60(5):658–664.

Adhikari S, Chakraborty T and Bagchi DK. 2005. Bioeconomic evaluation of maize (*Zea mays*) and groundnut (*Arachis hypogaea*) intercropping in drought-prone areas of Chotanagpur plateau region of Jharkhand. Indian Journal of Agronomy 50(2):113–115.

Adomou M, Prasad PVV, Boote KJ and Detongnon J. 2005. Disease assessment methods and their use in simulating growth and yield of peanut crops affected by leafspot disease. Annals of Applied Biology 146(4):469–479.

Aganga AA, Karikari SK and **Fabi L.** 2005. Incorporation of grass-hay, whole cereal grains (*Segaolane*) and bambara groundnut meal compared to lucerne into the diets of rabbits. Journal of Animal and Veterinary Advances 4(5):529–531.

Akhtar H, Hamid S and Khan JI. 2005. Comparative study of fatty acid composition and characterization of fixed oil of four peanut varieties available in Pakistan. Natural Product Sciences 11(4):225–228.

Akhtar J, Mehdi SM, Obaid-ur-Rehman, Mahmood K and Sarfraz M. 2005. Effect of deep tillage practices on moisture preservation and yield of groundnut under rainfed conditions. Journal of Agriculture and Social Sciences 1(2):98–101.

Amaefule KU and **Osuagwu FM.** 2005. Performance of pullet chicks fed graded levels of raw bambarra groundnut (*Vigna subterranean* (L.) Verdc) offal diets as replacement for soybean meal and maize. Livestock Research for Rural Development 17(5):55.

Angelini J, Taurian T, Morgante C, Ibáñez F, Castro S and **Fabra A.** 2005. Peanut nodulation kinetics in response to low pH. Plant Physiology and Biochemistry 43(8):754–759.

Anitha V, Wightman J and Rogers DJ. 2005. Management of white grubs (*Coleoptera: Scarabaeidae*) on groundnut in southern India. International Journal of Pest Management 51(4):315–322.

Anjana RM, Bharathi M, Rao PS and Kumar SS. 2005. Characterization of groundnut (*Arachis hypogaea* L.) genotypes through electrophoretic seed protein banding patterns. Research on Crops 6(3):600–605.

Arslan M. 2005. Effects of haulm cutting time on haulm and pod yield of peanut. Journal of Agronomy 4(1):39–43.

Asawaphan P, Mangkita W, Kachonpadungkitti Y, Matsuyama S, Satake T and Hisajima S. 2005. Efficient flower induction from peanut (*Arachis hypogaea* L.) seedling in vitro. SABRAO Journal of Breeding and Genetics 37(2):131–140.

Asis R, Barrionuevo DL, Giorda LM, Nores ML and Aldao MA. 2005. Aflatoxin production in six peanut (*Arachis hypogaea* L.) genotypes infected with *Aspergillus flavus* and *Aspergillus parasiticus*, isolated from peanut production areas of Cordoba, Argentina. Journal of Agricultural and Food Chemistry 53(23):9274–9280.

Avinash Dhawale A. 2005. Peanut and sunflower meal to replace SBM. World Poultry 21(2):12–13.

Azeredo GA de, Bruno R de LA, Lopes KP, Silva A da, Diniz E and Lima AA de. 2005. Peanut seeds conservation (*Arachis hypogaea* L.) as a function of conditioning, package and storage environment. Pesquisa Agropecuária Tropical 35(1):37–44.

Balkcom KS, Arriaga FJ and **Hartzog DL.** 2005. Narrow and wide strip tillage production for peanut. Pages 47–54 *in* Proceedings of the 27th Southern Conservation Tillage Systems Conference, Florence, South Carolina, USA, 27–29 June 2005.

Bandyopadhyay PK, Mallick S and **Rana SK.** 2005. Water balance and crop coefficients of summer-grown peanut (*Arachis hypogaea* L.) in a humid tropical region of India. Irrigation Science 23(4):161–169.

Banerjee K, Sounda G and **Mandal A.** 2005. Effect of different levels of irrigation and cobalt on growth and nodulation of summer groundnut (*Arachis hypogaea*). Journal of Interacademicia 9(2):235–241.

Barimalaa IS, Agoha G, Oboh CA and **Kiin-Kabari DB.** 2005. Studies on bambara groundnut flour performance in okpa preparation. Journal of the Science of Food and Agriculture 85(3):413–417.

Barros G, Torres A and **Chulze S.** 2005. *Aspergillus flavus* population isolated from soil of Argentina's peanut-growing region. Sclerotia production and toxigenic profile. Journal of the Science of Food and Agriculture 85(14):2349–2353.

Beena MR, Jami SK, Srinivasan T, Anuradha TS, Padmaja G and **Kirti PB.** 2005. Efficient direct shoot regeneration from cotyledonary node explants of peanut (*Arachis hypogaea* L. cv. JL-24). Indian Journal of Plant Physiology 10(3):205–210.

Bellettini NMT, Endo RM, Miglioranza É and **Santiago DC.** 2005. Pathogenicity of seed-borne and seedling fungi of groundnut cv. Tatu. Semina: Ciencias Agrárias (Londrina) 26(2):167–172.

Bergtold J, Norton G and **Brewster C.** 2005. Lomé to Cotonou Conventions: trade policy alternatives for the Senegalese groundnut sector. Agricultural Economics 33(3):315–323.

Bhanumathi P, Ganesan M and **Jayabalan N.** 2005. Physiological effect of organic mercury on the growth of peanut (*Arachis hypogaea* L.) seedlings. Plant Archives 5(2):665–669.

Bharud RW and **Pawar MR.** 2005. Physiological basis of yield variation in groundnut varieties under summer conditions. Journal of Maharashtra Agricultural Universities 30(1):100–102.

Bhatt RS, Sharma SR, Singh U, Kumar D and **Risam KS.** 2005. Effect of substituting groundnut cake for different levels of rice bran on growth and wool production of German Angora rabbits. World Rabbit Science 13(3):179–187.

Boldt A, Fortunato D, Conti A, Petersen A, Ballmer-Weber B, Lepp U, Reese G and **Becker WM.** 2005. Analysis of the composition of an immunoglobulin E reactive high molecular weight protein complex of peanut extract containing Ara h 1 and Ara h 3/4. Proteomics 5(3):675–686.

Branch WD. 2005. Registration of 'Georgia-04S' peanut. Crop Science 45(4):1653–1654.

Breuil S de, Giolitti F and Lenardon S. 2005. Detection of cucumber mosaic virus in peanut (*Arachis hypogaea* L.) in Argentina. Journal of Phytopathology 153(11/12):722–725.

Brown SL, Culbreath AK, Todd JW, Gorbet DW, Baldwin JA and **Beasley JP.** 2005. Development of a method of risk assessment to facilitate integrated management of spotted wilt of peanut. Plant Disease 89(4):348–356.

Casanoves F, Baldessari J and **Balzarini M.** 2005. Evaluation of multienvironment trials of peanut cultivars. Crop Science 45(1):18–26.

Casanoves F, Macchiavelli R and **Balzarini M.** 2005. Error variation in multienvironment peanut trials: within-trial spatial correlation and between-trial heterogeneity. Crop Science 45(5):1927–1933.

Chatterjee A, Dosani AAK, Talashikar SC and **Mehta VB.** 2005. Effect of lime on yield, quality and nutrient uptake by six groundnut varieties and properties of an alfisol. Journal of the Indian Society of Soil Science 53(1):128–132.

Chen L and **Phillips RD.** 2005. Effects of twin-screw extrusion of peanut flour on in vitro digestion of potentially allergenic peanut proteins. Journal of Food Protection 68(8):1712–1719.

Chenault KD, Melouk HA and **Payton ME.** 2005. Field reaction to sclerotinia blight among transgenic peanut lines containing antifungal genes. Crop Science 45(2):511–515.

Chi KL and **Lin CP.** 2005. Cloning and analysis of polC gene of phytoplasma associated with peanut witches' broom. Plant Pathology Bulletin 14(1):51–58.

Chitodkar SS, Bhoi PG, Patil HE and **Pawar PP.** 2005. Effect of irrigation regimes, mulches and antitranspirant on yield and yield contributing characters of summer groundnut. Journal of Maharashtra Agricultural Universities 30(2):230–232.

