

Pulse Pathology (Pigeonpea)

Report of Work

(June 1985 - May 1986)



ICRISAT

International Crops Research Institute for the Semi-Arid Tropics
Patancheru, Andhra Pradesh 507 324, India

This report has been prepared to share the information with scientists having interest in pigeonpea improvement. This is not an official publication of the Institute and should not be cited.

YLN

PULSE PATHOLOGIST STAFF

Dr. Y.L. Nene	- Director, Legumes Program and Principal Plant Pathologist (Pulses)
Dr. M.V. Reddy	Plant Pathologist
Dr. M.P. Havare	Plant Pathologist
Dr. A.M. Ghanekar	- Plant Pathologist
Mrs. Sheila Vijayakumar	- Sr. Research Associate I
Mr. J. Narayana Rao	- Research Associate II
Mr. T.N. Raju	- Research Associate I
Mr. S. Veera Reddy	- Research Associate I
Mrs. Ganeswari Bajpai	- Clerk/Typist
Mr. A.P. Raju	- Sr. Field Assistant
Mr. K. Prabhakar Reddy	- Sr. Field Assistant
Mr. G. Musala Reddy	Sr. Field Assistant
Mr. M. Sharfuddin Khan	- Sr. Driver-cum-General Assistant II
Mr. M. Ramulu	Field Attendant
Mr. R. Penta Reddy	- Field Attendant
Mr. M. Satyanarayana	- Field Attendant
Mr. Rahim Pasha	- Field Attendant

PULSE PAID OLOGY (PIGEONPEA)
LIST OF APPROVED PROJECTS (1985-86)

No.	Title	Project Scientist(s)	Cooperators
CP-117(85)IC	Studies on pathogens causing wilt and root rots of chickpea and pigeonpea	M.P. Havare	Laxman Singh H.A. van Rheenen R. Jambunathan Y.S. Chauhan
CP-118(85)IC	Studies on pathogens causing blights of chickpea and pigeonpea	Y.L. Nene	B.L. Jalali (Hisar) S.P.S. Beniwal
CP-119(85)IC	Studies on the viruses affecting chickpea and pigeonpea	A.M. Ghanekar Y.L. Nene	D.V.R. Reddy S. Sithanantham Laxman Singh H.A. van Rheenen
CP-120(85)IC	Screening for disease resistance in chickpea and pigeonpea	M.V. Reddy A.M. Ghanekar H.A. van Rheenen Laxman Singh	Y.P.S. Rathi (Pantnagar) B.L. Jalali (Hisar)

Project: CP-117(85)IC - Studies on pathogens causing wilts and root rots of chickpea and pigeonpea

	Page No.
I. Summary	1
II. Introduction	1
III. Wilt	
A. Residual effect of solarization	1
1. Effect on <i>Fusarium</i> propagules in the soil	2
2. Effect on wilt	2
B. Confirmation of solarization effects	
1. Effect on <i>Fusarium</i> propagules in the soil	4
2. Effect on wilt	4
C. Isolation of antagonists.	4
D. Seed health testing	5
E. Greenhouse testing of soil (solarised and nonsolarised) for pigeonpea pathogens.	5
F. Seed pathology	
1. Studies on internal seed borne <i>F. udum</i> .	7
2. Eradication of <i>F. udum</i> from pigeonpea seed by seed treatment.	7
G. Search for perfect state of <i>F. udum</i>	
1. Examination of host tissue	9
2. Cultures of <i>F. udum</i> used in perfect state study	11
3. Methods used to induce perithecia	11
H. Studies on inheritance of wilt resistance	11

Project: CP-118(85)IC - Studies on the pathogens causing blights of chickpea and pigeonpea

	Page No.
I. Summary	14
II. Introduction	14
III. Phytophthora blight	
A. Pot screening method	14
1. Coordinated Varietal Trial (ACT)	14
2. Germplasm lines	19
3. Phytophthora blight resistant selections	19
4. Sterility mosaic promising lines	19
5. Phytophthora blight promising lines	19
6. Advanced lines	19
7. <u>Atylosia</u> species	19
B. Isolates from different locations	19
C. Physiologic Races	19
IV. Alternaria blight	27

Project: CP-119(85)IC - Studies on the viruses affecting chickpea and pigeonpea

	Page No.
I. Summary	28
II. Introduction	28
III. Biology of pathogens	
A. Sterility mosaic	28
1. Purification procedures	28
a) Using protoplasts	28
b) Using enzymes driselase	30
c) Using enzyme Rohament-P	31
d) Using procedures used for other mite transmitted viruses	32
2. Electron microscopy	
a) Mite-dip	34
b) Ultrathin sections of mite	34
c) Ultrathin sections of leaf	34
3. Mechanical sap transmission	35
4. Graft transmission	35
B. Yellow Mosaic	
1. Experimental host range of YM	36
2. Mechanical sap transmission of YM	36

Project: CP-120(85)IC - Screening for disease resistance in chickpea and pigeonpea

	Page No.
I. Summary	36
II. Introduction	40
III. Screening nurseries	
A. Wilt sick plots	40
B. Sterility mosaic nursery	41
C. Multiple disease nurseries	44
D. Wilt + <u>Heliothis</u> nursery	44
IV. Screening for resistance	
A. Wilt	44
1. Germplasm selections	44
2. ICPL lines	44
3. ACT materials	45
4. Breeding materials	45
5. MAHYCO lines	47
6. Multilocation testing	45
B. Sterility mosaic	
1. Germplasm	46
2. Germplasm selections	46
3. ACT materials	57
4. ACT selections	57
5. Breeding materials	57
6. MAHYCO lines	57
7. <u>Heliothis</u> tolerant lines	57
8. Pot screening of early ICPL's	57
9. Multilocation testing	71

	Page No
C. Phytophthora blight	71
1. Multilocation testing	71
D. Multiple disease resistance	
1. Wilt + SM	71
2. Wilt + SM + Phytophthora blight	72
E. Wilt + <u>Heliothis</u>	
1. Wilt + <u>Heliothis</u> tolerant lines	84
2. Breeding materials	84

Project: CP-117(85)IC - Studies on pathogens causing wilts and root rots of chickpea and pigeonpea

I. Summary

Previous season's results on solarization were confirmed. The solar heating of Fusarium infested soil reduced the Fusarium population in the soil. It also reduced the wilt significantly.

Solarization did not affect the pigeonpea seed quality i.e. seed germination and seed weight.

A soil antagonist, Penicillium pinophilum was isolated from solarised soil. It was found to be antagonistic to pigeonpea wilt pathogen (Fusarium udum) in laboratory studies.

Internal seedborne F. udum was eradicated from pigeonpea seed by treating them with mixture of Benlate + thiram (1:1).

Search for perfect state of F. udum continued without any positive evidence of its existence.

Studies on inheritance of wilt resistance in pigeonpea were initiated with a modified pot screening technique.

II. Introduction

The project became operative from May 1985 with the following objectives:

- (a) Investigate the biology and epidemiology of the pathogens.
- (b) Work out an integrated control system for each disease.
- (c) Assist breeders in inheritance studies.

Work carried out during 1985-86 is reported.

III. Wilt

A. Residual effect of solarization

The solar heating of the soil in a part of wilt sick plot (BIL 2B) by application of transparent polythene mulch during summer months in 1984, resulted in a significant decrease in Fusarium propagules in solarized soil and pigeonpea wilt. The experiment was repeated in 1985-86 season on the same plot to obtain confirmation of the previous season's results. Residual effects of solarization were also measured by halving block number and imposing + and - solarization treatments on each of the previous season's treatments.

Experiment 1: To compare fresh, residual and double solarization effects on pigeonpea in the same area where 1984-85 trial was conducted.

Details of the experiment

Location : Wilt sick plot (BIL 2B)

Treatments:

Main plot : 1. Irrigation before solarization

: 2. No irrigation

Sub plot : 8 treatments

Design : Split plot

Replications : 3

Sub plot size : 6 x 6 m

Genotypes : LRG-30(wilt susceptible);

ICP 8863(wilt resistant)

Duration of

Solarization : 26-4-1985 to 6-6-1985

1. Effect on Fusarium propagules in the soil

For Fusarium population count soil samples were collected immediately before and after solarization, in mid season (September) and at the time of plant maturity (January). For each treatment, 5 cores of soil to a depth of 15 cm were collected from 5 locations in a plot, bulked and thoroughly mixed. Soil samples were air-dried on a laboratory bench, grinded and sieved to remove stones. Population of F. udum was estimated on modified Nash and Snyder's medium. Measured quantity of soil (100 mg) was evenly spread on the surface of the medium in a sterilized petriplate. Petriplates were incubated at 25°C for 5 days and number of Fusarium colonies counted and calculated per gram of soil.

Residual effect of last year's solarization was evident when the soil samples were analysed for Fusarium propagules before polythene was laid on 26-4-84 (Table 1). Two years solarization (1984 and 1985) further reduced the population of Fusarium to less than 40 per gram of soil. Population of Fusarium was significantly less in the treatments where solarization was done this year (1985) than in the treatments with previous year's (1984) solarization. Fusarium propagules were estimated over 1000 per gram of soil in nonsolarized treatments. Similar trend was evident in all the treatments throughout the season, though steady increase of Fusarium propagules was observed in all the treatments.

2. Effect on wilt

The effect of soil solarization was clearly evident in reduced wilt incidence in the treatment with solarization in 1985. Residual effect of solarization in reducing the wilt incidence was not observed (Table 2). Also there was no significant difference in wilt incidence among the treatments with 2 years and one year (1985) solarization.

Table 1. Residual effect of solarisation on *Fusarium* propagules in pigeonpea wilt sick plot, ICRISAT Center, 1984 and 1985.

Sl. No.	Irrigation	Solarisation		Genotype	No. of <i>Fusarium</i> propagules/gm soil ¹			
		84	85		10.4.85	6.6.85	10.9.85	29.1.87
1.	Unirrigated	P	P	LRG 30	509.9	32.2	136.6	944.3
2.	Unirrigated	NP	P	"	1079.9	131.1	303.3	866.5
3.	Unirrigated	P	NP	"	276.6	809.9	744.4	860.0
4.	Unirrigated	NP	NP	"	777.7	1236.6	1351.0	1800.0
5.	Unirrigated	P	P	ICP 8863	561.1	31.1	112.2	394.3
6.	Unirrigated	NP	P	"	838.8	97.7	115.5	893.5
7.	Unirrigated	P	NP	"	445.5	541.0	545.5	1049.6
8.	Unirrigated	NP	NP	"	917.7	1131.0	1259.9	1966.1
9.	Irrigated	P	P	LRG 30	334.4	47.7	111.1	605.5
10.	Irrigated	NP	P	"	802.2	167.7	279.9	866.5
11.	Irrigated	P	NP	"	405.5	601.0	673.3	1611.0
12.	Irrigated	NP	NP	"	989.9	1199.8	1419.9	1502.3
13.	Irrigated	P	P	ICP 8863	252.2	39.9	124.4	577.4
14.	Irrigated	NP	P	"	927.7	137.7	356.6	544.3
15.	Irrigated	P	NP	"	294.4	809.9	759.9	1211.0
16.	Irrigated	NP	NP	"	798.8	1224.4	1311.1	1683.1
SE of solarisation with same or different levels of irrigation					91.9	91.5	92.3	176.6
SE of solarisation with same level of irrigation					82.4	84.2	89.7	188.7
CV(%)					22.3	28.3	26.2	30.0

¹Average of 3 replications; P=Polythene; NP=No Polythene.

pt

Table 2. Residual Effect of solarisation on wilt incidence in pigeonpea in wilt sick plot, ICRISAT Center, 1984 and 85.

Sl. No.	Irrigation	Solarisation		Genotype	Percent wilt ¹
		84	85		
1.	Unirrigated	P	P	LRG 30	30.99 (30.3)
2.	Unirrigated	NP	P	"	24.76 (28.0)
3.	Unirrigated	P	NP	"	60.46 (51.5)
4.	Unirrigated	NP	NP	"	51.36 (45.8)
5.	Unirrigated	P	P	ICP 8863	0.00 (0.00)
6.	Unirrigated	NP	P	"	0.09 (1.0)
7.	Unirrigated	P	NP	"	0.106 (1.1)
8.	Unirrigated	NP	NP	"	0.03 (2.5)
9.	Irrigated	P	P	LRG 30	4.95 (12.7)
10.	Irrigated	NP	P	"	6.15 (13.3)
11.	Irrigated	P	NP	"	62.84 (54.4)
12.	Irrigated	NP	NP	"	53.06 (47.0)
13.	Irrigated	P	P	ICP 8863	0.00 (0.0)
14.	Irrigated	NP	P	"	0.10 (1.0)
15.	Irrigated	P	NP	"	0.00 (0.0)
16.	Irrigated	NP	NP	"	0.20 (2.1)

SE of solarisation with same or different levels of irrigation (6.88)

SE of solarisation with same level of irrigation (7.07)

CV(%) (67.4)

The values in parentheses are after angular transformation.

¹Average of 3 replications; P=Polythene; NP=No Polythene

B. Confirmation of solarization effects

Experiment 2.(New):To study effect of solarization in pigeonpea.

The experiment was conducted in a new area in wilt sick plot to confirm the effects of soil solarization in pigeonpea obtained in previous year.

Treatments:

Main plot:1. Irrigation before solarization

2. No irrigation

Subplot: 1. Solarization, LRG 30

2. Solarization, ICP 8863

3. No solarization, LRG 30

4. No solarization, ICP 8863

Design: Split plot

Replications : 4

Duration of

solarization : 26-4-1985 - 6-6-1985

Subplot size : 6 x 6 m

1. Effect on *Fusarium* propagules in the soil

The effect of solarization on *Fusarium udum* population reduction was significant in irrigated and unirrigated treatments (Table 3). *Fusarium* propagules were drastically reduced to 190 to 275 per gram of soil in solarised treatments compared to 1045 to 1175 propagules per gram of soil in the nonsolarised treatments. The similar trend was observed throughout the season.

2. Effect on wilt

In genotype 1 (LRG 30) wilt was considerably reduced in unirrigated solarised treatment. However, similar trend was not observed in irrigated plots (Table 4). This may be due to contamination of solarised soil with *F. udum* from unsolarized plots and or ineffective solar treatment due to torned polythene. Wilt resistant, ICP 8863 remained practically wilt-free in all the treatments.