Chu CA and **Resurreccion AVA.** 2005. Sensory profiling and characterization of chocolate peanut spread using response surface methodology. Journal of Sensory Studies 20(3):243–274.

Chung SY and **Champagne ET.** 2005. Peanut polyamines may be non-allergenic. Journal of the Science of Food and Agriculture 85(6):990–994.

Chung SY, Kato Y and **Champagne ET.** 2005. Polyphenol oxidase/caffeic acid may reduce the allergenic properties of peanut allergens. Journal of the Science of Food and Agriculture 85(15):2631–2637.

Clavel D, Drame NK, Diop ND and **Zuily-Fodil Y.** 2005. Drought adaptation and breeding: the case of groundnut cultivated in Sahel area. OCL - Oléagineux, Corps Gras, Lipides 12(3):248–260.

Clavel D, Drame NK, Roy-Macauley H, Braconnier S and **Laffray D.** 2005. Analysis of early responses to drought associated with field drought adaptation in four Sahelian groundnut (*Arachis hypogaea* L.) cultivars. Environmental and Experimental Botany 54(3):219–230.

Das A and **Singh GP.** 2005. Effect of partial replacement of groundnut cake with berseem (*Trifolium alexandrinum*) on intake, rumen fermentation pattern, blood metabolites and growth of crossbred calves. Animal Nutrition and Feed Technology 5(1):61–72.

Deshmukh DD and **Dev DV.** 2005. Effect of package of practices on nodulation, branching, nitrogen and crude protein content in groundnut. Legume Research 28(1):17–21.

Deshpande RP, Chinnan MS and **McWatters KH.** 2005. Nutritional, physical and sensory characteristics of various chocolate-flavored peanut-soy beverage formulations. Journal of Sensory Studies 20(2):130–146.

Devi DR and **Rao NV.** 2005. Note on the performance of different groundnut pod-protectants against groundnut bruchid, *Caryedon serratus* (Olivier). Legume Research 28(2):155–156.

Devi DR and **Rao NV.** 2005. Some observations on the biology of groundnut seed beetle *Caryedon serratus* (Olivier) (Coleoptera; Bruchidae). Legume Research 28(3):229–230.

Devi SI, Vashista P and **Sharma CB.** 2005. Purification to homogeneity and characterization of two antifungal proteins from the roots of *Arachis hypogaea* L. National Academy Science Letters 28(1/2):21–28.

Dhawale MB and **Charjan YD.** 2005. Response of groundnut (Tag-24) grown after rabi sorghum to levels of fertilizers and plant densities. Journal of Soils and Crops 15(1):199–200.

Djè Y, Bonny BS and Zoro Bi IA. 2005. Preliminary observations of variability between some morphotypes of bambara groundnut (*Vigna subterranea* L. Verdc., Fabaceae)

from Côte d'Ivoire. Biotechnologie, Agronomie, Société et Environnement 9(4):249–258.

Dohlman E and **Livezey J.** 2005. Peanut backgrounder. Electronic Outlook Report from the Economic Research Service OCS-05i-01:30.

Dubey SC. 2005. Role of weather on development of cercospora leaf spot (*Cercospora arachidicola*) on groundnut (*Arachis hypogaea*). Indian Journal of Agricultural Sciences 75(4):232–234.

Dudde KB and **Malewar GU.** 2005. Soil test and plant analysis for diagnosing zinc deficiency in groundnut and mung. Annals of Plant Physiology 19(1):75–79.

Dudde KB and **Raut RS.** 2005. Combined effects of *Rhizobium* and VA-mycorrhiza inoculation on groundnut. Journal of Soils and Crops 15(2):315–318.

Dutta D and **Patra BC.** 2005. Response of groundnut (*Arachis hypogaea* L.) to sources and levels of sulphur fertilization in alluvial soils of West Bengal. Journal of Interacademicia 9(1):45–48.

Ebregt E, Struik PC, Odongo B and **Abidin PE.** 2005. Pest damage in sweet potato, groundnut and maize in northeastern Uganda, with special reference to damage by millipedes (Diplopoda). NJAS – Wageningen Journal of Life Sciences 5(1):49–69.

Egal S, Hounsa A, Gong YY, Turner PC, Wild CP, Hall AJ, Hell K and Cardwell KF. 2005. Dietary exposure to aflatoxin from maize and groundnut in young children from Benin and Togo, West Africa. International Journal of Food Microbiology 104(2):215–224.

Eizendeher LB, Freitas RJS de and **Cançado RA.** 2005. Incidence of aflatoxins B_1 , B_2 , G_1 and G_2 in groundnut sweets and natural groundnuts sold in the state of Paraná. Higiene Alimentar 19(129):101–104.

Ejigui J, Savoie L, Marin J and **Desrosiers T.** 2005. Influence of traditional processing methods on the nutritional composition and antinutritional factors of red peanuts (*Arachis hypogaea*) and small red kidney beans (*Phaseolus vulgaris*). Journal of Biological Sciences 5(5):597–605.

El-Habbasha SF, Kandil AA, Abu-Hagaza NS, El-Haleem AKA, Khalafallah MA and **Behairy TG.** 2005. Effect of phosphorus levels and some bio-fertilizers on dry matter, yield and yield attributes of groundnut. Bulletin of Faculty of Agriculture, Cairo University 56(2):237–252.

El-Shehaby AI and **Morsy SMA.** 2005. Biological control of peanut damping-off disease by *Bacillus sphaericus* soil treatment. Egyptian Journal of Agricultural Research 83(1):1–9.

Evangelista AR, Lopes J, Peron AJ, Castro Neto P and **Fraga AC.** 2005. Evaluation of nutritional value of sunflower, peanut, forage turnip and castor bean cakes. Documentos – Embrapa Soja 261:130–131.

Farris RL, Gray CJ, Murray DS and **Verhalen LM.** 2005. Time of removal of crownbeard (*Verbesina encelioides*) on peanut yield. Weed Technology 19(2):380–384.

Fischer R, McGhee JR, Huong Lan Vu, Atkinson TP, Jackson RJ, Tomé D and **Boyaka PN.** 2005. Oral and nasal sensitization promote distinct immune responses and lung reactivity in a mouse model of peanut allergy. American Journal of Pathology 167(6):1621–1630.

Frick OL, Teuber SS, Buchanan BB, Morigasaki S and **Umetsu DT.** 2005. Allergen immunotherapy with heat-killed *Listeria monocytogenes* alleviates peanut and food-induced anaphylaxis in dogs. Allergy 60(2):243–250.

Gandh AP, Joshi KC, Jha K, Parihar VS, Srivastav DC, Raghunadh P, Kawalkar J, Jain SK and **Tripathi RN.** 2005. Studies on alternative solvents for the extraction of peanut oil. Journal of Food Science and Technology 42(4):352–355.

Garg AK, Singh P, Sastry VRB and **Agrawal DK.** 2005. Replacement effect of groundnut cake with castor bean meal (*Ricinus communis*) in concentrate mixture of adult sheep. Indian Journal of Animal Sciences 75(6):688–690.

Ghosh PK and **Dayal D.** 2005. Optimization of date of sowing in a new groundnut-wheat relay cropping system in semi-arid tropics of India. Journal of Sustainable Agriculture 26(3):83–94.

Ghosh PK, Singh RK, Bandyopadhyay KK, Misra AK and **Manna MC.** 2005. Role of integrated plant nutrient supply for sustainable production in groundnut based cropping system in India. Fertiliser News 50(3):45–53.

Girdhar IK, Bhalodia PK, Misra JB, Girdhar V and **Dayal D.** 2005. Performance of groundnut, *Arachis hypogaea* L. as influenced by soil salinity and saline water irrigation in black clay soils. Journal of Oilseeds Research 22(1):183-187.

Glaspole IN, Leon MP de, Rolland JM and **O'Hehir RE.** 2005. Characterization of the T-cell epitopes of a major peanut allergen, Ara h 2. Allergy 60(1):35–40.

Golakia PR, Makne VG and **Monpara BA.** 2005. Heritable variation and association in Virginia runner and Spanish bunch group of groundnut. National Journal of Plant Improvement 7(1):50–53.

Gomes RLF and **Lopes** $\hat{A}C$ **de** A. 2005. Correlations and path analysis in peanut. Crop Breeding and Applied Biotechnology 5(1):105-110.

Gopalakrishna T and **Bhagwat AS.** 2005. Molecular marker studies in groundnut (*Arachis hypogaea* L.). Indian Journal of Genetics and Plant Breeding 65(3):159–166.

Gouri V, Reddy DR and **Rao SBSN.** 2005. Effect of weather parameters on total dry matter production and partitioning of rabi groundnut (*Arachis hypogaea* L.). Journal of Research ANGRAU 33(1):1–5.

Gouri V, Reddy DR, Rao SBSN and **Rao AY.** 2005. Thermal requirement of rabi groundnut in Southern Telangana Zone of Andhra Pradesh. Journal of Agrometeorology 7(1):90–94.

Grey TL and **Wehtje GR.** 2005. Residual herbicide weed control systems in peanut. Weed Technology 19(3):560–567.

Grichar WJ. 2005. Using herbicides in a peanut strip-tillage production system. Crop Management Aug:1–5.

Grichar WJ, Jaks AJ and **Besler BA.** 2005. Response of peanuts (*Arachis hypogaea*) to weather-based fungicide advisory sprays. Crop Protection 24(4):349–354.

Guarneri F, Guarneri C and **Benvenga S.** 2005. Identification of potentially cross-reactive peanut-lupine proteins by computer-assisted search for amino acid sequence homology. International Archives of Allergy and Immunology 138(4): 273–277.

Guerke WR. 2005. Evaluating peanut (*Arachis hypogaea* L.) seed vigor. Seed Technology 27(1):121–126.

Güzel E, Akcali ID, Mutlu H and **Ince A.** 2005. Research on the fatigue behavior for peanut shelling. Journal of Food Engineering 67(3):373–378.

Hadwani GJ and **Gundalia JD.** 2005. Effect of N, P and K levels on yield, nutrient content, uptake and quality of summer groundnut grown on Typic Haplustepts. Journal of the Indian Society of Soil Science 53(1):125–128.

Haggag WM and **Abo-Sedera SA.** 2005. Characteristics of three *Trichoderma* species in peanut haulms compost involved in biocontrol of cumin wilt disease. International Journal of Agriculture and Biology 7(2):222–229.

He GH, Meng RH, Gao H, Guo BZ, Gao GuoQing, Newman M, Pittman RN and Prakash CS. 2005. Simple sequence repeat markers for botanical varieties of cultivated peanut (*Arachis hypogaea* L.). Euphytica 142(1/2):131–136.

Holbrook CC and **Dong WB.** 2005. Development and evaluation of a mini core collection for the U.S. peanut germplasm collection. Crop Science 45(4):1540–1544.

Hurt CA, Brandenburg RL, Jordan DL, Kennedy GG and **Bailey JE.** 2005. Management of spotted wilt vectored by *Frankliniella fusca* (Thysanoptera: Thripidae) in Virginia market-type peanut. Journal of Economic Entomology 98(5):1435–1440.

Hussein GAM and **Yasin NMN.** 2005. Characteristics of extracted peanut oil and impact of its use in partial replacement of milk fat on the quality of ice cream. Egyptian Journal of Dairy Science 33(2):279–293.

Ibeawuchi II, Obiefuna JC and **Ofoh MC.** 2005. Effects of row spacing on yield and yield components of okra (*Abelmoschus esculentus*) and mixture groundnut (*Arachis hypogaea*). Journal of Agronomy 4(4):304–307.

Jain RK, Pandey AN, Krishnareddy M and Mandal B. 2005. Immunodiagnosis of groundnut and watermelon bud necrosis viruses using polyclonal antiserum to recombinant nucleocapsid protein of groundnut bud necrosis virus. Journal of Virological Methods 130(1/2):162–164.

Janakiraman N, Venkataravana P and Seenappa C. 2005. Effect of iron, zinc and boron on yield and seed quality of groundnut (*Arachis hypogaea* L.). Mysore Journal of Agricultural Sciences 39(2):286–288.

Jayalakshmi V and **Reddy GL.** 2005. Heterosis and inbreeding depression for yield and physiological attributes in groundnut. Indian Journal of Agricultural Research 39(1):25–30.

Jitendra Singh and **Singh DK.** 2005. Dehydrogenase and phosphomonoesterase activities in groundnut (*Arachis hypogaea* L.) field after diazinon, imidacloprid and lindane treatments. Chemosphere 60(1):32–42.

John K, Vasanthi RP, Venkateswarlu O and Naidu PH. 2005. Variability and correlation studies for quantitative traits in Spanish bunch groundnut (*Arachis hypogaea* L.) genotypes. Legume Research 28(3):189–193.

Johnson WC, Prostko EP and Mullinix BG. 2005. Improving the management of dicot weeds in peanut with narrow row spacings and residual herbicides. Agronomy Journal 97(1):85–88.

Jonnala RS, Dunford NT and **Chenault K.** 2005. Nutritional composition of genetically modified peanut varieties. Journal of Food Science 70(4):S254–S256.

Jonnala RS, Dunford NT and **Dashiell KE.** 2005. New higholeic peanut cultivars grown in the Southwestern United State. Journal of the American Oil Chemists' Society 82(2):125–128.

Joshi M, Niu C, Fleming G, Hazra S, Chu Y, Nairn CJ, Yang HY and Ozias-Akins P. 2005. Use of green fluorescent protein as a non-destructive marker for peanut genetic transformation. In Vitro Cellular & Developmental Biology – Plant 41(4):437–445.

Kalyani G, Sonali S, Reddy AS, Reddy AGS, Waliyar F and Nigam SN. 2005. Resistance to tobacco streak virus in groundnut, *Arachis hypogaea* L. Journal of Oilseeds Research 22(1):105–107.

Karanjikar PN, Jadhav GS, Wakle PK and **Pawar SB.** 2005. Effect of sowing dates and genotypes on dry matter partitioning in groundnut during post monsoon season. Journal of Maharashtra Agricultural Universities 30(1):83–84.

Karmakar S, Mittra BN and **Ghosh BC.** 2005. Effect of different organic materials with fly ash in integrated plant nutrient system for groundnut (*Arachis hypogaea*). Indian Journal of Agronomy 50(2):152–155.

Kaur G and **Singh P.** 2005. Effect of foliar application of ethrel and cobalt chloride on flower production pod to peg ratio (PPR) and seed yield in groundnut (*Arachis hypogaea* L.) cvs M-522 and SG-84. Environment and Ecology 23S(4):707–710.

Kiening M, Niessner R, Elizabeth, Baumgartner S, Krska R, Bremer M, Tomkies V, Reece P, Danks C, Immer U and Weller MG. 2005. Sandwich immunoassays for the determination of peanut and hazelnut traces in foods. Journal of Agricultural and Food Chemistry 53(9):3321–3327. King N, Helm R, Stanley JS, Vieths S, Lüttkopf D, Hatahet L, Sampson H, Pons L, Burks W and Bannon GA. 2005. Allergenic characteristics of a modified peanut allergen. Molecular Nutrition & Food Research 49(10):963–971.

Kiniry JR, Simpson CE, Schubert AM and Reed JD. 2005. Peanut leaf area index, light interception, radiation use efficiency, and harvest index at three sites in Texas. Field Crops Research 91(2/3):297–306.

Kishore GK and **Pande S.** 2005. Integrated applications of aqueous leaf extract of *Datura metel* and chlorothalonil improved control of late leaf spot and rust of groundnut. Australasian Plant Pathology 34(2):261–264.

Kishore GK and **Pande S.** 2005. Integrated management of late leaf spot and rust diseases of groundnut (*Arachis hypogaea* L.) with *Prosopis juliflora* leaf extract and chlorothalonil. International Journal of Pest Management 51(4):327–334.

Kishore GK, Pande S and **Podile AR.** 2005. Biological control of collar rot disease with broad-spectrum antifungal bacteria associated with groundnut. Canadian Journal of Microbiology 51(2):123–132.

Kishore GK, Pande S and **Podile AR.** 2005. Chitinsupplemented foliar application of *Serratia marcescens* GPS 5 improves control of late leaf spot disease of groundnut by activating defence-related enzymes. Journal of Phytopathology 153(3):169–173.