C. Isolation of antagonists

In order to study the changes in soil fungal flora, that would have been brought out due to soil solarization, soil samples from solarised and nonsolarised plots were analysed for fungi.

Potato dextrose agar to which Dicrysticin-S (1g/litre) was added after autoclaving was used for isolations. Measured quantity of soil (100 mg) was evenly spread on the surface of the medium in petriplates. Petriplates were incubated at 25°C for 5 days.

On the basis of their colony characters, 40 different fungi were isolated from solarised soils of chickpea and pigeonpea experiments. The isolated fungi were grown on PDA as pure cultures. On the basis of their frequency and growth rate, 15 fungal cultures were selected for further study.

These fungal cultures were then screened in the laboratory against F. udum and F. oxysporum f.sp. ciceri. All the cultures were multiplied on PDA in petriplates for a week. Active growth of the fungus from the edge of the colony was used by cutting with sterilized cork borer (3 mm). Growth of the test fungus along with one of the Fusarium was transferred in a petriplate containing potato-dextrose agar to grow as dual culture. Petriplates were incubated at 25°C and examined daily. By 7th day two test fungi over grew and covered petriplates. These two fungi showed their antagonism against F. udum in laboratory tests.

These two fungal cultures were sent to CMI, London for identification and were identified as Penicillium pinophilum Hedgcock. Further work is in progress.

D. Seed health testing

Pigeonpea seed harvested from solarization experiment was examined critically for any external infection and abnormality. Seed health testing of samples obtained from different treatments was done using blotter paper technique. Percent germination and 100 seed wt. were also recorded (Table 5). These studies were conducted with only irrigated treatments.

Seeds were also plated on selective medium (NS medium) for detection of Fusarium. Seeds from solarised and nonsolarised treatments were free from infection by Fusarium or any other fungus. On blotter test also we could not detect any pathogen on the seed. There was no apparent effect of solarization of seed germination and 100 seed wt. in wilt-susceptible and resistant cultivars.

E. Greenhouse testing of soil (solarised and nonsolarised) for pigeonpea pathogens

After the harvest of the crop in January 86, soil samples were collected from all the treatments for greenhouse testing. The objective was to examine if any other pigeonpea pathogen became dominant in solarised soil in which Fusarium was suppressed. Soil samples from 16 treatments were filled in 15-cm plastic pots (3pot/treatment). Five seeds of LRG 30 were planted in each pot. Pots were irrigated regularly and observed for 60 days for wilt and any other root rot.

Wilted plants were carefully examined from solarized (they were few) and also from nonsolarised treatment. Isolations resulted only in F. udum. There was no other pathogen detected in this experiment.

Table 3. Effect of solarisation on *Fusarium* propagules in pigeonpea wilt sick plot, ICRISAT Center, 1985

Sl. No.	Particulars	No. of <i>Fusarium</i> propagules/gm soil ¹			
		22.4.85	6.6.85	10.9.85	29.1.86
1.	Unirrigated polythene LRG 30	1272.4	190.0	168.3	677.5
2.	Unirrigated no polythene LRG 30	1190.8	1104.9	1549.1	1973.7
3.	Unirrigated polythene ICP 8863	1197.4	247.4	269.9	298.7
4.	Unirrigated no polythene ICP 8863	1323.3	1105.7	1517.4	2026.2
5.	Irrigated polythene LRG 30	1251.6	275.8	542.4	727.5
6.	Irrigated no polythene LRG 30	1279.1	1174.9	1474.1	2165.0
7.	Irrigated polythene ICP 8863	1326.6	254.9	499.9	798.7
8.	Irrigated no polythene ICP 8863	1217.4	1045.6	1467.4	2040.0
	SE of solarisation with same or different levels of irrigation	83.6	59.9	74.9	146.6
	SE of solarisation with same levels of irrigation	75.8	48.6	65.8	139.1
	CV(%)	12.0	14.7	14.0	20.8
Average of 4 replications					

Table 4. Effect of solarization on pigeonpea wilt in sick plot, ICRISAT Center, 1985.

Sl. No.	Particulars	Percent wilt ¹
1.	Unirrigated polythene LRG 30	25.40 (29.0)
2.	Unirrigated no polythene LRG 30	56.55 (53.9)
3.	Unirrigated polythene ICP 8863	0.07 (0.8)
4.	Unirrigated no polythene ICP 8863	0.24 (2.0)
5.	Irrigated polythene LRG 30	44.70 (42.1)
6.	Irrigated no polythene LRG 30	41.04 (39.5)
7.	Irrigated polythene ICP 8863	0.00 (0.0)
8.	Irrigated no polythene ICP 8863	0.16 (1.6)
	SE of solarisation with same or different levels of irrigation	(7.83)
	SE of solarisation with same levels of irrigation	(7.02)
	CV(%)	(66.50)

Values in parentheses are after angular transformation.

¹Average of 4 replications

F. Seed Pathology

In our earlier report (1983-84) we have reported the internal seed-borne nature of *Fusarium udum*. During the year of reporting we have confirmed our earlier finding that *F. udum* in pigeonpea seed was successfully eradicated by fungicidal seed treatment.

1. Studies on internal seedborne *F. udum*

Pigeonpea seeds were collected from seven cultivars at the time of wilting during January to April 85. Seeds were cleaned, air-dried and kept in closed containers in cold storage.

In all the experiments, unless stated otherwise, (i) 400 seeds of each cultivar were used, (ii) seeds were surface sterilized by a 2-min dip in 2.5% Sodium hypochlorite (clorox) and (iii) Nash and Snyder's medium was used for detection of *F. udum*. Surface sterilized and/or fungicide treated seeds (10/petriplate) were placed on medium. Petriplates were incubated at 25 C for 10 days.

The 'growing on' test was carried out by sowing seeds in autoclaved, fine riverbed sand in 5-cm plastic pots in a greenhouse. The plants were observed 60 days for wilt symptoms. Isolations were attempted from wilted plants.

It was confirmed that highly wilt susceptible cultivars like LRG-30 and ICP-2376 did not carry seed borne infection. Tolerant cultivars like BDN 1, HY 3C and C-11 allow the fungus to get establish in the seed before wilting (Table 6).

Isolates from seeds and wilted plants were identified as *F. udum*. They were pathogenic to pigeonpea.

2. Eradication of *F. udum* from pigeonpea seed by seed treatment

Seeds collected from diseased plants of BDN 1 and HY 3C in December 1986 were used in seed-treatment studies. Systemic fungicide formulations Benlate 50 W.p. (benomyl), Bavistin WP (carbendazim), Tecto-60 (Thiabendazole), thiram 75 WP and Dithane M-45 were tried. Two fungicidal mixtures, benlate + thiram (1:1) and bavistin + thiram (1:1) were also included for seed-treatment. The fungicidal dosage was 2.5 g of commercial formulation per kg of seed. The treated seeds were plated on medium and sown in pots in green house.

F. udum could not be detected from the seed treated with Benlate + thiram mixture. Benlate and Bavistin alone or with thiram reduced the incidence of the pathogen in the seed to the near check level.

Table 5. Percent seed germination and 100 seed wt.(g) of pigeonpea harvested from solarisation experiment.

S.No.	Irrigation	Solarization 84-85	Genotype	% Seed Germination	100 seed wt.
1	Irri.	P - P	LRG 30	93.50	6.47
2	Irri.	NP - P	LRG 30	94.25	6.88
3	Irri.	P - NP	LRG 30	96.25	6.38
4	Irri.	NP - NP	LRG 30	96.25	6.54
5	Irri.	P - P	ICP 8863	96.25	8.38
6	Irri.	NP - P	ICP 8863	94.75	8.47
7	Irri.	P - NP	ICP 8863	98.00	8.27
8	Irri.	NP - NP	ICP 8863	92.00	8.39
SE				1.103	
CV%				2.3	

P-Polythene, NP-No Polythene

of pigeonpea cultivars on selective medium
and in 'growing on' test.

Cultivar	Percent seed infected		Percent wilt	
	A	B	A	B
ICP 2376	0	0	0	0
BDN 1	3.75	0	3.75	0
ICPX 78148	2.50	0	2.25	0
ICP 1903	2.75	0	2.00	0
LRG-30	0	0	0	0
HY3C	4.50	0	4.0	0
C-11	2.00	0	1.50	0

A-Seeds from diseased plants;

B-Seeds from healthy plants.

Mixture of Benlate and thiram at the equivalent rate were again tried to confirm the earlier findings. Complete eradication was obtained with a fungicidal mixture of Benlate + thiram (1:1) at the rate of 0.25%. The treatment showed its effectiveness on the medium and also in 'growing on' test (Table 7A & 7B).

G. Search for perfect state of F. udum

The two major asexual spore forms of Fusaria are microconidia and macroconidia. Asexual state is important, since it is pathogenic and more frequently seen in nature.

In some of Fusaria, perfect state is reported. Sexual stage of Fusarium is ascomycetous, which is represented by one to two celled ascospores formed in ascocarp or perithecium. Recently, Gibberella indica has been reported as the perfect state of F. udum [from Varanasi]. Association of perithecia of Gibberella with wilted plants of pigeonpea was reported from Allahabad. Association of perithecia with wilted and dried pigeonpea plants was also observed by us at ICRISAT Center. But there was no evidence to relate its appearance to F. udum. In view of this, work was taken up to investigate the perfect state of F. udum.

1. Examination of the host tissue

Every year we distribute wilted pigeonpea plant pieces in wilt sick plot to maintain its sickness. Diseased material collected in January-February 85 was chopped into small pieces and spread in wilt sick plot in June 1985. Small stem pieces, thus distributed in the field were examined regularly under stereobinocular for perithecia. It was possible to collect the host tissues till December 85.

Wilted pigeonpea plants in fields at ICRISAT Center were also examined periodically. Diseased samples collected from Rahuri, Akola and Gwalior were also examined. Isolations were also attempted, whenever, fruiting bodies were seen on the surface of the stem pieces.

Following perithecial fungi were isolated from wilted pigeonpea plants.

- (a) Eupenicillium sp.
- (b) Ramichloridium sp.
- (c) Neocosmospora vasinfecta

N. vasinfecta was also isolated from wilted pigeonpea plants collected from Coimbatore.

Table 7A. Effect of fungicidal seed treatment on seed-borne Fusarium udum.

Fungicide	Infection detected on NS Medium (%)	
	BDN 1	HY3C
Benlate	2	5
Bevistin	3	2
Tecto 60	4	3
Thiram	4	7
Dithane M-45	7	8
Benlate + Thiram	0	0
Bevistin + Thiram	2	1
Check	12	17

Table 7B. Effect of fungicidal seed treatment on seed-borne Fusarium udum.

Cultivar	Infection(%)				Germination %	
	Nash & Snyder's medium		In sand		A	B
	A	B	A	B		
BDN 1	13.0	0	3	0	87.5	89.0
HY3C	19.25	0	4.25	0	96.25	95.0
ICP 1903 ²	9.75	0	-	-	-	-

1-Plants were observed for wilting up to 60 days after sowing.

2-ICP-1903 was used only in laboratory test due to shortage of seed

-Chlorax treated (Control), B-Fungicide treated

In addition, two fungi, Gliocladium caterulatum and Colletotrichum gloeosporioides were also isolated from pigeonpea plants showing wilt symptoms.

2. Cultures of F. udum used in perfect state study

F. udum cultures isolated from wilted pigeonpea plants from the following places are being used to induce perfect state. They are all pathogenic to pigeonpea.

Culture No. Location

- 1 ICRISAT Center (used in routine work)
- 2 ICRISAT Center (isolated from ICP 8863 in 1984.
- 3 Varanasi
- 4 ICRISAT Center (isolated from ICP 8863 in 1985
- 5 Gwalior
- 6 Isolated from several places in A.P.

3. Methods adopted to induce perithecia

- (a) Mating in culture
- (b) Inoculation of stem pieces with F. udum
- (c) Inoculation of pigeonpea seeds
- (d) Use of synthetic and nonsynthetic culture media

Inoculated materials are subjected to different temperatures. They are incubated for longer periods and examined periodically.

We hope to conclude this study next year and we will report our work in detail.

B. Studies on inheritance of wilt resistance

After perfecting the pot-screening technique (Pulse Pathology (Pigeonpea) Report of Work, June 1984-May 1985) we decided to use it in genetic studies. In this method 5-7-day-old seedlings grown in sterilized sand are root-inoculated with the inoculum multiplied on potato-sucrose broth. Seedlings are then transplanted in autoclaved riverbed sand or soil in 15-cm plastic pots.

Two separate screening tests were conducted to study the inheritance of resistance to F. udum in pigeonpea. In the first test, 7 parents and 21 F_2 populations of single crosses in diallel design were tested (Table 8). In the second test 6 lines, 4 testers and F_3 's were screened.

Table 8. F_2 populations of 21 single crosses (diallel design) screened for P. udum resistance by root dip technique.

Sl.No.	Cross	No. of pots	No. of plants tested
1.	ICP 8859 x ICP 8861	8	72
2.	ICP 8859 x ICP 8863	9	85
3.	ICP 8859 x ICPL 131	9	86
4.	ICP 8859 x ICP 10958	9	90
5.	ICP 8859 x ICPL 138	8	79
6.	ICP 8859 x ICP 6997	10	97
7.	ICP 8861 x ICP 8863	6	59
8.	ICP 8861 x ICPL 131	9	88
9.	ICP 8861 x ICP 10958	9	90
10.	ICP 8861 x ICPL 138	7	68
11.	ICP 8861 x ICP 6997	9	99
12.	ICP 8863 x ICPL 131	9	87
13.	ICP 8863 x ICP 10958	7	65
14.	ICP 8863 x ICPL 138	9	90
15.	ICP 8863 x ICP 6997	9	85
16.	ICPL 131 x ICP 10958	10	98
17.	ICPL 131 x ICPL 138	9	90
18.	ICPL 131 x ICP 6997	9	89
19.	ICP 10958 x ICPL 138	6	60
20.	ICP 10958 x ICP 6997	7	70
21.	ICPL 138 x ICP 6997	9	87

Seedlings (5-day old) were used. The roots were dipped in inoculum derived from single-spore culture of *F. udum* (ICRISAT isolate) for a minute and seedlings transplanted in sterilised sand in plastic pots (22 cm d.). Each pot had 10 F plants, 2 plants of susceptible check (LRG 30) and parents. Number of plants showing wilt symptoms were recorded periodically. Final observations were taken after 60 days.

Project: CP-118(85) IC - Studies on the pathogens causing blights of chickpea and pigeonpea

I. Summary

1. A total of 116 ACT entries were screened in pots against P3 isolate of *Phytophthora* blight; only one entry, T 7 showed a minimum of 57.4 percent blight.
2. Of the 13 species of *Atylosia* tested, only *A. goensis* and *A. platycarpa* showed low blight incidence under greenhouse conditions against P3 isolate.
3. A preliminary test indicated the existence of physiologic races in *Phytophthora drechsleri* f.sp. *cajani*.

II. Introduction

The project was formulated in 1985 with the objective to investigate the biology and epidemiology of the pathogens causing foliar blights of pigeonpea.

III. Phytophthora blight

A. Pot screening method

Pigeonpea lines were screened against the more virulent P3 isolate in pots in the greenhouse by the 'drench inoculation' method. Planting, inoculation and recording observations were done as described in Pulse Pathology (Pigeonpea) Report of Work 1977-78. But, 50 ml of the mycelial suspension was poured into each 10 cm dia plastic pots, instead of 100 ml suspension in 20 cm dia pots. The surviving plants were reinoculated by 'rub' method and then transplanted in 30 cm dia plastic pots containing Vertisol. Selfed seed are collected from the surviving plants for retesting.

1. Coordinated Varietal Trial (ACT)

Eighteen entries included in the Extra early Arhar Coordinated Trial (EXACT), nineteen in Early Arhar Coordinated Trial (EACT), seventeen in Arhar Coordinated Trial-1 (ACT-1), thirteen in ACT-2, twenty-five in ACT-3, twenty-one in Medium maturity pigeonpea sterility mosaic and wilt resistant yield trial (MPSMVRY), and three entries in late maturity pigeonpea sterility mosaic and wilt resistant yield trial (LPSMVRY) were tested against P3 isolate. All the 116 CVT entries were found susceptible to blight. Only one entry, T 7 from ACT 3 showed a minimum of 57.4 percent blight (Table 9).

Table 9. Results of pot of screening of ACT entries (1985-86) against Phytophthora blight in the greenhouse at ICRISAT Center, Patancheru.

S.No.	Entry	Total plants		Percent blight 1		
		Rep I	Rep II	Rep I	Rep II	Average
1.	AL 1	21	23	100.0	100.0	100.0
2.	AL 15	23	21	95.7	100.0	97.9
3.	AL 101	23	24	100.0	100.0	100.0
4.	DL 78-1	23	26	100.0	100.0	100.0
5.	ICPL 317	20	22	100.0	100.0	100.0
6.	ICPL 8306	19	20	100.0	100.0	100.0
7.	Ry 10	6	13	100.0	100.0	100.0
8.	H 76-11	21	18	100.0	100.0	100.0
9.	H 76-44	24	23	100.0	100.0	100.0
10.	H 76-51	22	23	100.0	91.3	95.7
11.	H 76-65	22	22	100.0	95.5	97.8
12.	H 81-1	19	22	100.0	100.0	100.0
13.	H 82-1	20	19	100.0	100.0	100.0
14.	H 82-12	20	25	100.0	100.0	100.0
15.	Pusa 85	24	22	100.0	100.0	100.0
16.	Pusa 851	24	21	100.0	100.0	100.0
17.	TAT 10	22	22	100.0	100.0	100.0
18.	TPT 11	24	24	100.0	100.0	100.0

EACT

1.	AL 13	21	25	100.0	100.0	100.0
2.	AL 56	22	23	100.0	95.7	97.9
3.	AL 57	18	23	94.4	95.7	95.1
4.	BSMR 294	-	-	-	-	-
5.	GAUT 82-53	23	25	100.0	100.0	100.0
6.	GAUT 82-55	22	22	95.5	90.9	93.2
7.	H 76-24	20	21	100.0	100.0	100.0
8.	H 80-110	21	25	100.0	100.0	100.0
9.	H 82-26	19	21	100.0	100.0	100.0
10.	Ry 11	-	-	-	-	-
11.	ICPL 151	10	15	100.0	100.0	100.0
12.	ICPL 269	17	19	100.0	100.0	100.0
13.	ICPL 317	19	16	100.0	100.0	100.0
14.	ICPL 8327	21	23	100.0	87.0	93.5
15.	MTB 6	13	17	100.0	100.0	100.0

S.No.	Entry	Total plants		Percent blight 1		
		Rep I	Rep II	Rep I	Rep II	Average
16.	MTB 10	19	24	100.0	100.0	100.0
17.	Pant A 1-1	22	20	100.0	94.5	97.3
18.	Pant A 10	20	22	100.0	100.0	100.0
19.	Pusa Sveta 2	17	19	100.0	100.0	100.0

ACT 1

1.	CORG 5	20	20	95.0	100.0	97.5
2.	DA 8	22	20	100.0	100.0	100.0
3.	ICPL 176	23	17	100.0	100.0	100.0
4.	ICPL 186	19	23	100.0	100.0	100.0
5.	ICPL 288	13	18	100.0	100.0	100.0
6.	ICPL 8308	16	18	100.0	100.0	100.0
7.	ICPL 8324	11	12	90.9	60.0	75.5
8.	ICPL 84074	-	-	-	-	-
9.	ICPL 84077	19	20	100.0	100.0	100.0
10.	Pusa 85	23	18	100.0	100.0	100.0
11.	Pant 102	20	16	100.0	100.0	100.0
12.	Pant 103	20	22	95.5	100.0	97.8
13.	Pant 104	18	19	100.0	100.0	100.0
14.	PT 14	23	23	100.0	100.0	100.0
15.	PT 20	21	9	100.0	100.0	100.0
16.	TT 5	20	19	100.0	100.0	100.0
17.	TT 6	21	22	100.0	100.0	100.0

ACT 2

1.	AGS 498	23	24	91.3	100.0	95.7
2.	AKT 1	22	22	100.0	100.0	100.0
3.	AKT 6	22	25	77.3	76.0	76.7
4.	C 11	22	19	95.5	100.0	97.8
5.	G 78-3	22	25	81.8	88.0	84.9
6.	MA 162	13	15	100.0	93.3	96.7
7.	MTB 8	21	23	100.0	95.7	97.9
8.	MTB 9	25	23	100.0	100.0	100.0
9.	MTB 11	22	22	90.9	100.0	95.5
10.	MRG 66	22	22	86.4	90.9	88.7
11.	PT 17	19	23	100.0	95.7	97.9
12.	PT 18	22	22	100.0	100.0	100.0
13.	No. 148	23	25	100.0	96.0	98.0

.No.	Entry	Total plants		Percent blight 1		
		Rep I	Rep II	Rep I	Rep II	Average
ACT 3						
1.	AGS 522	26	22	88.5	63.6	76.1
2.	Bahar	22	21	100.0	100.0	100.0
3.	DA 15	17	25	70.6	76.0	73.3
4.	ICPL 146	23	20	100.0	100.0	100.0
5.	ICPL 161	21	23	90.5	95.7	93.1
6.	ICPL 360	21	21	100.0	100.0	100.0
7.	ICPL 366	22	23	90.9	100.0	95.5
8.	KA 1	-	-	-	-	-
9.	KA 25-1	22	22	100.0	100.0	100.0
10.	KA 28	-	-	-	-	-
11.	KA 32	-	-	-	-	-
12.	MA 2	22	24	100.0	100.0	100.0
13.	MA 95-2	18	23	88.9	100.0	94.5
14.	MA 97	23	21	100.0	85.7	92.9
15.	MA 128-2	12	20	100.0	100.0	100.0
16.	MA 165	22	25	100.0	100.0	100.0
17.	MA 166	-	-	-	-	-
18.	MA 167	-	-	-	-	-
19.	PDA 1	-	-	-	-	-
20.	PDA 9	-	-	-	-	-
21.	PDA 10	22	28	77.3	57.1	67.2
22.	PDA 83-3	-	-	-	-	-
23.	PT 20	22	24	95.5	100.0	97.8
24.	T 7	24	23	58.3	56.5	57.4
25.	KA 73-1	-	-	-	-	-

NPSHRY

1.	ICPL 227	25	24	92.0	100.0	96.0
2.	ICPL 332	22	22	86.4	100.0	93.2
3.	ICPL 333	24	25	95.8	100.0	97.9
4.	ICPL 335	22	22	100.0	95.4	97.7
5.	ICPL 342	23	26	95.7	100.0	97.9
6.	ICPL 343	26	25	76.9	88.0	82.5
7.	ICPL 345	25	24	100.0	100.0	100.0
8.	ICPL 8356	23	22	100.0	100.0	100.0
9.	ICPL 8357	20	21	100.0	100.0	100.0
10.	ICPL 8358	23	21	100.0	100.0	100.0

S.No.	Entry	Total plants		Percent blight 1		
		Rep I	Rep II	Rep I	Rep II	Average
11.	ICPL 8362	20	23	100.0	97.5	98.8
12.	ICPL 8363	23	22	73.9	77.3	75.6
13.	ICPL 84001	23	24	87.0	91.7	89.4
14.	ICPL 84002	25	20	92.0	95.0	93.5
15.	ICPL 84008	24	20	100.0	100.0	100.0
16.	ICPL 84011	23	21	95.7	100.0	97.9
17.	ICPL 84016	23	22	95.7	100.0	97.9
18.	ICPL 85061	22	24	100.0	100.0	100.0
19.	ICPL 85062	25	26	100.0	100.0	100.0
20.	ICPL 85063	17	22	100.0	100.0	100.0
21.	ICPL 85064	23	18	100.0	100.0	100.0
<u>LPSHURY</u>						
1.	ICPL 84072	20	23	100.0	95.7	97.9
2.	ICP 8121	25	19	100.0	100.0	100.0
3.	ICPL 83143	20	19	90.0	78.9	84.5
<u>Susceptible checks</u>						
	HY 3C	25	24	100.0	100.0	100.0
	ICP 2376	22	19	100.0	100.0	100.0

Blight incidence was recorded 10 days after inoculation with the P₃ isolate of Phytophthora drechsleri f.sp. cajani.

2. Germplasm lines

Fifteen germplasm lines found resistant to less virulent P_2 isolate were tested against P_3 isolate; only two lines ICP 8805 and ICP 8700 showed <50 percent blight (Table 10).

3. Phytophthora blight resistant selections

Forty-eight progenies of plants resistant to the P_3 isolate in earlier pot screening were retested. Three lines, ICP 7815, ICP 8129 and APP 1384 showed <50 percent blight (Table 11).

4. Sterility mosaic promising lines

Thirteen sterility mosaic (SM) and wilt promising lines from SM and wilt nursery (RM 3C field, 1984 Kharif), two SM resistant germplasm lines, ICP 10976 and 10977, and one SM resistant selection from Badnapur, BSMR 225 were screened; all the lines were found susceptible to the P_3 isolate of blight (Table 12).

5. Phytophthora blight promising lines

A total of 79 germplasm lines that showed less than 50 percent blight in earlier pot screening against the P_3 isolate were retested; all the lines showed more than 50 percent blight (Table 13).

6. Advanced lines

The ten advanced lines tested showed more than 50 percent blight (Table 14) against P_3 isolate.

Of the 13 species of Atylosia tested, only A. goensis and A. platycarpa showed less than 20 percent blight (Table 15) against P_3 isolate.

B. Isolates from different locations

Phytophthora drechsleri f.sp. cajani was isolated from diseased pigeonpea plants collected from Pantnagar (Nainital district), Sivkasi Ka Pura (Bhind district), Haath Rathi Ka Pura (Morena district) and JNKVV Farm (Morena district). All the isolates, except the one obtained from Sivkasi Ka Pura were found to be pathogenic to HY 3C, a blight susceptible cultivar.

C. Physiologic Races

Pathogenicity of seven isolates of P. drechsleri f.sp. cajani from different locations in India was proven by drench-inoculation method. Fourteen genotypes were used. Each genotype was planted in four replications (one pot per replication) with twenty-five seeds per pot (10 cm dia plastic). The reaction of these genotypes to the seven isolates indicate the existence of physiologic races (Table 16). The test will be repeated to confirm the results.

Table 10. Results of pot screening of P2 isolate resistant
 eraplasm lines against the P₃ isolate of *Phytophthora*
drechsleri f.sp. *cajani*

No.	Entry	Total plants	Percent blight
1.	ICP 6974	392	79.6
2.	ICP 7798	22	95.5
3.	ICP 7810	24	66.7
4.	ICP 8282	25	76.0
5.	ICP 8287	20	90.0
6.	ICP 8328	22	100.0
7.	ICP 8332	24	100.0
8.	ICP 8562	20	90.0
9.	ICP 8564	19	89.5
10.	ICP 8568	22	72.7
11.	ICP 8619	23	69.6
12.	ICP 8688	24	62.5
13.	ICP 8700	24	50.0
14.	ICP 8701	25	100.0
15.	ICP 8805	20	40.0
	HY 3C (check)	24	95.8
	ICP 2376 (check)	25	100.0

Table 11. Results of repeat pot screening of *Phytophthora* blight resistant selections against the P₃ isolate of *Phytophthora drechsleri* f.sp. *cajani*.