Kishore GK, Pande S and **Podile AR.** 2005. Management of late leaf spot of groundnut (*Arachis hypogaea*) with chlorothalonil-tolerant isolates of *Pseudomonas aeruginosa*. Plant Pathology 54(3):401–408.

Kishore GK, Pande S and **Podile AR.** 2005. Phylloplane bacteria increase seedling emergence, growth and yield of field-grown groundnut (*Arachis hypogaea* L.). Letters in Applied Microbiology 40(4):260–268.

Kishore GK, Pande S and **Podile AR.** 2005. Biological control of late leaf spot of peanut (*Arachis hypogaea*) with chitinolytic bacteria. Phytopathology 95(10):1157–1165.

Kishore GK, Pande S, Rao JN and **Podile AR.** 2005. *Pseudomonas aeruginosa* inhibits the plant cell wall degrading enzymes of *Sclerotium rolfsii* and reduces the severity of groundnut stem rot. European Journal of Plant Pathology 113(3):315–320.

Kladpan S, Mahakachanakul W, Yongmanitchai V, Boonyaratanakornkit M and Chinbuti A. 2005. Situation of aflatoxin contamination in groundnut and groundnut products in Thailand in 2004. Pages 557–564 *in* Proceedings of 43rd Kasetsart University Annual Conference, Thailand, 1–4 Feb 2005. Thailand: Kasetsart University.

Koppelman SJ, Jong GAH de, Laaper-Ertmann M, Peeters KABM, Knulst AC, Hefle SL and Knol EF. 2005. Purification and immunoglobulin E-binding properties of

peanut allergen Ara h 6: evidence for cross-reactivity with Ara h 2. Clinical and Experimental Allergy 35(4):490–497.

Kraitong T, Sarobol E, Sooksathan I and **Thongpae S.** 2005. Application of cow dung and chemical fertilizer for increasing yield of groundnut grown before rice in farmer's field at Amphur Non Sung Changwat Nakhon Ratchasima. Pages 288– 295 *in* Proceedings of 43rd Kasetsart University Annual Conference, Thailand, 1–4 Feb 2005. Thailand: Kasetsart University.

Krishna A and **Reddy MD.** 2005. Effect of methods of irrigation on performance of rabi groundnut (*Arachis hypogaea*) in red sandy loam soil. Journal of Research ANGRAU 33(2):66–68.

Krishnakanth A, Naidu GK and Gowda MVC. 2005. Selection for resistance to stem and pod rot in groundnut, *Arachis hypogaea* L. Journal of Oilseeds Research 22(1):226–227.

Krishnappa N, Narayanaswamy S and **Sreerama R.** 2005. Biochemical changes during storability in groundnut (*Arachis hypogaea* L.) as influenced by packing materials. Mysore Journal of Agricultural Sciences 39(2):214–218.

Kumar A, Lal D, Seth R and **Sharma V.** 2005. Detection of groundnut oil and vanaspati in ghee by thin layer chromatography. Indian Journal of Dairy Science 58(4):250–252.

Lancaster SH, Jordan DL, York AC, Wilcut JW, Brandenburg RL and Monks DW. 2005. Interactions of lateseason morningglory (*Ipomoea* spp.) management practices in peanut (*Arachis hypogaea*). Weed Technology 19(4):803–808.

Lancaster SH, Jordan DL, York AC, Wilcut JW, Monks DW and Brandenburg RL. 2005. Interactions of clethodim and sethoxydim with selected agrochemicals applied to peanut. Weed Technology 19(2):456–461.

Lanier JE, Jordan DL, Spears JF, Wells R and Johnson PD. 2005. Peanut response to inoculation and nitrogen fertilizer. Agronomy Journal 97(1):79–84.

Lavia GI and Fernández A. 2005. Karyotypic studies in *Arachis hypogaea* L. varieties. Caryologia 57(4):353–359.

Laxminarayana K and **Patiram.** 2005. Influence of inorganic, biological and organic manures on yield and nutrient uptake of groundnut (*Arachis hypogaea*) and soil properties. Indian Journal of Agricultural Sciences 75(4):218–221.

Lekberg Y and **Koide RT.** 2005. Arbuscular mycorrhizal fungi, rhizobia, available soil P and nodulation of groundnut (*Arachis hypogaea*) in Zimbabwe. Agriculture, Ecosystems & Environment 110(3/4):143–148.

Leon MP de, Drew AC, Glaspole IN, Suphioglu C, Rolland JM and O'Hehir RE. 2005. Functional analysis of cross-reactive immunoglobulin E antibodies: peanut-specific immunoglobulin E sensitizes basophils to tree nut allergens. Clinical and Experimental Allergy 35(8):1056–1064. Lewis SA, Grimshaw KEC, Warner JO and Hourihane JO. 2005. Promiscuity of immunoglobulin E binding to peanut allergens, as determined by Western blotting, correlates with the severity of clinical symptoms. Clinical and Experimental Allergy 35(6):767–773.

Li XM. 2005. Beyond allergen avoidance: update on developing therapies for peanut allergy. Current Opinion in Allergy and Clinical Immunology 5(3):287–292.

Liang XQ, Holbrook CC, Lynch RE and **Guo BZ.** 2005. β-1,3-glucanase activity in peanut (*Arachis hypogaea*) seed is induced by inoculation with *Aspergillus flavus* and copurifies with a conglutin-like protein. Phytopathology 95(5):506–511.

Lifrani A, Dubarry M, Rautureau M, Aattouri N, Boyaka PN and **Tomé D.** 2005. Peanut-lupine antibody cross-reactivity is not associated to cross-allergenicity in peanut-sensitized mouse strains. International Immunopharmacology 5(9):1427–1435.

Liu H, McCarty LB, Wells CE, Baldwin CM and Brown PJ. 2005. Greenhouse establishment study comparing peanut shell bio-solid with peat moss as an organic source for root zone mix. International Turfgrass Society Research Journal 10:1108–1114.

Livingstone DM, Hampton JL, Phipps PM and **Grabau EA.** 2005. Enhancing resistance to *Sclerotinia minor* in peanut by expressing a barley oxalate oxidase gene. Plant Physiology 137(4):1354–1362.

Luo M, Dang P, Bausher MG, Holbrook CC, Lee RD, Lynch RE and **Guo BZ.** 2005. Identification of transcripts involved in resistance responses to leaf spot disease caused by *Cercosporidium personatum* in peanut (*Arachis hypogaea*). Phytopathology 95(4):381–387.

Luo M, Dang P, Guo BZ, He G, Holbrook CC, Bausher MG and Lee RD. 2005. Generation of expressed sequence tags (ESTs) for gene discovery and marker development in cultivated peanut. Crop Science 45(1):346–353.

Luo M, Liang XQ, Dang P, Holbrook CC, Bausher MG, Lee RD and Guo BZ. 2005. Microarray-based screening of differentially expressed genes in peanut in response to *Aspergillus parasiticus* infection and drought stress. Plant Science 169(4):695–703.

Maeda K. 2005. Influence of the shell on the moisture content of fresh and dried groundnut seed. Japanese Journal of Crop Science 74(4):431–437.

Mahalakshmi P, Manivannan N and **Muralidharan V.** 2005. Variability and correlation studies in groundnut (*Arachis hypogaea* L.). Legume Research 28(3):194–197.

Mahalakshmi P, Manivannan N and Muralidharan V. 2005. Genetic divergence of groundnut (*Arachis hypogaea* L.) germplasm. Legume Research 28(3):220–222.

Mahan JR, Burke JJ, Wanjura DF and **Upchurch DR.** 2005. Determination of temperature and time thresholds for biotic irrigation of peanut on the Southern High Plains of Texas. Irrigation Science 23(4):145–152.

Main CL, Ducar JT, Whitty EB and MacDonald GE. 2005. Weed management in southeastern peanut with diclosulam and flumioxazin. Weed Technology 19(4):870–874.

Maiti MK, Raj SK and **Das S.** 2005. Management of leaf spot (*Cercospora arachidicola* and *Phaeoisariopsis personata*) of groundnut (*Arachis hypogaea*) by seed treatment with non-conventional chemicals. Indian Journal of Agricultural Sciences 75(7):452–453.

Mandal S, Samui RC and **Mondal A.** 2005. Growth, yield and yield attributes of groundnut (*Arachis hypogaea* L) cultivars as influenced by gypsum application. Legume Research 28(2): 119–121.