S.No.	Entry	Total plants	Percent blight
1.	ICP 2719-1-Pot P10	25	52.0
2.	ICP 2719-1-Pot P20	34	100.0
3.	ICP 3753-Pot P10	4	100.0
4.	ICP 5097-SWP19 sel.-Pot P10	20	90.0
5.	ICP 5097-SWP19 sel.-Pot P20	135	90.4
6.	ICP 7200-1-Pot P10	6	100.0
7.	ICP 7200-1-Pot P20	17	100.0
8.	ICP 7200-1- Pot P30	26	100.0
9.	ICP 7200-3-Pot P10	29	79.3
10.	ICP 7200-3-Pot P20	8	62.5
11.	ICP 7269-Pot P10	13	100.0
12.	ICP 7269-Pot P20	24	100.0
13.	ICP 7493-Pot P10	86	100.0
14.	ICP 7493-Pot P20	21	90.5
15.	ICP 7815-Pot P10	18	88.9
16.	ICP 7815-Pot P20	13	31.8
17.	ICP 7917-Pot PB0	11	54.5
18.	ICP 7917-Pot P30	23	87.0
19.	ICP 7917-Pot P40	8	100.0
20.	ICP 8055-Pot P10	9	77.8
21.	ICP 8087-Pot P10	42	83.3
22.	ICP 8087-Pot PB0	18	100.0
23.	ICP 8087-Pot P40	27	96.3
24.	ICP 8087-Pot P50	71	97.2
25.	ICP 8087-Pot P60	16	93.8
26.	ICP 8105-Pot P10	17	82.4
27.	ICP 8105-Pot P20	20	70.0
28.	ICP 8129-Pot P10	4	25.0
29.	ICP 8216-Pot P10	8	87.5
30.	ICPL 87-Pot P10	4	100.0
31.	KPBR 80-2-Pot P10	43	88.4
32.	KPBR 80-2-Pot P20	25	76.0
33.	KPBR 80-2-Pot P30	18	61.1
34.	KPBR 80-2-1-SWP2-Pot PB0	18	72.2
35.	KPBR 80-2-1-SWP2-Pot P20	13	53.8
36.	KPBR 80-2-2-SWP1-Pot P10	369	92.9
37.	KPBR 80-2-2-SWP1-Pot P20	61	82.7
38.	KPBR 80-2-2-SWP1-Pot P30	10	60.0
39.	KPBR 80-3-SWP1-Pot P10	50	100.0
40.	KPBR 80-3-SWP1-Pot P20	28	100.0
41.	KPBR 80-3-SWP1-Pot P30	10	100.0
42.	RL 2 (KPR)-1-Pot P10	15	100.0
43.	RL 2 (KPR)-1-Pot P20	81	100.0
44.	RL 2 (KPR)-1-Pot P30	22	100.0
45.	NKR 13-ENT-Pot P10	22	100.0
46.	APP 6-B0-Pot P10	36	100.0
47.	APP 8-B-Pot P10	3	66.7
48.	APP 1384-Pot P10	2	50.0
	HY 3C (check)	24	95.8
	ICP 2376 (check)	25	100.0

Table 12. Performance of sterility mosaic promising lines against F3 isolate of Phytophthora blight under greenhouse conditions.

S.No.	Entry	Total plants	Percent blight
1.	ICP 11438-1-S10-SVB0	29	89.7
2.	ICP 11844-SVB0	29	93.1
3.	ICP 11934-SVB0	22	100.0
4.	PR 5118-2-1-S10-SVB0	22	95.5
5.	PR 5137-1-S10-SVB0	24	95.8
6.	PR 5140-1-S10-SVB0	44	100.0
7.	PR 5145-2-1-S10-SVB0	22	90.9
8.	PR 5149-1-1-S10-SVB0	30	100.0
9.	PR 5164-1-S10-SVB0	20	95.0
10.	PR 5294-1-S10-SV10	12	100.0
11.	PI 397456-1-2-1-S10-SVB0	25	100.0
12.	PI 397630-1-1-S10-SVB0	27	100.0
13.	Srilanka 477-1-S10-SVB0	22	100.0
14.	ICP 10976	10	90.0
15.	ICP 10977	23	82.6
16.	BSMR 225	21	76.2
	HY 3C (check)	15	100.0
	ICP 2376 (check)	20	100.0

Table 13. Results of repeat pot screening of Phytophthora blight promising lines against the P₃ isolate of Phytophthora drechsleri f.sp. cajani.

S.No.	Entry	Total plants	Percent blight
1.	ICP 5498	23	100.0
2.	ICP 7330	30	100.0
3.	ICP 7362	14	100.0
4.	ICP 7371	9	88.9
5.	ICP 7381	23	78.3
6.	ICP 7385	31	93.5
7.	ICP 7387	34	94.1
8.	ICP 7396	32	100.0
9.	ICP 7409	12	83.3
10.	ICP 7412	38	94.7
11.	ICP 7417	37	97.3
12.	ICP 7419	34	88.2
13.	ICP 7420	34	88.2
14.	ICP 7423	10	100.0
15.	ICP 7428	45	93.3
16.	ICP 7476	29	82.8
17.	ICP 7492	34	91.2
18.	ICP 7493	32	87.5
19.	ICP 7494	35	100.0
20.	ICP 7518	36	100.0
21.	ICP 7600	41	80.5
22.	ICP 7643	28	82.1
23.	ICP 7685	34	85.3
24.	ICP 7697	37	97.3
25.	ICP 7710	34	97.1
26.	ICP 7712	40	100.0
27.	ICP 7713	29	100.0
28.	ICP 7714	26	96.2
29.	ICP 7720	30	93.3
30.	ICP 7724	33	100.0
31.	ICP 7728	23	95.7
32.	ICP 7734	26	92.3
33.	ICP 7735	37	82.2
34.	ICP 7736	25	88.0
35.	ICP 7739	35	91.4
36.	ICP 7740	39	100.0
37.	ICP 7743	28	96.4
38.	ICP 7745	35	100.0
39.	ICP 7751	29	96.6
40.	ICP 7782	32	100.0
41.	ICP 7784	31	93.5
42.	ICP 7785	34	100.0
43.	ICP 7792	32	90.6
44.	ICP 7793	34	100.0
45.	ICP 7796	24	91.7

S.No.	Entry	Total plants	Percent blight
46.	ICP 7801	32	100.0
47.	ICP 7803	28	100.0
48.	ICP 7804	33	97.0
49.	ICP 7811	32	96.9
50.	ICP 7816	15	86.7
51.	ICP 7824	28	100.0
52.	ICP 7830	33	100.0
53.	ICP 7832	29	100.0
54.	ICP 7835	32	100.0
55.	ICP 7838	45	97.8
56.	ICP 7841	36	100.0
57.	ICP 7848	34	100.0
58.	ICP 7849	40	92.5
59.	ICP 7852	4	100.0
60.	ICP 7862	32	100.0
61.	ICP 7873	23	100.0
62.	ICP 7962	33	100.0
63.	ICP 8055	28	92.9
64.	ICP 8060	31	100.0
65.	ICP 8065	31	100.0
66.	ICP 8080	45	100.0
67.	ICP 8081	33	93.9
68.	ICP 8083	33	100.0
69.	ICP 8092	33	90.9
70.	ICP 8099	32	100.0
71.	ICP 8105	32	100.0
72.	ICP 8127	41	100.0
73.	ICP 8129	32	96.9
74.	ICP 8145	41	90.2
75.	ICP 8146	18	66.7
76.	ICP 8150	30	90.0
77.	ICP 8205	36	100.0
78.	ICP 8216	30	80.0
79.	ICP 8219	32	100.0
	NY 3C (check)	23	100.0
	ICP 2376 (check)	20	100.0

Table 14. Performance of pigeonpea advanced lines against P₃ isolate of *Phytophthora* blight under greenhouse conditions.

S.No.	Entry	Total plants		Percent blight	
1.	ICPL 87	2483		95.3	
2.	ICPL 151	1933		95.6	
3.	ICPL 8309	18		66.7	
4.	ICPL 8315	35		100.0	
5.	ICPL 8322	47		95.8	
6.	ICPL 8398	32		75.3	
7.	ICPL 84020	41		100.0	
8.	ICPL 84031	40		100.0	
9.	ICPL 84078	41		100.0	
10.	ICPM 8	38		94.5	
	HT 3C (check)	47		98.0	
	ICP 2376 (check)	44		100.0	

Table 15. Results of pot screening of *Atylosia* species against the P₃ isolate of *Phytophthora drechsleri* f.sp. *cajani*.

S.No.	<i>Atylosia</i> species	Total plants		Percent blight		
		Rep I	Rep II	Rep I	Rep II	Average
1.	<i>A. scutifolia</i> (A 68)	17	18	100.0	100.0	100.0
2.	<i>A. albicans</i> (PR 4816)	22	23	100.0	100.0	100.0
3.	<i>A. cajaniifolia</i> (PR 4876)	22	24	100.0	100.0	100.0
4.	<i>A. goensis</i> (JN 3501)	18	21	0.0	4.8	2.4
5.	<i>A. grandifolia</i> (BC 124363)	20	18	100.0	94.4	97.2
6.	<i>A. lanceolata</i> (CQ 1619)	26	22	100.0	100.0	100.0
7.	<i>A. lineata</i> (JN 3366)	22	23	100.0	95.7	97.9
8.	<i>A. mollis</i> (JN 4311)	22	20	100.0	100.0	100.0
9.	<i>A. platycarpa</i> (PR 4557)	18	23	11.1	17.4	14.3
10.	<i>A. reticulata</i> (IBS 2443)	17	24	100.0	100.0	100.0
11.	<i>A. scarabaeoides</i> (JN 4221)	25	22	100.0	95.5	97.8
12.	<i>A. sericea</i> (JN 1961)	19	26	94.7	92.3	93.5
13.	<i>A. volubilis</i> (JN 4200)	18	22	83.3	68.2	75.8
	HT 3C (check)	20	23	100.0	100.0	100.0
	ICP 2376 (check)	23	20	100.0	100.0	100.0

Table 16. Reaction of pigeonpea genotypes to seven isolates of *Phytophthora drechsleri* f.sp. *cajani*.

Genotype	P 3 (ICRISAT)	Misar (Varanasi)	BRU (Varanasi)	Kanpur	TARI (Delhi)	P 2 (ICRISAT)	BP (ICRISAT)
ICP 6997	R	S	S	S	R	S	R
NY 3C	R	S	S	S	S	S	R
ICP 7010	R	S	S	S	S	S	R
ICP 113	R	S	S	S	R	S	R
ICP 1709	R	S	S	S	R	R	R
ICP 4802	R	S	S	S	R	R	R
ICP 7657	R	S	S	S	R	R	R
ICP 782	R	S	S	S	R	R	R
ICP 2376	R	S	S	R	R	R	R
NPBR 80-1-4	R	S	S	R	R	R	R
ICP 7049	R	S	R	S	R	R	R
ICP 7269	R	S	S	S	R	R	R
ICPL 7	R	S	S	S	R	R	R
ICP 7795	R	S	R	R	R	R	R

1=Data from one test only.

R=0 to 20 percent blight - resistant.

R=21 to 50 percent blight - moderately susceptible.

R=51 to 100 percent blight - susceptible.

V. *Alternaria* blight

Alternaria tenuissima was isolated from stems of infected pigeonpeas in BW 21 field. The fungus was not pathogenic when tested under greenhouse conditions. A high concentration of spore suspension was sprayed on the test seedlings. Symptoms were not produced on ICP 7197 (Bahar) (*Alternaria* blight susceptible) and ICP 2376 (resistant).

The BHU isolate of *A. tenuissima* was multiplied in a 250 ml flask containing 100 ml potato dextrose broth (PDB) and incubated at 30°C in dark for 20 days. Very few conidia were produced. When the mycelial suspension was sprayed on the test seedlings, ICP 7197 and 2376 showed 100 and 0 percent blight, respectively. Good sporulation was observed when the fungus was multiplied in PDB and incubated at 25°C under 8 hr near-UV for 20 days.

Project: CP-119(85)IC - Studies on the viruses affecting pigeonpea

I. Summary

1. Different purification methods were used to purify and isolate the causal pathogen of SM. None of the methods worked except the one where the enzyme Driselase was incorporated in purification schedule, flexuous virus-like particles (VLP) of 12-13 nm width and 250-600 nm in length were observed. However, these VLP did not show ultraviolet absorbance at 260 nm and similar flexuous structures in very low concentration were also observed from healthy leaf tissue.
2. Ultrathin sections of infective mites and SM-infected leaves did not show virus-like particles or any viral inclusion.
3. Mechanical sap transmission of SM pathogen was not successful.
4. SM pathogen was graft transmissible to pigeonpea and bean (Phaseolus vulgaris cv. Bountiful).
5. Using its whitefly vector, Bemisia tabaci Genn. the yellow mosaic (YM) was transmitted to 4 herbaceous plant hosts, namely horsegram (Macrotyloma uniflorum), mungbean (Vigna radiata var-Pusa Baisakhi), limabean (Phaseolus lunatus cv. Henderson Baby Bush), field bean (Phaseolus vulgaris cv.Topcrop), in addition to pigeonpea.
6. Attempts in sap transmission of YM gave a low rate of success on to field bean ((Phaseolus vulgaris cv. top crop) and mungbean ((Vigna radiata cv. Pusa Baisakhi).

II. Introduction

The work on the project 'Studies on the viruses affecting pigeonpea' was started from May 1985 and this is the first year of report under this new project. Work done on sterility mosaic (SM) and yellow mosaic (YM) that infect pigeonpea is reported here.

III. Biology of pathogens

A. Sterility mosaic

We continued efforts to know the nature of the pathogen causing SM.

1. Purification procedures

a. Using protoplasts

Since it was difficult to isolate the causal pathogen of SM from leaves, we made an attempt to isolate protoplasts first, and then rupture them which may hopefully release the causal

pathogen. Because the exact nature of SM pathogen is still not known and with a purpose to have positive control for comparison, we also processed the groundnut leaf tissue infected with peanut mottle virus (PMV) and its control. As we can handle only small quantity of leaf material for protoplast isolation, we started with 30 leaflets from pigeonpea and groundnut. The following procedure for mesophyll protoplast isolation was done three times.

- (i) Leaflets were washed with 70% ethanol for 5 minutes.
- (ii) Leaflets were transferred to 0.3M clorox solution for 10 minutes.
- (iii) Leaves were washed three times with sterilized distilled water, total time 15 minutes for 3 washes.
- (iv) Leaves were dried on paper towels for 15 minutes, then floated in 1% solution of Rohment-P, an enzyme, in a petri plate, incubated at 30 C for 30 minutes.
- (v) Leaves were transferred in 0.6M sterilized Mannitol solution in a petriplate for 4 hours at room temperature, which helped in plasmolysis of mesophyll cells.
- (vi) Leaves were transferred to enzyme solution 2 (0.7% cellulase and 0.1% Macerozyme), incubated at 30°C overnight.
- (vii) The petriplates were then swirled to help in release of protoplasts from leaf tissue.
- (viii) The protoplast soup was strained with a fine nylon mesh and transferred to clinical centrifuge tubes.
- (ix) The tubes were spun in a clinical centrifuge at 400 rpm for 10 minutes. The protoplast pellets which were loose, were resuspended in 0.6 M mannitol, pooled together and centrifuged. The final pellet was resuspended in 1 ml of 0.6M mannitol.
- (x) Examined for protoplasts under a light microscope.
- (xi) Resuspended protoplast pellet low speeded at 8000 rpm for 10 min.

The protoplasts burst: A pinchful of carborundum may be added before going in for low speed centrifugation as it helps in rupture of more number of protoplasts.
- (xii) The supernatant was examined under electron microscope for virus-like particles, if any.
- (xiii) The supernatant was layered on a 10 to 40% linear

sucrose gradient and spun at 20,000 rpm, for 2 hours and examined for a opalescence band.

By using the above procedure, we noticed by light microscopy that more number of protoplasts could be isolated from groundnut than from pigeonpea, which had a poor harvest of protoplasts. However, the samples from low speed centrifugation and bands from sucrose density gradients upon examination under electron microscope did not show virus-like particles (VLP) either from pigeonpea (SM infected) or from groundnut (PMV infected). Based on these results we feel that this system may work successfully if we start with 25 g or more of leaf tissue.

b. Using Enzyme Driselase

The procedure used is a slight modification of the procedure used earlier by Takanami and Kubo (J.Gen.Viro. 1979. 44: 153-159).

Infected leaves (fresh)

Using liquid nitrogen, ground the tissue into powder; using a varying blender, further homogenized in 0.1M citrate buffer containing 0.1% sodium thioglycollate and 0.5% Driselase, pH 6.0, 3 ml/g leaf. Dispensed extract in flasks, shaken for 2 hours, expressed through cheesecloth

Extract (Infectivity assay)

Emulsified with one-third volume chloroform, stirred for 20 min, centrifuges at 5000 rpm, 15 min

Aqueous phase

Sodium chloride and polyethylene glycol 8000 were added to make final concentration of 0.2M and 6%, dissolved on a stirrer, and stored for 120 minutes in a refrigerator, centrifuged 8000 rpm, 15 min

Precipitate

Resuspended in 0.01M citrate buffer pH 6.0, stored overnight, centrifuged at 5000 rpm, 10 minutes.

Supernatant (Infectivity assay and EM examination)

30% sucrose cushion in 0.01M citrate containing NaCl (0.2M) and PEG 8000 (6%) cushion occupied 1/3 portion of tube and in rest of the tube, sample layered centrifuged at 20,000 rpm for 120 min.

Pellet

Resuspended in 0.01M citrate buffer, pH 6.0 centrifuged at 5000 rpm, 10 min.

Supernatant (Infectivity assay, EM examination and UV absorbance)

Prepared 10-40% cesium chloride gradients, centrifuged at 25,000 rpm, 1 hour in SV 41 rotor.

The fractions collected at different stages in purification schedule were used for sap inoculation, examined under electron microscope, and their ultraviolet absorbance was seen. Sap inoculation of different fractions on to pigeonpea and other herbaceous indicator hosts did not give positive results. Because earlier we did not get a band in linear sucrose gradient, we shifted to 10-40% cesium chloride gradient and got a diffused band at 2.5 cm from top of the tube. This band when seen under electron microscope showed virus like particles (VLP) of approx. dimensions 12 nm width x 250-600 nm length; similar VLP were seen earlier in PEG precipitate and in sucrose cushion (ICRISAT Annual Report 1985). However, these VLP did not show ultraviolet absorbance typical of nucleoprotein.

Similar VLP were seen in healthy extracts after cesium chloride gradient centrifugation, although their concentration was low. The VLP from infected and healthy preparations were similar.

If we omitted Driselase from the purification schedule, VLP were not seen but instead masses of broom like structures were seen under electron microscope.

C. Using enzyme Rohament-P

The procedure given below, in which Rohament-P was used, is adapted from a procedure used earlier by Cleora J.D'Arcy *et al.*, (Phytopathology 1983(73):755-759).

Infected leaves (pigeonpea)

Use liquid nitrogen, grind the tissue into powder in a mortar-pestle; using a blender, further homogenize in 0.1M sodium phosphate buffer, pH 6.0 containing 0.1% sodium thiglycollate and 1.5% Rohament-P (3ml buffer/G of leaf), dispensed the extract in flasks, shaken for 24 hours, 28°C, express through cheesecloth

Extract

Adjust the extract to pH 6.4 by adding 1N NaOH, measure volume, emulsified with one-third volume chloroform, stirred for 15 min. centrifuged at 4000 rpm 15 min.

Aqueous phase

Sodium chloride and polyethylene glycol (PEG) 8000 was added to make final concentration of 0.2M and 5%, stirred for 30 min at room temperature, stored in refrigerator for another 15 min, centrifuged at 8000 rpm, 15 min.

Precipitate (EM examination)

Resuspended in 50 ml of 0.01M sodium phosphate buffer, pH 6.5

containing 1% Triton x-100, stored overnight in refrigerator, centrifuged at 8000 rpm for 10 minutes.

Super (EM examination)

20% sucrose cushion, centrifugation at 20,000 rpm for 2 hours (the volume of sucrose cushion was 10 ml, rest of the tube filled with the sample).

Pellet (UV absorbance)

Resuspended in 0.01M sodium phosphate buffer, pH 6.5, centrifuged at 5000 rpm 10 min.

Super (EM examination and UV absorbance)

Prepared 10-40% linear sucrose gradients layer the sample, centrifuged at 20,000 rpm for 1 hour.

The pellets obtained after NaCl-PEG precipitation and sucrose cushion did not show virus-like particles (VLP) when examined under electron microscope. After sucrose density gradient centrifugation, there was no opalescent band, and ultraviolet absorbance typical of nucleoprotein was not noticed.

d) Using procedures used for other mite transmitted viruses

There are two procedures reported in literature earlier for purification of wheat streak mosaic and rye grass mosaic viruses. Both of these cereal viruses are transmitted by eriophyid mites.

Purification procedure used for wheat streak mosaic (after, Brakke, H.K. and Ball, E.M. 1968. Purification and antigenicity of wheat streak mosaic virus. Phytopathology 58:963-971).

Infected leaves (100 G)

Used a waring blender, the buffer was 0.01M K₂HPO₄, pH 8.5, 2 ml buffer/G of leaf, express sap thru cheesecloth, measure volume, adjust to pH 6.0 with 1N acetic acid, heated sap 40°C, 1 hour, kept in cold room 1 hour which helped in precipitation of host proteins.

Extract

Centrifuged the extract at 8000 rpm, 20 min

supernatant

Measured volume, its pH was raised to pH 8.0. To this sufficient sodium citrate salt was added to give a 0.01 M concentration. Used a 20% sucrose cushion in 0.01 M sodium citrate and was centrifuged at 28,000 rpm, 2 hours.

pellet

pellet was resuspended in 10 ml of 0.01 sodium citrate, pH 8.0 (1 ml/10 G of leaf), allowed to stand overnight in refrigerator, centrifuged at 8000 rpm, 20 min.

Supernatant

Two ml of super was layered on 10-40% linear sucrose gradient and spun at 23000 rpm, 2 hr. Tubes observed for light scattering.

Samples for electron microscopic examination were taken at various stages during purification and virus-like particles not seen.

Purification procedure used Rye grass mosaic virus (after, Paliwal, Y.C., and Tremaine, J.H. 1976. Multiplication, purification, and properties of ryegrass mosaic virus. *Phytopathology* 66: 406-414).

Infected leaves (Tissue from ICP 8136 served as control)

Use liquid nitrogen, ground the tissue into powder in a mortar-pestle, by using a ball-mill, further homogenize in 0.01M sodium citrate, pH 7.0, containing 0.02M 2-mercaptoethanol, 3.5 ml/G leaf tissue. In ball mill, the homogenization was for 1 hour in a walk-in cold room (purpose of using ball mill: we thought that in case the SM pathogen is flexuous rod, use of a ball mill will gradually release virus particles from leaf tissue without their breakage), express the extract through muslin cloth.

Extract

Heat the extract at 40°C for 5 minutes, cool it immediately in an ice bath for 1 hour, centrifuged at 8000 rpm for 20 min in GS-3 rotor.

super

Measured volume and sodium chloride and PEG 8000 were added to make a final concentration of 0.2M and 6%, dissolved with a magnetic stirrer for 30 min, and kept for another 90 min in a freezer (-20°C). centrifuged at 7500 rpm 15 min.

precipitate

Resuspended in 0.01M sodium citrate, pH 7.0 (in one-fourth of the original volume), halt overnight, low speed at 5000 rpm, 15 min

Supernatant

centrifuged at 25000 rpm, 2 hrs. in R-35 rotor

Pellet

Resuspended in 2 ml of 0.1M sodium citrate, pH 7.0, allowed to soak for 2 hours, low speed 5000 rpm, 15 min

super

centrifuged at 25000 rpm, 2 hours in R35 rotor

Pellet

Resuspended the pellet in 1 ml of 0.01M sodium citrate, pH 7.0, soak overnight, low speed at 5000 rpm, 15 min.

Super

At different stages during purification, the samples were examined under electron microscope for virus like particles. Virus-like particles were not seen and the ultraviolet absorbance readings at 260 nm and 280 nm were not indicative of a nucleoprotein.

2. Electron microscopy

a) Mite dip

About 500 live eriophyid mites from infected pigeonpea leaves were picked with the help of single eyelash transfer brush, deposited in a cavity slide, and later crushed these in a glass tissue homogenizer. The drop with triturates of infective mites was put on a strip of parafilm. Copper grids were put on the drop for about 5 minutes, lifted with forceps, lightly rinsed with water, and then was put into a drop of 2% uranyl acetate for a few seconds. When these grids were examined under electron microscope (EM) few elongated flexuous particles of various dimensions were seen and these were similar to the viruslike particles (VLP) that we earlier examined from infected pigeonpea leaves. Such particles were not seen when triturates of healthy mites were examined under EM. However, we could do this just once and this needs to be confirmed.

b) Ultrathin sections of mite

The mites collected from SM-infected leaves and pathogen-free mites from pigeonpea cv. ICP 8136 were used. The processing of material before it is ready for ultrathin sectioning was earlier described in Pulse Pathology Progress Report 45 (June 1984-May 1985). VLP were not identified in ultrathin sections of mites and further attempts are necessary.

c) Ultrathin sections of leaf

Infected pigeonpea leaves of cv. NP-(VR) 15 and cv BDN-1 and their healthy leaf tissue picked up from plants raised in glasshouse were used. The processing of leaf material for ultrathin sectioning was described earlier in Pulse Pathology Progress Report 45 (June 1984-May 1985).

Definite virus inclusion or any other type of inclusion was not observed in infected material.

In ultrathin sections of infected leaves of cv.BDN-1 some nuclear changes were observed in the form of increased electron density but no conclusion could be derived. No VLP was seen in ultrathin sections.

In ultrathin sections of infected leaves of cv.WP(VR)15, some membrane bound inclusions with radiating bands were found. However, enough number of ultrathin sections were not scanned to draw a definite conclusion.

3. Mechanical sap transmission

The sterility mosaic infected leaf tissue (about 20 G) was triturated in liquid nitrogen in a mortar pestle and the fine leaf powder was homogenized in 0.1M sodium citrate buffer, pH 6.0 containing 0.1% sodium thiglycollate and 0.5% Driselase, an enzyme, in a Waring blender, in ratio of 5 ml buffer/G of leaf (volume/vt). Following herbaceous plants (5 plants of each, minimum) were used in sap inoculation. Plants were earlier dusted with carborundum. Bean (Phaseolus vulgaris cv.Bountiful), Nicotiana clevelandii, N. tabacum cv. Samsun-NN, Tetragonia expansa (new Zealand spinach), limabean (Phaseolus lunatus cv.Henderson Baby Bush), and pigeonpea cv.BDN 1. Results of mechanical sap transmission onto pigeonpea or other plant sp. were negative.

4. Graft transmission

The tissue implantation method of grafting was used. In this, approx. 1 cm length stem pieces from infected pigeonpea, earlier treated with 0.3% Metasystox to kill mites, were used as scions and these were implanted by making a slit on the stem below the growing point of healthy plants (served as stock) of pigeonpea cv BDN-1, bean (Phaseolus vulgaris cv.Bountiful), limabean (Phaseolus lunatus cv.Henderson Baby Bush). At least 10 plants of each plant sp.were used and adequate controls kept.

Of the 24 pigeonpea grafted, 1 plant showed symptoms of SM and of 10 bean cv.Bountiful grafted, 4 plants showed symptoms of yellowing of veins and mosaic. In Bountiful bean, the pathogen was systemic up to second trifoliate; however, later the plants own defence system seemed to have not allowed further advance of the pathogen and symptoms disappeared in new leaves. We were not successful in graft transmission of SM in bean cv.Kentuckey, and in limabean cv.Henderson Baby Bush.

B. Yellow Mosaic

Yellow mosaic (YM) of pigeonpea is not economically important disease at the moment, but has a potential to become serious.

We studied the experimental host range of YM via its whitefly vector. Also, since YM has not been found sap transmissible, we made attempts to sap-transmit this virus to pigeonpea and to other herbaceous plant species.

1. Experimental host range of YM

The cotton whitefly, *Bemisia tabaci* Genn. was collected from a cotton field and was maintained on cotton in cages and used in experimental host range studies.

The table below shows the different herbaceous hosts used, besides pigeonpea, to know the host range of the YM (Table 17).

Whiteflies used in transmission studies were given a pre-acquisition starvation of about 1 hour; these were given acquisition access feeding for 2 days on a yellow mosaic infected pigeonpea twig which was kept in a water bottle to keep it turgid. These whiteflies were released for transmission feeding for 2 days on different test plant species. We released 50 whiteflies per plant and at least 5 plants of each host species were used except for lima bean. Among the different host species tested, *Vigna radiata* (mungbean cv. Pusa Baisakhi) was observed as a good assay host for YM.

2. Mechanical sap transmission of YM

Infected leaf tissue of YM established in mungbean var. Pusa Baisakhi via whiteflies was used in mechanical sap transmission. The leaf tissue was triturated in 0.1M potassium phosphate buffer, pH 7.8 containing 0.2% mercaptoethanol in a mortar-pestle in proportion of 1:4 (wt:vol). The sap was kept cold and was rubbed onto carborundum-dusted leaves of test plants. A minimum of 10 plants of mungbean, *Phaseolus vulgaris* cv. Topcrop, pigeonpea cv. ICP 11242 were used.

Of 24 plants of mungbean var Pusa Baisakhi sap inoculated, one plant showed yellow mosaic symptoms. Similarly of 10 plants of bean (*Phaseolus vulgaris* cv. Topcrop), 2 plants showed leaf curling and death of the plant; this symptom in Top crop bean was similar to the symptoms earlier observed after release of whiteflies.