Marchiando NC, Zón MA and **Fernández H.** 2005. Determination of cercosporin (CER) phytotoxin isolated from infected peanut leaves by using adsorptive stripping square wave voltammetry. Analytica Chimica Acta 550(1/2):199–203.

Massawe FJ, Mwale SS, Azam-Ali SN and **Roberts JA.** 2005. Breeding in bambara groundnut (*Vigna subterranea* (L.) Verdc.): strategic considerations. African Journal of Biotechnology 4(6):463–471.

McDonald AH, Loots GC, Fourie H and **Waele D de.** 2005. Microplot study on *Ditylenchus africanus* population densities and damage symptoms on groundnut in relation to commercial yields. Nematology 7(5):647–653.

Misra JB and **Nautiyal PC.** 2005. Influence of imposition of soil moisture-deficit stress on some quality components of groundnut, *Arachis hypogaea* L. kernel. Journal of Oilseeds Research 22(1):119–124.

Mondoulet L, Paty E, Drumare MF, Ah-Leung S, Scheinmann P, Willemot RM, Wal JM and Bernard H. 2005. Influence of thermal processing on the allergenicity of peanut proteins. Journal of Agricultural and Food Chemistry 53(11):4547–4553.

Moraes ARA de, Lourenção AL, Godoy IJ de and Teixeira G de C. 2005. Infestation by *Enneothrips flavens* Moulton and yield of peanut cultivars. Scientia Agricola 62(5):469–472.

Morgante C, Angelini J, Castro S and **Fabra A.** 2005. Role of rhizobial exopolysaccharides in crack entry/intercellular infection of peanut. Soil Biology & Biochemistry 37(8):1436–1444.

Mothilal A, Muralidharan V and **Manivannan N.** 2005. Variability among five F_2 populations of intraspecific crosses of groundnut (*Arachis hypogaea* L.). Environment and Ecology 23(2):265–270.

Mupangwa WT and **Tagwira F.** 2005. Groundnut yield response to single superphosphate, calcitic lime and gypsum on acid granitic sandy soil. Nutrient Cycling in Agroecosystems 73(2/3):161-169.

Muthukumar N, Prasad JR and **Rao ZP.** 2005. Evaluation of legume tree leaves and groundnut haulms in sheep. Animal Nutrition and Feed Technology 5(2):153–162.

Mwale SS and **Azam-Ali SN.** 2005. Root growth and water extraction pattern of bambara groundnut (*Vigna subterranea* (L.) Verdc.) landraces. Aspects of Applied Biology 73:187–194.

Naab JB, Tsigbey FK, Prasad PVV, Boote KJ, Bailey JE and **Brandenburg RL.** 2005. Effects of sowing date and fungicide application on yield of early and late maturing peanut cultivars grown under rainfed conditions in Ghana. Crop Protection 24(4):325–332.

Nagaraja R, Murthy KVK and **Nagaraju.** 2005. Serological diagnosis of weeds and thrips harboring on them for the presence of peanut bud necrosis virus (PBNV). Environment and Ecology 23S(1):107–110.

Nagaraja R, Venugopal R, Murthy KVK, Jagadish KS and **Nagaraju.** 2005. Evaluation of groundnut genotypes against peanut bud necrosis virus (PBNV) and its thrips vector at Bangalore. Environment and Ecology 23S(1):118–120.

Naik CM and **Ramaswamy GR.** 2005. Role of spray adjuvants in improving efficacy of *Spodoptera litura* NPV on groundnut in coastal zone. Environment and Ecology 23S(1):17–19.

Naik RW, Deotale RD, Thorat A and **Titare P.** 2005. Effect of hormone and nutrients on yield and yield contributing characters of groundnut. Journal of Soils and Crops 15(1):213–215.

Nautiyal PC and **Misra JB.** 2005. Effect of drying methods on seed germination and seed protein profile in groundnut, *Arachis hypogaea* L. Journal of Oilseeds Research 22(1):125–128.

Nelson RG, Jolly CM, Hinds MJ, Donis Y and **Prophete E.** 2005. Conjoint analysis of consumer preferences for roasted peanut products in Haiti. International Journal of Consumer Studies 29(3):208–215.

Nepote V, Grosso NR and **Guzmán CA.** 2005. Optimization of extraction of phenolic antioxidants from peanut skins. Journal of the Science of Food and Agriculture 85(1):33–38.

Newman YC, Sollenberger LE, Boote KJ, Allen LH Jr, Vu JCV and **Hall MB.** 2005. Temperature and carbon dioxide effects on nutritive value of rhizoma peanut herbage. Crop Science 45(1):316–321.

Nigam SN, Chandra S, Sridevi KR, Bhukta M, Reddy AGS, Rachaputi NR, Wright GC, Reddy PV, Deshmukh MP, Mathur RK, Basu MS, Vasundhara S, Varman PV and Nagda AK. 2005. Efficiency of physiological trait-based and empirical selection approaches for drought tolerance in groundnut. Annals of Applied Biology 146(4):433–439.

Nolte C, Tiki-Manga T, Badjel-Badjel S, Gockowski J and **Hauser S.** 2005. Groundnut, maize and cassava yields in mixed-food crop fields after calliandra tree fallow in southern Cameroon. Experimental Agriculture 41(1):21–37.

Obasi MO. 2005. Influence of herbicides on growth and nodulation in Kerstings groundnut (*Kerstingiella geocarpa* Harms.). Crop Research (Hisar) 30(2):167–176.

Olorede BR and **Ajayi AF.** 2005. Replacement of groundnut cake and maize with *Falderbia albida* Goa in the diets of broiler chickens. Bulletin of Animal Health and Production in Africa 53(1):61–67.

Oumarou H, Ejoh R, Ndjouenkeu R and **Tanya A.** 2005. Nutrient content of complementary foods based on processed and fermented sorghum, groundnut, spinach, and mango. Food and Nutrition Bulletin 26(4):385–392.

Ozudogru EA, Ozden-Tokatli Y and **Akcin A.** 2005. Effect of silver nitrate on multiple shoot formation of virginia-type peanut through shoot tip culture. In Vitro Cellular & Developmental Biology – Plant 41(2):151–156.

Palanimuthu V, Ranganna B, Munishamanna KB, Gowda NAJ and **Kammar C.** 2005. Storage of groundnut seed in coastal region of Karnataka. Mysore Journal of Agricultural Sciences 39(1):64–69.

Panda PK and **Behera SK.** 2005. Irrigation water management strategy for peanut under deficit conditions. Zeitschrift für Bewässerungswirtschaft 40(1):91–114.

Passone MA, Resnik SL and **Etcheverry MG.** 2005. In vitro effect of phenolic antioxidants on germination, growth and aflatoxin B_1 accumulation by peanut *Aspergillus* section Flavi. Journal of Applied Microbiology 99(3):682–691.

Patel DP, Munda GC and **Islam M.** 2005. Dry matter partitioning and yield performance of HPS groundnut. Crop Research 30(2):156–161.

Pathirana RR, Watson L, Chen B, Leung S, Voisey C, Murray T and McManus MT. 2005. Removal of the N-linked glycan structure from the peanut peroxidase prxPNC2: influence on protein stability and activity. Phytochemistry 66(16):1869–1879.

Piersma SR, Gaspari M, Hefle SL and **Koppelman SJ.** 2005. Proteolytic processing of the peanut allergen Ara h 3. Molecular Nutrition & Food Research 49(8):744–755.

Plaut Z and **Ben-Hur M.** 2005. Irrigation management of peanut with a moving sprinkler system: runoff, yield, and water use efficiency. Agronomy Journal 97(4):1202–1209.

Poms RE, Agazzi ME, Bau A, Brohee M, Capelletti C, Norgaard JV and **Anklam E.** 2005. Inter-laboratory validation study of five commercial ELISA test kits for the determination of peanut proteins in biscuits and dark chocolate. Food Additives and Contaminants 22(2):104–112.

Pons L, Chéry C, Mrabet N, Schohn H, Lapicque F and **Guéant JL.** 2005. Purification and cloning of two high molecular mass isoforms of peanut seed oleosin encoded by cDNAs of equal sizes. Plant Physiology and Biochemistry 43(7):659–668.