Table 17. Experimental host range of yellow mosaic virus of pigeonpea.

S.No.	Plant species	# plants infected/ # of plants inoculated	Symptoms	Recovery Assay on pigeonpea (ICP-1) or mungbean var. Pusa Balakshi
1.	<u>Horrogram</u> (<u>Macrotyloma</u> <u>uniflorum</u>)	2/4	Yellow mosaic, systemic	Positive
2.	<u>Chickpea</u> (<u>Cicer arietinum</u>) var. <u>Glabrous</u> Mutant	0/5	-	-
3.	<u>Mungbean</u> (<u>Vigna radiata</u>) var. <u>Pusa Balakshi</u>	5/8	Yellow mosaic, systemic, sometimes downward leaf curling noticed first, followed by yellow mosaic in new leaves.	Positive
4.	<u>Groundnut</u> (<u>Arachis hypogaea</u>) var. <u>Kopergaon</u> (ICG 80366)	0/5	-	-
5.	<u>Pigeonpea</u> (<u>Cajanus cajan</u>)			
	ICP 1	3/20	Islands of intense yellow mosaic lesions on leaf lamina, sometimes yellow mosaic covers the entire leaf lamina	Positive
	ICP 11242	1/9	Similar as above	Positive
6.	<u>Rice bean</u> (<u>Vigna unguiculata</u>) var. <u>ABL 13</u>	0/6	-	-
7.	<u>Bean</u> (<u>Phaseolus vulgaris</u>)	4/8	Downward curling of leaves, new trifoliates shortened, and become brittle. Intense yellow mosaic, as seen in pigeonpea and mungbean not observed	Not done
8.	<u>Bean</u> (<u>Phaseolus vulgaris</u>) cv. <u>Plato</u>	0/5	-	-
9.	<u>Lima bean</u> (<u>Phaseolus lunatus</u>)	1/3	Intense yellow mosaic not seen; the new trifoliates instead showed reduction in size of trifoliates and mosaic mottle.	Not done
10.	<u>Squash</u> (<u>Cucurbita maxima</u>) var. <u>Black magic</u>	0/6	-	-

- = indicates no symptoms or negative results

Project: CP-120(85) IC - Screening for disease resistance in chickpea and pigeonpea

I. Summary

Screening nurseries

1. Very high, early and uniform incidence of wilt (average 98% with a range of 97-100%) was achieved in wilt sick plots by criss-cross ploughing of the plots, adding inoculum just before sowing, early sowing (in the first fortnight of June) and good weed control especially through the application of pre-emergence herbicides. This has enabled very effective screening of germplasm and breeding materials.
2. Equally high level of incidence of sterility mosaic (SM) (average 99-100%) was achieved in 2.3 ha SM nursery by following infector-hedge and leaf-stapling inoculation techniques.
3. Screening for combined resistance to wilt and SM was effectively carried out in 0.4 ha. Alfisol field.
4. Screening for combined resistance to wilt, SM and phytophthora blight (PB) was done in 1 ha Alfisol field. Screening for wilt and SM resistance was very effective but screening against PB was not effective due to low disease development.
5. The Fusarium population in all the sick plots before sowing is being estimated to understand the variation in wilt incidence within and over the seasons.

Wilt resistance

1. Three additional germplasm lines resistant to wilt were identified. These were ICP-12942, -13056, and -13165. Among 6 ICPL lines screened, ICPL-84006, -84014, and -84015 showed <10% wilt.
2. The results of screening of ACT materials against wilt showed that except some ICPL lines from MPSMWRY most other lines are susceptible. Entries of EXACT and EACT showed comparatively less wilt than the late lines.
3. A total of 444 early, 5138 medium, 90 late, and 314 other breeding materials were evaluated for wilt resistance to assist the breeders in development of wilt resistant varieties.
4. None of the 10 Mahyco lines tested showed <20% wilt.

5. For wilt, 2 multilocation nurseries were operated; IIUTPWR in India and IPVN in East Africa. In IIUTPWR, 44 lines were provided to 11 cooperators and results were received from 8. ICP-9174 showed <20% wilt at all 8 locations in India. In IPVN, 34 lines were supplied to 3 cooperators. ICP 12733, -12738, -12741, -12748, and -12753 showed <10% wilt at ICRISAT and BVUMBWE in Malawi.

SM resistance

1. Fifty two additional germplasm lines resistant to SM (OX) were identified and 17 of these had <20% wilt.
2. Among the ACT lines, in addition to ICPLs, some lines in ACT3 were also found promising against SM. Some ACT entries have been purified for SM resistance by repeated screening and selfing of resistant plants.
3. A total of 545 early, 640 medium, 35 late, and 365 other breeding materials were evaluated for SM resistance to assist the breeders in developing SM resistant cultivars.
4. Out of 10 Mahyco lines, two lines; MPPL8 and MPPL10 showed <10% SM.
5. Out of 32 *Heliothis* promising lines, four lines; PI-396986, ICP-6831, ICP-7198 and ICP 4769 showed <20% wilt and SM.
6. SPP of three early ICPLs i.e. ICPL-146, ICPL-151, and ICPL-155 were screened for SM resistance in pots. ICPL-146 showed uniform ringspot reaction. None of 310 SPP of ICPL 151 was free from SM but some isolated plants showed ringspot or mild mosaic reaction. In ICPL 155, some SPP were uniformly resistant while others were susceptible.
7. One multilocation trial; IIUTPSMR was operated in India. In this trial, 39 lines were provided to 10 cooperators. Results were received from 6 locations. ICP 7035, 7234, 8862, 10976, 10979, 10984, 11049, 11207, BSMR 225, 235, 258 and ICPL 8324 showed <20% SM at all 6 locations.

Phytophthora blight resistance

1. For phytophthora blight, one multilocation trial, IIUTPBR was operated in India. Nine lines were supplied to 5 cooperators and results from 4 locations showed that no line was promising across locations.

Multiple disease resistance

1. Two germplasm lines; PR 5149 and PI 397630 showed <10% wilt and SM.
2. 230 medium, 10 late and 25 male sterile breeding materials were evaluated for combined resistance to wilt and SM.
3. Two multilocation trials for multiple disease resistance were conducted in India for the first year. In IIUTPMNR-A, for wilt and SM resistance, 54 lines were provided to 4 cooperators. The line ICP 8869 showed <10% and ICP 8860 <20% wilt and SM across locations. In IIUTPMNR-B, for SM and PB resistance, 9 lines were provided to 3 cooperators and ICP 11304 showed promise for SM and PB at all the locations.
4. 234 early, 259 medium and 56 late breeding materials were evaluated for wilt, SM, and PB resistance in the multiple disease nursery.

Wilt and Heliothis resistance

1. Wilt resistance of ICP 4769, ICP 6831 and ICP 7198, which were known to be Heliothis and wilt resistant was confirmed.

17 P2 and P3 wilt and Heliothis tolerant populations were evaluated for wilt resistance.

II. Introduction

This project has been formulated in January 1985 with the following objectives.

To identify broad based/durable sources of resistance for wilt, sterility mosaic, Phytophthora and Alternaria blights of pigeonpea to help breeders in developing pigeonpea varieties with broad based/ durable resistance to individual and combination of diseases.

During 1985-86 the work carried out under this project consisted of creation of large scale artificial epiphytotics of wilt, sterility mosaic and phytophthora blight in the field for screening germplasm and breeding materials, identification of new sources of resistance from germplasm, screening of breeding materials and multilocation testing of resistant lines to identify lines with broad based resistance and share the seed of resistant materials with the scientists of the national programs.

III. Screening nurseries

A. Wilt sick plots

Vertisol sick plots BIL 2A and BIL 2B (1.8 ha) at ICRISAT Center were used for screening for wilt resistance. These fields are being used since 1977-78 season. In order to obtain uniform, early and high incidence of wilt and maintain high level of

sickness for the subsequent seasons the following steps were undertaken.

1. Application of wilt inoculum (chopped pigeonpea wilted plants collected from sickplots and other fields) in the field just before sowing.
2. Criss cross ploughing of sick plots for uniform distribution of inoculum.
3. Early sowing i.e. first fortnight of June to get good growth of pigeonpea.
4. Good weed control thru pre-emergence application of herbicides (0.75 kg Basalin + 1 kg prometryne per ha) and hand weeding.
5. More frequent use of susceptible check ICP 2376 i.e. 1:2 instead of 1:4.

These above steps helped in obtaining more than 98 percent average wilt incidence in the susceptible check. The wilt incidence was also very uniform (range 97.0-100.0 percent) (Table 18). The wilt incidence in the two wilt sick plots for the past 9 years is given in Table 19. The other aspect observed during the current season was that the wilt incidence almost reached peak by flowering time and it has helped in selfing of the resistant material.

From this season data on *Fusarium* population in the sick plots before sowing is being collected. For this purpose sick plots were divided into 10 x 10 m grids and from each grid 20 sub-soil samples were drawn. The data on *Fusarium* propagules and final wilt incidence is presented in table 20. In general the *Fusarium* population was low in vertisols than in Alfisols but the wilt incidence was high in both soils. The data also revealed big variation in *Fusarium* population in the sick plots even after 9 years of their development.

B. Sterility mosaic nursery

A 2.3 ha plot in vertisol field BIL 7B was used for screening against sterility mosaic (SM). Both infector-hedge and leaf stapling inoculation techniques were used for creating the artificial epiphytotic of the disease. Inoculation through the leaf stapling technique was carried out as disease incidence in the infector hedge was very low. However the combined use of both the techniques helped in getting high incidence (99.0 percent) of the disease in the susceptible check BDN-1 which was planted after every 10 test rows. The range was very also narrow (97.0-100.0 percent) indicating uniform development of disease.

Table 18. Final disease incidence of wilt, SM and PB in susceptible checks in different pigeonpea disease nurseries during 1985-86 at ICRISAT Center.

Field	Diseases	Area (ha)	Check cultivar (Susc. check)	Percent disease incidence	Range	Frequency of susc. check
BIL 2A	Wilt	0.8	ICP 2376	99	99-100	1:2
BIL 2B	Wilt	1.0	ICP 2376	98	97-99	1:2
BIL 7C	SM	2.3	BDN 1	99	97-100	1:10
RM 8E	Wilt	0.3	ICP 2376	99	99-100	1:8
RM 3C	SM	0.1	ICP 8863	100		1:8
RP 18	Wilt	1.0	ICP 2376	91	86-100	1:2
	SM		ICP 8863	93	89-92	1:8
	PB		HY 3C	53	-	1:8
BM 16C	Wilt, <i>Heliothis</i>	1.0	T 21	91	-	1:4

SM=Sterility mosaic, PB=Phytophthora blight

Table 19. Pigeonpea wilt incidence in susceptible cv. ICP 2376 in vertisol sick plots BIL 2A & 2B at ICRISAT Center (1977-1986).

Year	BIL 2A		BIL 2B	
	% wilt		% wilt	
	Early	Harvest	Early	Harvest
1977-78	2.3	71.8	1.1	61.5
1978-79	24.0	93.5	27.3	93.3
1979-80	24.2	90.9	18.0	96.4
1980-81	26.2	74.1	30.9	77.4
1981-82	41.0	82.4	22.6	66.9
1982-83	17.9	92.8	17.3	90.0
1983-84	30.3	86.6	11.9	69.0
1984-85	11.6	90.8	13.1	84.0
1985-86	-	99.0	-	98.0

Table 20. *Fusarium* population and pigeonpea wilt incidence in different wilt sick plots during 1985-86 at ICRISAT Center.

Field/Nursery	<i>Fusarium</i> population/g soil	Percent wilt at maturity
RM 3C (Wilt + SM nursery)	4985 (4820-5380)	99.5 (99.3-99.7)
RM 8E (Wilt nursery)	8350 (6500-9300)	89.4 (80.5-98.1)
RP 18 (Wilt+SM+PB nursery)	4075 (3340-5060)	91.0 (86.0-99.5)
BIL 2A (Wilt nursery)	3086 (1600-4100)	99.5 (99.2-99.7)
BIL 2B (Wilt nursery)	2633 (2100-3900)	98.2 (97.4-99.1)
BM 16C (Wilt+ <i>Heliothis</i> nursery)	3240 (2300-5500)	91.0

1=*Fusarium* count at the time of sowing (First fortnight of June).

2=Average of 4 reps.

C. Multiple disease nurseries

1. Wilt + sterility mosaic

Alfisol sick plots RM 3C (0.1 ha) and RM 8E (0.3 ha) were used for screening for combined resistance to wilt and SM. For wilt all the steps taken for wilt sick plots were followed. Wilt and SM susceptible checks (ICP 2376 and ICP 8863) were alternated after every 4 test rows. For SM inoculation in RM 3C infector hedge technique was used and in RM 8E leaf stapling technique was followed. In both the plots, both wilt and SM incidence in the susceptible checks was nearly 100.0 percent.

2. Wilt + sterility mosaic + phytophthora blight

A 1.0 ha Alfisol plot (RP-18) was used for screening for multiple disease resistance. For obtaining high and uniform incidence of wilt, the steps mentioned under wilt sickplots were followed. For SM infector hedge technique was used. For phytophthora blight, inoculation with zoospores suspension (10,000 spores/ml) coupled with perfo-irrigation in the seedling stage and stem cut inoculation and furrow-irrigation in the adult plant stage were followed. The wilt and SM incidence was quite high but phytophthora blight incidence was low.

D. Wilt + Heliothis nursery

A 1.0 ha vertisol field (BM 16C) was used for this purpose. This plot was selected as it is adjacent to the unsprayed area where high incidence of Heliothis is expected. A large amount of pigeonpea wilt chopped material was incorporated before sowing. Surprisingly a very high incidence (91 percent) of wilt was obtained in the susceptible check in the first year itself.

IV. SCREENING FOR RESISTANCE

A. Wilt

Most of the pigeonpea germplasm available with GRU, ICRISAT has already been screened for wilt resistance and several good sources of resistance were identified. At present the emphasis is on multilocation testing of the resistant lines and screening of the breeding materials. The screening for wilt resistance this season has been very effective as the susceptible check showed near 100.0 percent wilt by flowering time throughout the sick plots.

1. Germplasm selections

Thirteen single plant germplasm selections from 1984-85 screening were retested in BIL 2B. The results are presented in table 21. The plant stand was very low for many lines. Three selections showed no wilt.

2. ICPL lines

Six ICPL lines and an advanced line were retested for wilt

resistance. The results presented in table 22 show that most of these lines are promising for wilt resistance.

3. ACT materials

The entries of EXACT, EACT, ACT 1, ACT 2, ACT 3, MPSMVRY, and LPSMVRY were tested for the r reaction against wilt. All the trials were laid out in RBD with two replications. ICP 2376 was the susceptible check. In EXACT, EACT and ACT 1, the wilt reaction is from the ratooned crop. In the main crop the early lines did not show much wilt. The results presented in tables 23,24,25,26,27,28 and 29 show that only some entries in MPSVRY are resistant to wilt.

The susceptibility of entries in ACT's to wilt indicate that not much progress has been made in breeding for wilt resistance in spite of availability of good sources of resistance.

4. Breeding materials

Screening of breeding materials in early, medium and late maturity groups was the major work under this project. The list of various breeding materials screened is presented in tables 30,31,32 and 33. The detailed results have been provided to the breeders. Generally the advanced lines which are in yield trials were tested in RBD with 2 replications.

5. MAHYCO lines

Ten lines received from MAHYCO were tested for their wilt reaction (table 34). None of them were resistant.

6. Multilocation testing

Along with screening of breeding materials, multilocation testing of resistant germplasm and advanced breeding lines was the major activity under this project. Two multilocation trials were organised for wilt resistance.

a) ICAR-ICRISAT Uniform Trial for Pigeonpea Wilt Resistance (IIUTPW) 1985-86

This trial is organised in India in collaboration with Directorate of Pulse Research (DPR), (ICAR), Kanpur. The list of cooperators and locations of this nursery are given in table 35. The reaction of the entries at different locations is given in table 36. The trial was well conducted at most of the locations and effective screening was obtained. Quite a few lines such as ICP 8858, 8863, 9174, 12745, ICPL 84008 and ICPL 84013 showed broad based resistance. (resistant in 7-8 out of 8 locations).

Table 21. Reaction of of pigeonpea germplasm selections (1984-85) against wilt in vertisol sick plot (BIL 2B) during 1985-86 at ICRISAT Center.

S.No.	Pedigree	Total plants	Percent wilt
1.	ICP-12807-W10	2	50.0
2.	ICP-12814-W10	14	21.4
3.	ICP-12880-W10	25	24.0
4.	ICP-12942-W10	14	0.0
5.	ICP-12942-W20	53	0.0
6.	ICP-12973-W10	20	40.0
7.	ICP-13056-W10	2	0.0
8.	ICP-13063-W10	4	25.0
9.	ICP-13073-W10	21	9.5
10.	ICP-13146-W10	-	-
11.	ICP-13164-W10	2	100.0
12.	ICP-13165-W10	7	14.3
13.	ICP-13165-W20	1	0.0
14.	ICP-2376 (wilt susc. check)	413	88.1

Table 22. Reaction of ICPL lines to wilt in vertisol sick plot (BIL 2B) during 1985-86 at ICRISAT Center.

S.No.	Pedigree	Total plants	Percent wilt
1.	ICPL 84005	99	18.2
2.	ICPL 84006	95	1.0
3.	ICPL 84009	100	14.0
4.	ICPL 84012	130	24.6
5.	ICPL 84014	123	9.8
6.	ICPL 84015	81	4.9
7.	ICPX 78153-W27-WB-WB0	102	7.8
8.	ICP 2376 wilt susc. check	549	92.0

Table 23. Reaction of EXACT entries to wilt in sick plot (BIL 2B) during 1985-86 at ICRISAT Center.

S.No.	Entry	Total plants		Percent wilt in ratoon crop		Mean
		R I	R II	R I	R II	
1.	AL 1	71	55	88.7	89.1	88.9
2.	AL 15	82	90	95.1	91.1	93.1
3.	AL 101	56	91	92.8	74.7	83.6
4.	DL 78-1	57	147	84.2	90.5	87.3
5.	ICPL 317	80	23	88.8	69.6	79.2
6.	ICPL 8306	81	90	90.1	83.3	86.7
7.	Hy 10	5	6	100.0	100.0	100.0
8.	H 76-11	92	140	100.0	88.6	94.3
9.	H 76-44	76	65	76.3	83.1	79.7
10.	H 76-51	132	60	93.2	93.3	93.2
11.	H 76-65	68	190	92.6	88.9	90.7
12.	H 81-1	92	84	94.6	54.8	74.7
13.	H 82-1	55	145	96.4	45.5	71.0
14.	H 82-12	66	92	83.3	97.8	90.5
15.	Pusa 85	88	82	80.8	78.0	83.9
16.	Pusa 851	72	94	81.9	88.3	85.1
17.	TAT 10	75	78	89.3	89.7	89.5
18.	TAT 11	18	22	94.4	95.4	94.9
19.	ICP 2376 (wilt check)	33	38	100.0	99.4	99.7

Table 24. Reaction of EACT entries to wilt in sick plot (BIL 2B) during 1985-86 at ICRISAT Center.

S.No.	Entry	Total plants		Percent wilt in ratoon crop		Mean
		R I	R II	R I	R II	
1.	AL 13	42	54	92.8	100.0	96.4
2.	AL 56	85	90	89.4	94.4	91.9
3.	AL 57	115	170	98.2	97.0	97.6
4.	BSMR 294	20	9	65.0	55.5	60.2
5.	GAUT 82-53	33	93	90.9	96.7	93.8
6.	GAUT 82-55	64	88	45.3	71.5	58.4
7.	H 76-24	85	93	98.8	94.6	96.7
8.	H 80-110	57	216	84.2	89.3	86.8
9.	H 82-26	98	89	93.8	93.2	93.5
10.	Hy 11	7	5	71.4	80.0	75.7
11.	ICPL 151	71	60	98.5	95.0	96.8
12.	ICPL 269	70	73	91.4	95.8	93.6
13.	ICPL 317	39	86	92.3	88.3	90.3
14.	ICPL 8327	57	79	67.2	93.6	80.4
15.	MTH 6	59	52	52.5	50.0	51.2
16.	MTH 10	42	92	95.2	95.6	95.4
17.	Pant A 1-1	61	114	93.4	96.4	94.9
18.	Pant A 10	63	78	92.0	89.7	90.8
19.	PUSA SWETA 2	77	88	85.7	87.5	86.6
	ICP 2376	67	70	100.0	99.8	99.9
	(wilt check)					

Table 25. Reaction of ACT1 entries to wilt in sick plot (SIL 2B) during 1985-86 at ICRISAT Center.

S.No.	Entry	Total plants		Percent wilt in ratoon crop		Mean
		R I	R II	R I	R II	
1.	CORG 5	100	100	98.0	91.0	94.5
2.	DA 8	31	68	90.3	00.0	95.1
3.	ICPL 176	26	48	92.3	97.9	95.1
4.	ICPL 186	62	68	91.9	61.7	76.8
5.	ICPL 288	61	101	67.2	42.5	54.9
6.	ICPL 8308	41	21	78.0	76.1	77.0
7.	ICPL 8324	12	19	33.3	47.3	40.3
8.	ICPL 84074	19	13	89.4	92.3	90.8
9.	ICPL 84077	66	83	83.3	89.1	86.2
10.	Pusa 85	52	25	94.2	92.0	93.1
11.	Pant 102	98	78	81.6	97.4	89.5
12.	Pant 103	63	75	87.3	100.0	93.6
13.	Pant 104	80	72	85.0	91.6	88.3
14.	PT 14	67	74	76.1	90.5	83.3
15.	PT 20	79	82	56.9	63.4	60.1
16.	TT 5	77	86	94.8	95.3	95.0
17.	TT 6	74	90	90.5	90.0	90.2
	ICP 2376	34	30	99.4	99.7	99.5
	(wilt check)					

Table 26. Reaction of ACT2 entries to wilt in sick plot (SIL 2A) during 1985-86 at ICRISAT Center.

S.No.	Entry	Total plants		Percent wilt		Mean
		R I	R II	R I	R II	
1.	AGE 498	52	74	75.0	64.8	69.9
2.	AKT 1	74	70	79.7	88.5	84.1
3.	AKT 6	84	140	97.6	96.4	97.0
4.	C 11	44	28	100.0	96.4	98.2
5.	G 78-3	72	39	100.0	100.0	100.0
6.	MA 162	39	28	94.8	92.8	93.8
7.	MTH 8	46	53	100.0	100.0	100.0
8.	MTH 9	74	80	93.2	91.2	92.2
9.	MTH 11	108	78	98.1	94.8	96.4
10.	MHG 66	77	95	100.0	98.9	99.4
11.	PT 17	76	115	98.6	95.6	97.1
12.	PT 18	64	46	96.8	93.4	95.1
13.	No. 148	31	60	96.7	96.6	96.6
	ICP 2376	105	84	99.9	100.0	99.9
	(wilt check)					

Table 27. Reaction of ACT3 entries to wilt in sick plot (BIL 2A) during 1985-86 at ICRISAT Center.

S.No.	Entry	Total plants		Percent wilt		Mean
		R I	R II	R I	R II	
1.	AGS 522	57	95	87.7	98.9	93.1
2.	Bahar	28	61	92.8	100.0	96.4
3.	DA 15	18	18	77.7	88.8	83.2
4.	ICPL 146	90	105	75.5	90.4	83.0
5.	ICPL 161	71	72	85.9	93.0	89.4
6.	ICPL 360	74	71	90.5	84.5	87.5
7.	ICPL 366	75	51	76.0	90.0	83.0
8.	KA 1	15	19	86.6	84.2	85.4
9.	KA 25-1	41	47	60.9	87.2	74.0
10.	KA 28	12	20	66.6	85.0	75.8
11.	KA 32	6	11	83.3	81.8	82.5
12.	MA 2	70	94	75.7	68.0	71.9
13.	MA 95-2	46	76	78.2	81.5	79.5
14.	MA 97	55	66	90.9	66.6	78.8
15.	MA 128-2	10	13	60.0	84.6	72.3
16.	MA 165	11	7	63.6	57.1	60.3
17.	MA 166	9	14	55.5	57.1	56.3
18.	MA 167	31	30	80.6	86.6	83.6
19.	PDA 1	8	5	75.0	80.0	77.5
20.	PDA 9	15	11	93.3	100.0	96.6
21.	PDA 10	44	57	100.0	91.2	95.6
22.	PDA 83-3	16	25	87.5	92.0	89.8
23.	PT 20	77	45	87.0	95.5	91.2
24.	T 7	22	32	72.7	81.2	77.0
25.	KA 73-1	10	12	20.0	25.0	22.5
	ICP 2376	119	105	97.3	99.4	98.3
	(wilt check)					

Table 28. Reaction of NPSMVRY entries to wilt in sick plot (BIL 2A) during 1985-86 at ICRISAT Center.

S.No.	Entry	Total plants		Percent wilt		Mean
		R I	R II	R I	R II	
1.	ICPL 227	77	76	2.6	13.3	7.9
2.	ICPL 332	77	127	100.0	100.0	100.0
3.	ICPL 333	45	72	0.0	25.0	12.5
4.	ICPL 335	61	62	16.4	25.8	21.1
5.	ICPL 342	68	96	82.3	89.5	85.9
6.	ICPL 343	59	70	100.0	98.5	99.2
7.	ICPL 345	72	110	70.8	77.2	74.0
8.	ICPL 8356	49	96	14.3	5.2	9.8
9.	ICPL 8357	74	185	10.8	20.5	15.7
10.	ICPL 8358	61	101	1.6	8.9	5.2
11.	ICPL 8362	96	70	13.5	15.7	14.6
12.	ICPL 8363	77	86	2.6	12.7	7.7
13.	ICPL 84001	27	43	44.4	51.1	47.8
14.	ICPL 84002	90	76	27.7	19.7	23.7
15.	ICPL 84008	82	65	11.0	9.2	10.1
16.	ICPL 84011	78	75	15.4	16.0	15.7
17.	ICPL 84016	37	61	13.5	4.9	9.2
18.	ICPL 85061	63	60	12.7	15.0	13.1
19.	ICPL 85062	58	84	12.1	20.2	16.1
20.	ICPL 85063	54	64	90.7	95.3	93.0
21.	ICPL 85064	40	96	67.5	26.0	46.1
	ICP 2376	99	105	99.1	99.7	99.4
	(wilt check)					

Table 29. Reaction of LPSMWRY entries to wilt in sick plot (BIL 2A) during 1985-86 at ICRISAT Center.

S.No.	Entry	Total plants		Percent wilt		Mean
		R I	R II	R I	R II	
22.	ICPL 84072	48	130	39.5	33.8	36.7
23.	ICP 8121	75	127	96.0	92.1	94.0
24.	ICPL 83143	68	115	60.2	69.5	64.9
	ICP 2376 (wilt check)	99	105	99.9	99.7	99.8

Table 30. List of early pigeonpea breeding materials screened for wilt resistance in sick plot (BIL 2B) at ICRISAT Center during 1985-86.

S.No.	Material	Entries Reps.	
1.	Multilocation trial entries	108	2
2.	Retesting and purification of ICPL's	19	2
3.	F2 bulks	3	-
4.	F3 populations	7	-
5.	Station trial entries	292	1
6.	Vegetable type F3 bulks	10	2
7.	SPVYT entries	5	2
	TOTAL	444	

Table 31. List of medium pigeonpea breeding materials screened for wilt resistance in sick plots (BIL 2A) at ICRISAT Center during 1985-86.

S.No.	Material	Entries
1.	MPSMWRY entries	9
2.	LRG 30 BC1F4 spp.	17
3.	F4 sps (inheritance study)	300
4.	F7 sps	258
5.	F7 spp	183
6.	F3, F4 and F6 bulks	4
7.	Advanced lines from trials	287
8.	LRG 30 M2 materials	4080
	TOTAL	5138

Table 32. List of late pigeonpea breeding materials screened for wilt resistance in sick plot (BIL 2B) at ICRISAT Center during 1985-86.

S.No.	Material	Entries
1.	SPS from advanced lines (ICPL and ICP)	43
2.	Advanced lines	45
3.	F5 and F6 bulks	2
	TOTAL	90

Table 33. List of male sterile, hybrid and high protein lines screened against wilt during 1985-86 (BIL 2A) at ICRISAT Center.