Potter TL, Strickland TC, Joo H and **Culbreath AK.** 2005. Accelerated soil dissipation of tebuconazole following multiple applications to peanut. Journal of Environmental Quality 34(4):1205–1213. **Price TJ, Lamb MC** and **Wetzstein ME.** 2005. Technology choice under changing peanut policies. Agricultural Economics 33(1):11–19.

Quilambo OA, Weissenhorn I, Doddema H, Kuiper PJC and **Stulen I.** 2005. Arbuscular mycorrhizal inoculation of peanut in low-fertile tropical soil. I. Host-fungus compatibility. Journal of Plant Nutrition 28(9):1633–1644.

Quilambo OA, Weissenhorn I, Doddema H, Kuiper PJC and **Stulen I.** 2005. Arbuscular mycorrhizal inoculation of peanut in low-fertile tropical soil. II. Alleviation of drought stress. Journal of Plant Nutrition 28(9):1645–1662.

Raj SK, Mandal D and **Das S.** 2005. Rhizosphere mycoflora of groundnut, *Arachis hypogaea* L. in different organic and inorganic amended soil under field condition. Journal of Oilseeds Research 22(1):129–135.

Ramachandran L, Singh S and **Rathour AK.** 2005. Preparation of kulfi from admixtures of partially de-oiled groundnut meal and milk/milk powders. Natural Product Radiance 4(2):90–96.

Ramesh G and **Rao KHP.** 2005. Nutrient status of groundnut growing soils under rainfed conditions. Indian Journal of Dryland Agricultural Research and Development 20(1):35–40.

Rani AR and **Padmaja G.** 2005. Protocol for high frequency plant conversion from somatic embryos of peanut (*Arachis hypogaea* L. cv. DRG-12). Journal of Plant Biotechnology 7(3):1–7.

Rani AR, Reddy VD, Babu PP and **Padmaja G.** 2005. Changes in protein profiles associated with somatic embryogenesis in peanut. Biologia Plantarum 49(3):347–354.

Rao IVYR and **Raju VT.** 2005. Growth and instability of groundnut, *Arachis hypogaea* L. production in Andhra Pradesh: districtwise analysis. Journal of Oilseeds Research 22(1):141–149.

Reddy BR and **Reddy PM.** 2005. Studies on time of sowing of sunflower and weed management practices in groundnut and sunflower intercropping during kharif season. Indian Journal of Dryland Agricultural Research and Development 20(1):19–30.

Reddy SS. 2005. Effect of different organic manures on available NPK status and organic carbon after harvest of groundnut (*Arachis hypogaea* L.). Crop Research 30(1):26–29.

Reddy SS, Shivaraj B, Reddy VC and **Ananda MG.** 2005. Direct effect of fertilizers and residual effect of organic manures on yield and economics of maize (*Zea mays* L.) in groundnut-maize cropping system. Crop Research 30(1):1–5.

Reddy YS and **Reddy GP.** 2005. Economic analysis of sunflower versus groundnut production under rainfed conditions in Kurnool District, Andhra Pradesh. International Journal of Tropical Agriculture 23(1–4):269–282.

Rejeb SB, Abbott M, Davies D, Cléroux C and **Delahaut P.** 2005. Multi-allergen screening immunoassay for the detection

of protein markers of peanut and four tree nuts in chocolate. Food Additives and Contaminants 22(8):709–715.

Roberts G and **Lack G.** 2005. Diagnosing peanut allergy with skin prick and specific IgE testing. Journal of Allergy and Clinical Immunology 115(6):1291–1296.

Rogers DJ, Ward AL and **Wightman JA.** 2005. Damage potential of two *Scarab* species on groundnut. International Journal of Pest Management 51(4):307–314.

Rossetto CAV, Silva OF and **Araújo AE da S.** 2005. Storage peanut kernels fungal contamination and aflatoxin as affected by liming, harvest time and drying. Ciência Rural 35(2):309–315.

Rowland D, Dorner J, Sorensen R, Beasley JP Jr and **Todd J.** 2005. Tomato spotted wilt virus in peanut tissue types and physiological effects related to disease incidence and severity. Plant Pathology 54(4):431–440.

Ruchikachorn N, Chompreeda P, Haruthaitanasan V and **Chuenput S.** 2005. Formulation and process optimization of peanut sauce. Pages 435–443 *in* Proceedings of 43rd Kasetsart University Annual Conference, Thailand, 1–4 Feb 2005. Thailand: Kasetsart University.

Rudolf JR and **Resurreccion AVA.** 2005. Elicitation of resveratrol in peanut kernels by application of abiotic stresses. Journal of Agricultural and Food Chemistry 53(26):10186–10192.

Rudolf JL, Resurreccion AVA, Saalia FK and **Phillips RD.** 2005. Development of a reverse-phase high-performance liquid chromatography method for analyzing trans-resveratrol in peanut kernels. Food Chemistry 89(4):623–638.

Sahayaraj K and **Amalraj A.** 2005. Impact of monocrotophos and neem oil mixture on defoliator management in groundnut. Journal of Food, Agriculture & Environment 3(2):313–315.

Sakonnakhon SPN, Toomsan B, Cadisch G, Baggs EM, Vityakon P, Limpinuntana V, Jogloy S and Patanothai A. 2005. Dry season groundnut stover management practices determine nitrogen cycling efficiency and subsequent maize yields. Plant and Soil 272(1/2):183–199.

Sales RL, Costa NMB, Monteiro JBR, Peluzio M do CG, Coelho SB, Oliveira CG de and Mattes R. 2005. Effects of peanut, safflower, and olive oil on body composition, energy metabolism, lipid profile and food intake of eutrophic, normolipidemic subjects. Revista de Nutrição 18(4):499–511.

Samdur MY, Manivel P and **Mathur RK.** 2005. Genetics of iron-chlorosis related characters in groundnut, *Arachis hypogaea* L. Journal of Oilseeds Research 22(1):162–163.

Santos CEReS., Stamford NP, Freitas ADS, Vieira IM de MB, Souto SM, Neves MCP and Rumjanek NG. 2005. Effectiveness of rhizobia on peanut (*Arachis hypogaea*) nitrogen fixation, isolated from semi-arid soils of northeastern Brazil. Acta Scientiarum – Agronomy 27(2):301–307.

Saravanamoorthy MD and **Kumari BDR.** 2005. Effect of cotton yarn dye effluent on physiological and biochemical contents of peanut (*Arachis hypogaea* L. cv. TMV-10) and green gram (*Phaseolus radiatus* L. cv. K1). Biochemical and Cellular Archives 5(1):113–117.

Sastawa BM, Maina YT and **Lawan M.** 2005. Effects of sowing date modification and intercropping on the distribution of *Aphis craccivora* Koch (Hemiptera: Aphididae) in groundnut (*Arachis hypogaea*) in the Nigerian Sudan Savanna and implications for management. International Journal of Agriculture and Biology 7(2):298–303.

Savu RM, Choubey NK, Shrivastava GK and **Tiwari N.** 2005. Effect of chemical weed control on nitrogen uptake, weed weight and yield of groundnut under Chhattisgarh plains. Environment and Ecology 23S(3):400–402.

Schein CH, Ivanciuc O and **Braun W.** 2005. Common physical-chemical properties correlate with similar structure of the IgE epitopes of peanut allergens. Journal of Agricultural and Food Chemistry 53(22):8752–8759.

Sconyers LE, Brenneman TB, Stevenson KL and **Mullinix BG.** 2005. Effects of plant spacing, inoculation date, and peanut cultivar on epidemics of peanut stem rot and tomato spotted wilt. Plant Disease 89(9):969–974.

Sellamuthu KM and **Govindaswamy M.** 2005. Effect of humic acid on the mitigation of iron chlorosis of groundnut in red calcareous soil. Crop Research 29(1):106–110.

Sforza S, Scaravelli E, Corradini R and **Marchelli R.** 2005. Unconventional method based on circular dichroism to detect peanut DNA in food by means of a PNA probe and a cyanine dye. Chirality 17(9):515–521.

Shanmugam PM and **Balusamy M.** 2005. Polyethylene film mulching in groundnut – a way to getting higher yield. Research on Crops 6(3):462–464.

Sherif SA, Zohry AA and **Ibrahim ST.** 2005. Effect of planting dates and densities of maize intercropped with groundnut on growth, yield and yield components of both crops. Arab Universities Journal of Agricultural Sciences 13(3):771–791.

Shreffler WG, Lencer DA, Bardina L and **Sampson HA.** 2005. IgE and IgG4 epitope mapping by microarray immunoassay reveals the diversity of immune response to the peanut allergen, Ara h 2. Journal of Allergy and Clinical Immunology 116(4):893–899.

Singh J and **Singh DK.** 2005. Bacterial, azotobacter, actinomycetes, and fungal population in soil after diazinon, imidacloprid, and lindane treatments in groundnut (*Arachis hypogaea* L.) fields. Journal of Environmental Science and Health, Part B, Pesticides, Food Contaminants, and Agricultural Wastes 40(5):785–800.

Singh PK, Bhat AS, Ganai AM, Sarkar TK, Khan HM and Islam R. 2005. Effect of substitution of groundnut cake with

mustard cake on the growth performance and nutrients utilization of Corriedale lambs. Animal Nutrition and Feed Technology 5(2):163–170.

Singh RA. 2005. Response of fertilizers application on yield of chickpea under groundnut-chickpea cropping system. Farm Science Journal 14(1):16–18.

Singh RB and **Ali S.** 2005. Evaluation of groundnut genotypes for resistance against bud necrosis virus. Farm Science Journal 14(1):70.

Singh RN, Kumar B, Singh S and **Prasad NK.** 2005. Effect of boron application on groundnut and pigeonpea production in acid soils. Journal of Research, Birsa Agricultural University 17(1):7–10.

Singh S, Kaul JN and **Kaur N.** 2005. Productivity of summer planted groundnut in relation to land configurations and the seeding rates. Environment and Ecology 23(2):246–249.

Singh SK, Singh S and **Katti P.** 2005. Evaluation of IPM technology for groundnut- and sunflower-based production system. Entomon 30(3):201–205.

Singh YP, Sharma SC and **Maan JS.** 2005. Effect of sulphur on yield and its uptake in groundnut (*Arachis hypogaea*) and their residual effect on succeeding wheat (*Triticum aestivum*). Indian Journal of Agronomy 50(2):116–118.

Solanki RM, Bhalu VB, Jadav KV and **Kelaiya GR.** 2005. Studies on integrated weed management in irrigated groundnut. Indian Journal of Weed Science 37(1/2):119–120.

Sonali Shukla, Kalyani G, Kulkarni N, Waliyar F and **Nigam SN.** 2005. Mechanism of transmission of tobacco streak virus by *Scirtothrips dorsalis, Frankliniella schultzei* and *Megalurothrips usitatus* in groundnut, *Arachis hypogaea* L. Journal of Oilseeds Research 22(1):215–217.

Sounda G, Sharma A, Banerjee K and **Dey J.** 2005. Effects of different levels and sources of calcium on growth and nodulation of groundnut (*Arachis hypogaea* L.) during summer season. Environment and Ecology 23S(3):592–594.

Srichantawong M, Toomsan B, Limpinuntana V, Cadisch G, Jogloy S and Patanothai A. 2005. Evaluation of groundnut stover management strategies in a legume-rice rotation. Biological Agriculture & Horticulture 23(1):29–44.

Srinivas M, Mohammad S and **Sairam A.** 2005. Yield components and yield of castor (*Ricinus communis*) as influenced by different planting geometries and row proportions of intercropped groundnut or pearl millet. Crop Research 30(3):349–354.

Srinivas MS and Nagalingam B. 2005. Influence of certain microbial agents on *Spodoptera litura* (Hubner) and coccinellids in groundnut. Journal of Entomological Research 29(1):31–34.

Srinivas T, Manjulatha M and Venkateswarlu D. 2005. Biointensive management of collar rot, *Aspergillus niger* and stem rot, *Sclerotium rolfsii* Sacc. in groundnut, *Arachis hypogaea* L. Journal of Oilseeds Research 22(1):103–104.

Srivastava AK, Garg SK and **Dubey AK.** 2005. Optimization of operating parameters of a power operated groundnut decorticator. JNKVV Research Journal 38(1):70–77.

Srivastava KD, Kattan JD, Zou ZM, Li JH, Zhang LB, Wallenstein S, Goldfarb J, Sampson HA and Li XM. 2005. Chinese herbal medicine formula FAHF-2 completely blocks anaphylactic reactions in a murine model of peanut allergy. Journal of Allergy and Clinical Immunology 115(1):171–178.

Stein C, Koch P, Gude T, Battaglia R, Poms RE, Amadò R and **Anklam E.** 2005. How does the food industry handle the problem of peanut allergens during production? Deutsche Lebensmittel-Rundschau 101(7):293–301.

Strid J, Hourihane J, Kimber I, Callard R and **Strobel S.** 2005. Epicutaneous exposure to peanut protein prevents oral tolerance and enhances allergic sensitization. Clinical and Experimental Allergy 35(6):757–766.

Subrahmaniyan K and **Kalaiselvan P.** 2005. Flowering behaviour and reproductive growth of polyethylene filmmulched groundnut (*Arachis hypogaea*) intercropped with cotton (*Gossypium hirsutum*) under irrigated situation. Indian Journal of Agronomy 50(2):126–128.

Suriharn B, Patanothai A and Jogloy S. 2005. Gene effects for specific leaf area and harvest index in peanut (*Arachis hypogaea* L.). Asian Journal of Plant Sciences 4(6):667–672.

Tambe AD, Gaikwad CB, Pawar AD and **Walke VN.** 2005. Performance of herbicides against weeds in kharif groundnut. Annals of Plant Physiology 19(1):59–60.

Terzo E, Natera V, Isola MC, Fabra A, Franzoni L and **Castro S.** 2005. Effect of low pH on the enzyme activities of the ammonium assimilation pathways in the symbiotic association *Bradyrhizobium* sp. peanut (*Arachis hypogaea* L.). Symbiosis (Rehovot) 40(1):1–6.

Thomas WE, Troxler SC, Smith WD, Fisher LR and **Wilcut JW.** 2005. Uptake, translocation, and metabolism of sulfentrazone in peanut, prickly sida (*Sida spinosa*), and pitted morningglory (*Ipomoea lacunosa*). Weed Science 53(4):446–450.

Tschakert P, Khouma M and **Sène M.** 2005. Biophysical potential for soil carbon sequestration in agricultural systems of the Old Peanut Basin of Senegal. Journal of Arid Environments 59(3):511–533.

Tschakert P and **Tappan G.** 2005. Social context of carbon sequestration: considerations from a multi-scale environmental history of the Old Peanut Basin of Senegal. Journal of Arid Environments 59(3):535–564.

Tsitsigiannis DI, Kunze S, Willis DK, Feussner I and **Keller NP.** 2005. *Aspergillus* infection inhibits the expression of peanut 13S-HPODE-forming seed lipoxygenases. Molecular Plant-Microbe Interactions 18(10):1081–1089.

Tubbs RS and **Gallaher RN.** 2005. Conservation tillage and herbicide management for two peanut cultivars. Agronomy Journal 97(2):500–504.

Upadhyaya HD. 2005. Variability for drought resistance related traits in the mini core collection of peanut. Crop Science 45(4):1432–1440.

Upadhyaya HD, Swamy BPM, Goudar PVK, Kullaiswamy BY and **Singh S.** 2005. Identification of diverse groundnut germplasm through multienvironment evaluation of a core collection for Asia. Field Crops Research 93(2/3):293–299.

Vaghasia PM, Savalia RL and **Kelaiya GR.** 2005. Evaluation of frontline demonstrations on groundnut, *Arachis hypogaea* L. in Saurashtra region of Gujarat. Journal of Oilseeds Research 22(1):238–239.

van Odijk J, Ahlstedt S, Bengtsson U, Borres MP and Hulthén L. 2005. Double-blind placebo-controlled challenges for peanut allergy the efficiency of blinding procedures and the allergenic activity of peanut availability in the recipes. Allergy 60(5):602–605.

van Wijk F, Nierkens S, Hassing I, Feijen M, Koppelman SJ, Jong GAH de, Pieters R and Knippels LMJ. 2005. Effect of the food matrix on in vivo immune responses to purified peanut allergens. Toxicological Sciences 86(2):333–341.

Vanzolini S and **Nakagawa J.** 2005. Electrical conductivity test in peanut seeds. Revista Brasileira de Sementes 27(2):151–158.

Varma TSN, Dwivedi SL, Pande S and **Gowda MVC.** 2005. SSR markers associated with resistance to rust (*Puccinia arachidis* Speg.) in groundnut (*Arachis hypogaea* L.). SABRAO Journal of Breeding and Genetics 37(2):107–119.

Vasanthi RP, Devi GS, Babitha M and **Sudhakar P.** 2005. Inheritance of leaf chlorophyll content in groundnut (*Arachis hypogaea* L.). Indian Journal of Genetics and Plant Breeding 65(3):196–198.

Veenakumari K, Rabindra RJ, Naik CDS and Shubha MR. 2005. Field efficacy of nuclear polyhedrosis virus against the red hairy caterpillar, *Amsacta albistriga* (Walker) (Lepidoptera: Arctiidae) on groundnut in Karnataka. Journal of Biological Control 19(2):141–144.

Veeramani BB, Prakash M, Jagadeesan S, Kavimani S, Saravanan K and Ganesan J. 2005. Variability studies in M_2 generation of groundnut (*Arachis hypogaea* L.) var. VRI 2. Legume Research 28(1):68–70.

Vega VA. 2005. Rapid extraction of aflatoxin from creamy and crunchy peanut butter. Journal of AOAC International 88(5): 1383–1386.

Venkataramana P, Madhuprasad VL and **Anand TN.** 2005. Yield gaps and constraints in kharif groundnut cultivation – a case study of Kolar District in Karnataka. Mysore Journal of Agricultural Sciences 39(2):262–267. **Verstraeten SV, Hammerstone JF, Keen CL, Fraga CG** and **Oteiza PI.** 2005. Antioxidant and membrane effects of procyanidin dimers and trimers isolated from peanut and cocoa. Journal of Agricultural and Food Chemistry 53(12):5041–5048.

Viegas EC, Nascimento FG, Meyrelles BG and **Rossetto CAV.** 2005. Physiological quality of stored peanut seeds influenced by synthetic and vegetable products. Revista Brasileira de Plantas Medicinais 7(3):79–85.

Virk AS, Kaul JN, Bhangoo BS and **Singh A.** 2005. Influence of planting techniques and plant population on biology and pod productivity of summer groundnut varieties. Research on Crops 6(1):173–174.

Vu JCV. 2005. Acclimation of peanut (*Arachis hypogaea* L.) leaf photosynthesis to elevated growth CO_2 and temperature. Environmental and Experimental Botany 53(1):85–95.

Wen HW, Borejsza-Wysocki W, DeCory TR, Baeumner AJ and Durst RA. 2005. Novel extraction method for peanut allergenic proteins in chocolate and their detection by a liposome-based lateral flow assay. European Food Research and Technology 221(3/4):564–569.

Wen HW, Borejsza-Wysocki W, DeCory TR and Durst RA. 2005. Development of a competitive liposome-based lateral flow assay for the rapid detection of the allergenic peanut protein Ara h1. Analytical and Bioanalytical Chemistry 382(5):1217–1226.

Wheeler TA and **Black MC.** 2005. First report of cylindrocladium black rot caused by *Cylindrocladium parasiticum* on peanut in Texas. Plant Disease 89(11):1245.

Whitaker TB, Williams KM, Trucksess MW and Slate AB. 2005. Immunochemical analytical methods for the determination of peanut proteins in foods. Journal of AOAC International 88(1):161–174.

Woodward JE, Brenneman TB, Kemerait RC, Culbreath AK and Clark JR. 2005. First report of botrytis blight of peanut caused by *Botrytis cinerea* in Georgia. Plant Disease 89(8):910.

Xue HQ, Isleib TG, Payne GA, Novitzky WF and **Obrian G.** 2005. Aflatoxin production in peanut lines selected to represent a range of linoleic acid concentrations. Journal of Food Protection 68(1):126–132.

Yan LY, Xu ZY, Goldbach R, Kunrong C and **Prins M**. 2005. Nucleotide sequence analyses of genomic RNAs of peanut stunt virus Mi, the type strain representative of a novel PSV subgroup from China. Archives of Virology 150(6):1203– 1211.

Yang CMJ. 2005. Proteolysis, fermentation efficiency, and in vitro ruminal digestion of peanut stover ensiled with raw or heated corn. Journal of Dairy Science 88(8):2903–2910.

Yang CMJ. 2005. Soybean milk residue ensiled with peanut hulls: fermentation acids, cell wall composition, and silage utilization by mixed ruminal microorganisms. Bioresource Technology 96(12):1419–1424.

Yaranal RS and **Guruswamy T.** 2005. Performance of I.C. engine using blends of groundnut oil. Mysore Journal of Agricultural Sciences 39(3):294–299.

Yoshida H, Hirakawa Y, Tomiyama Y, Nagamizu T and **Mizushina Y.** 2005. Fatty acid distributions of triacylglycerols and phospholipids in peanut seeds (*Arachis hypogaea* L.) following microwave treatment. Journal of Food Composition and Analysis 18(1):3–14.

Yu JM, Ahmedna M and Goktepe I. 2005. Effects of processing methods and extraction solvents on concentration and antioxidant activity of peanut skin phenolics. Food Chemistry 90(1/2):199–206.

Yüksel B, Bowers JE, Estill J, Goff L, Lemke C and Paterson AH. 2005. Exploratory integration of peanut genetic and physical maps and possible contributions from *Arabidopsis*. Theoretical and Applied Genetics 111(1):87–94.

Yüksel B, Estill JC, Schulze SR and **Paterson AH.** 2005. Organization and evolution of resistance gene analogs in peanut. Molecular Genetics and Genomics 274(3):248–263.

Yüksel B and **Paterson AH.** 2005. Construction and characterization of a peanut HindIII BAC library. Theoretical and Applied Genetics 111(4):630–639.

Zade SR, Buldeo AN, Lanje PW and Gulhane VG. 2005. Evaluation of plant extracts and culture filtrates of bioagents against *Puccinia arachidis* Speg. in groundnut. Journal of Soils and Crops 15(1):150–154.

Information for IAN contributors

Publishing objectives

The International *Arachis* Newsletter (IAN) is published annually by the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) and the Peanut Collaborative Research Support Program (Peanut CRSP), USA. It is intended as a worldwide communication link for all those who are interested in the research and development of groundnut or peanut (*Arachis hypogaea* L.) and its wild relatives. Though the contributions that appear in IAN are peer reviewed and edited, it is expected that the work reported will be developed further and formally published later in refereed journals.

IAN welcomes short contributions (not exceeding 1000 words) about matters of interest to its readers. A few high quality full length papers may be accepted.

What to contribute?

Send us the kind of information you would like to see in IAN.

- Contributions should be current, scholarly, and their inclusion well-justified on the grounds of new information.
- · Results of recently concluded experiments, newly released varieties, recent additions to germplasm collections, etc.
- Genome maps and information on probe-availability and sequences, and populations synthesized for specific traits being mapped. Glossy black and white prints of maps should be included, if possible. Partial maps can also be submitted.
- Short reports of workshops, conferences, symposia, field days, meetings, tours, surveys, network activities and recently launched or concluded projects.
- · Details of recent publications, with full bibliographic information and 'mini reviews' whenever possible.
- Personal news (new appointments, awards, promotions, change of address, etc.)

How to format contributions?

- Keep the items brief remember, IAN is a newsletter and not a primary journal. About 1000 words is the upper limit (no more than four double-spaced pages). In exceptional cases, longer articles may be accepted.
- If necessary, include one or two small tables (and no more). Supply only the essential information; round off the data-values to just one place of decimal whenever appropriate; choose suitable units to keep the values small (eg, use tons instead of kg). Every table should fit within the normal type-written area of a standard upright page (not a 'landscape' page). Do not use the table-making feature of the word processing package; use simple tab set to prepare tables.
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About ICRISAT[®]

The International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) is a nonprofit, non-political organization that does innovative agricultural research and capacity building for sustainable development with a wide array of partners across the globe. ICRISAT's mission is to help empower 600 million poor people to overcome hunger, poverty and a degraded environment in the dry tropics through better agriculture. ICRISAT belongs to the Alliance of Centers of the Consultative Group on International Agricultural Research (CGIAR).

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