S.No.	Material	Entries
1.	MS 3783 BC2F4	23
2.	MS 3783 BC5 F1/BC5 F2	8
3.	MS 7086 BC5 F2	6
4.	WR MS 3A	7
5.	WR MS 4A	47
6.	MS BDN-1, C 11 sibs and MS BDN-1, selfed progenies	102
7.	Wilt resistant hybrids	30
8.	Yield test lines	27
9.	High protein genetic stocks	64
	TOTAL	314

Table 34. Performance of Mahyco Pigeonpea lines against wilt in vertisol sick plot (BIL 2B) during 1985-86 at ICRISAT Center.

S.No.	Pedigree	Total plants	Percent wilt
1.	MPPL-1	5	40.0
2.	MPPL-2	4	75.0
3.	MPPL-3	13	69.2
4.	MPPL-4	14	92.9
5.	MPPL-5	9	22.2
6.	MPPL-6	2	100.0
7.	MPPL-7	17	47.1
8.	MPPL-8	138	30.8
9.	MPPL-9	9	77.8
10.	MPPL-10	7	28.6
	ICP 2376 (wilt susc. check)	306	91.0

Table 35. List of cooperators and locations of IIUTPVR 1985-86.

1. Dr. T.B. Anilkumar, (Bangalore), Annigeri
2. Dr. K.K. Zote, Badnapur
3. Dr. S.W. Pillai, Baroda
4. Dr. B. Misra, Dholi
5. Dr. V.B. Bidari, Gulbarga
6. Dr. K.S. Amin, DPR, Kanpur
7. Dr. Mahendra Pal, IARI, New Delhi
8. Dr. G. Arjunan, Pudukottai
9. Dr. D.K. Jha, Ranchi
10. Mr. N.J. Bindre, Rahuri
11. Dr. S.C. Agarwal, Sehore
12. Dr. M.V. Reddy and Dr. Y.L. Nene, ICRISAT, Patancheru

b) International Pigeonpea Wilt Nursery (IPWN) 1985-86

This nursery is conducted in India, Kenya, and Malawi. The cooperators and locations are given in table 37. The reaction of the entries at ICRISAT and Malawi is given in table 38. Many lines showed higher wilt incidence in Malawi than at ICRISAT. The lines ICP 12733, ICP 12738, ICP 12741, ICP 12750, ICP 12753 and ICP 12755 showed less than 10% wilt at both the locations.

B. STERILITY MOSAIC

As in case of wilt, screening for SM resistance was very effective. In SM screening nursery very high natural incidence of wilt also developed enabling evaluation of all the materials both for SM and wilt resistance.

1. Germplasm

Two hundred and thirty eight new germplasm accessions from GRU mostly originating from Kenya were screened for SM resistance. Of these thirty seven lines were free from SM infection. These were ICP 12768, 12782, 12807, 12808, 12819, 12885, 12898, 12904, 12923, 12928, 12936, 12942, 12971, 12972, 12973, 13004, 13027, 13030, 13031, 13035, 13046, 13051, 13069, 13070, 13071, 13072, 13073, 13076, 13087, 13108, 13129, 13142, 13152, 13153, 13164, 13166, and 13169.

Several other lines showed less than 10% SM infection. It is interesting to see such high frequency of resistance from the germplasm originating from Kenya where SM incidence is not reported. Most of these lines were very late, tall and less branching types with large, thick, and dark green leaves. It will be interesting to study whether the thick leaves have any association with SM resistance.

Seventeen of the 37 lines which showed 0.0 percent SM had less than 20.0 percent wilt. These were ICP 12807, 12808, 12973, 13046, 13051, 13069, 13070, 13071, 13073, 13076, 13087, 13108, 13142, 13152, 13153, 13164 and 13166.

Most of the exotic germplasm lines showed high susceptibility to bacterial leaf spot and stem canker while indigenous lines such as BDN-1 showed high level of tolerance. Compared to green stem types, purple stem types were more tolerant.

2. Germplasm selections

Thirty eight single plant selections made from twentyone germplasm lines during 1983-84 season were retested. The data presented in table 39 show that 15 accessions had no SM. The seed of these lines has been stored in the cold room and is available for cooperators.

3. ACT materials

The reactions of EXACT, EACT, ACT-1, ACT-2, ACT-3, NPSMVRY, and LPSMVRY entries to SM are given in tables 40,41,42,43,44,45 and 46. In addition to ICPL's, some other lines in ACT 3 showed resistance to SM. These were Bahar, DA 15, ICPL 146, ICPL 366, KA 1, KA 32, MA 97, MA 166, MA 167, PDA 10 and KA 73 1.

4. ACT selections

The seed of resistant plants selected from 1983 ACT entries were retested. The data presented in table 47 show that most of these are resistant to SM. However, most of them showed susceptibility to wilt except ICPL 311.

5. Breeding material

The list of early, medium, late and other breeding materials screened for SM resistance is given in table 48. The detailed data has been provided to the breeders.

6. MAHYCO lines

Ten lines received from MAHYCO were tested for their reaction to sterility mosaic (table 49). MPPL-8 and 10 showed promise against SM.

7. Heliothis tolerant lines

Thirty two lines that showed promise against Heliothis and SM in the previous seasons were retested. Most of these lines were promising against SM. Few lines; PI 396986, ICP 4769, ICP 6831, ICP 7198, also showed promise against wilt (Table 50).

8. Pot screening of early ICPL's

Three hundred and ten SPP of ICPL 151, and 100 SPP of each of ICPL 155 and ICPL 146 were screened against SM in pots. BDN-1 and ICP 7867 were used as susceptible and resistant checks, respectively. Each SPP was planted in one 20 cm diameter earthen pot (10 plants). The susceptible check BDN1 showed 100.0 percent SM while ICP 7867 remained free.

In case of ICPL 151, all the 310 SPP showed 100.0 percent infection. But in some SPP there were a few plants with mild mosaic or ring spot symptoms. Selfed seed of these plants was handed over to the breeders. In ICPL 146, all the 100 SPP showed uniformly ring spot symptoms. In ICPL 155, while most of the progenies showed 100.0 percent susceptibility, 8 progenies showed 0.0 to 11.0 percent infection. These were ICPL 155-26, -27, -29, -74, -75, -76, -77 and -78. ICPL 155-97 showed uniformly mild mosaic symptoms.

**Table 37. List of cooperators and locations of
IPWN 1985-86.**

1. Dr. V.V. Saka
Bunda College of Agriculture
University of Malawi
P.O. Box 219
Lilongwe
Malawi
2. Dr. Andrew T. Daudi
Plant Nematologist
Byumbwe Agricultural Research Station
P.O. Box 5748
Limbe
Malawi
3. Dr. Abdul Sakoor
Plant Breeder
Katumani Dryland Farming Research
and Development Project
P.O. Box 340
Machakos
Kenya
4. Dr. M.V. Reddy, and Dr. Y.L. Nene, ICRISAT,
Patancheru, India

Table 38. Reaction of IPUN 1985-86 entries to wilt at ICRISAT, Hyderabad and Bvumbwe, Malawi during 1985-86.

Cultivar	Percent wilt	
	ICRISAT	Malawi
ICP 3461	0.0	65.5
ICP 3465	4.6	52.3
ICP 6997	88.2	100.0
ICP 8858	4.3	54.8
ICP 8859	8.7	57.7
ICP 8861	4.3	73.9
ICP 8862	88.1	100.0
ICP 8863	3.1	35.6
ICP 8864	10.5	95.0
ICP 9174	1.0	55.0
ICP 10958	14.4	71.3
ICP 11294	10.2	100.0
ICP 11295	4.5	16.7
ICP 11297	50.0	58.3
ICP 12725	11.1	73.3
ICP 12726	1.4	41.7
ICP 12727	2.1	37.3
ICP 12729	4.1	59.5
ICP 12730	15.8	27.5
ICP 12731	68.8	16.7
ICP 12733	4.3	9.1
ICP 12738	3.6	4.8
ICP 12741	3.3	2.4
ICP 12744	2.5	47.6
ICP 12745	1.0	32.1
ICP 12746	56.8	100.0
ICP 12748	3.4	8.9
ICP 12750	17.1	25.5
ICP 12752	2.2	100.0
ICP 12753	2.2	6.3
ICP 12755	3.0	15.3
ICP 12758	2.1	16.8
ICP 12759	6.0	65.9
ICPL 8343	54.0	97.1
ICP 2376	86.3	100.0
(wilt susc. check)	(69.9-98.1)	

Table 39. Reaction of 1983-84 sterility mosaic promising germplasm selections to sterility mosaic and wilt during 1985-86 at ICRISAT Center.

S.No.	Pedigree	Total plants	% SM	% Wilt
1.	ICP-6005 1 1 S10	1	0.0	0.0
2.	ICP-6005-1-1 S20	2	0.0	0.0
3.	ICP-6005 3 1 S10	37	5.4	91.8
4.	ICP-6005 3 1 S20	6	0.0	100.0
5.	ICP-6006 1-S10	37	67.5	94.5
6.	ICP-6006 1-S20	51	41.7	70.5
7.	ICP-6008-1-S10	1	100.0	-
8.	ICP-6008 1-S20	1	0.0	0.0
9.	ICP-6013 1-S10	17	0.0	29.4
10.	ICP-6013 1-S20	19	5.6	26.3
11.	ICP-6017 1-S10	14	0.0	7.1
12.	ICP-6017 1-S20	5	0.0	0.0
13.	ICP-6018 1-S10	25	0.0	72.0
14.	ICP-6018-1-S20	5	0.0	80.0
15.	ICP-6036-1-S10	20	5.0	85.0
16.	ICP-6036-1-S20	13	38.5	46.1
17.	PR 6037 1 1 S10			-
18.	PR 6061 1 1 S10	4	0.0	0.0
19.	PR 1680-1-S10	50	2.0	84.0
20.	PR 1680-1-S20	16	0.0	50.0
21.	PR 6119-1-S10	69	0.0	30.4
22.	PR 6119-1-S20	39	56.1	69.2
23.	PR 6120-1-S10	43	0.0	88.3
24.	PR 6120-1-S20	31	9.7	96.7
25.	PR 6121-1-S10	59	1.7	38.9
26.	PR 6121-1-S20	54	9.3	48.1
27.	PR 6140-1-1-S10	7	0.0	14.2
28.	PR 5354-B-1-S10	43	7.0	30.2
29.	PR 5354-B-1-S20	52	0.0	34.6
30.	PR 5364-1-S10	21	19.0	71.3
31.	PR 5370-1-S10	25	0.0	56.0
32.	PR 5370-1-S20	38	0.0	23.6
33.	PR 5322-2-1-S10	5	0.0	40.0
34.	PR 5509-1-S10	51	0.0	74.5
35.	PR 5509-1-S20	15	0.0	40.0
36.	PR 5564-1-S10	38	2.6	31.5
37.	PR 6567-1-S10	31	16.1	61.2
38.	PR 6567-1-S20	3	0.0	100.0
39.	MON-1 (SM susc. check)	40	100.0	100.0

In the SM nursery high incidence of wilt was observed and reaction of the lines to both diseases was studied.

Table 40. Reaction of EXACT entries to SM at ICRISAT Center during 1985-86.

S.No.	Entry	Total plants		Percent SM		Mean
		R I	R II	R I	R II	
1.	AL 1	48	37	100.0	100.0	100.0
2.	AL 15	47	72	100.0	97.2	98.6
3.	AL 101	89	70	98.8	98.5	98.6
4.	DL 78-1	53	46	100.0	100.0	100.0
5.	ICPL 317	69	14	98.5	100.0	99.2
6.	ICPL 8306	42	74	100.0	98.6	99.3
7.	Hy 10	13	2	100.0	100.0	100.0
8.	H 76-11	65	77	100.0	100.0	100.0
9.	H 76-44	122	74	100.0	100.0	100.0
10.	H 76-51	77	60	100.0	100.0	100.0
11.	H 76-65	97	72	100.0	100.0	100.0
12.	H 81-1	47	106	97.8	100.0	98.5
13.	H 82-1	78	79	100.0	100.0	100.0
14.	H 82-12	93	124	100.0	100.0	100.0
15.	Pusa 85	68	84	95.5	92.8	94.1
16.	Pusa 851	90	80	100.0	100.0	100.0
17.	TAT 10	45	77	97.7	100.0	98.5
18.	TAT 11	21	20	100.0	100.0	100.0
	BDN 1	79	67	95.5	97.0	96.0
	(SM check)					

Table 41. Reaction of EACT entries to SM at ICRISAT Center during 1985-86.

S.No.	Entry	Total plants		Percent disease		Mean
		R I	R II	R I	R II	
1.	AL 13	77	57	100.0	100.0	100.0
2.	AL 56	74	96	97.2	97.9	97.5
3.	AL 57	107	98	100.0	100.0	100.0
4.	BSMR 294	23	22	0.0	0.0	0.0
5.	GAUT 82-53	24	70	100.0	100.0	100.0
6.	GAUT 82-55	59	59	100.0	100.0	100.0
7.	H 76-24	86	56	95.3	86.1	90.7
8.	H 80-110	126	85	94.4	100.0	97.2
9.	H 82-26	102	91	100.0	98.9	99.4
10.	Hy 11	12	8	66.6	100.0	83.3
11.	ICPL 151	43	31	83.7	100.0	91.9
12.	ICPL 269	63	45	19.0	28.8	23.9
13.	ICPL 317	63	49	100.0	100.0	100.0
14.	ICPL 8327	77	80	62.3	65.0	63.6
15.	MTH 6	46	87	84.7	89.6	87.1
16.	MTH 10	62	30	100.0	100.0	100.0
17.	Pant A 1-1	54	51	92.5	94.1	93.3
18.	Pant A 10	43	29	88.3	100.0	94.1
19.	Pusa Sveta 2	37	54	100.0	96.2	98.1
	BDN 1	44	56	100.0	100.0	100.0
	(SM check)					

Table 42. Reaction of ACT I entries to SM at YCRISAT Center during 1985-86.

S.No.	Entry	Total plants		Percent disease		Mean
		R I	R II	R I	R II	
1.	CORG 5	65	93	95.3	93.5	94.4
2.	DA 8	63	62	85.7	69.3	77.5
3.	ICPL 176	56	46	0.0	0.0	0.0
4.	ICPL 186	47	84	87.2	95.2	91.2
5.	ICPL 288	57	64	0.0	0.0	0.0
6.	ICPL 8308	38	42	0.0	4.7	2.3
7.	ICPL 8324	19	21	0.0	9.5	4.8
8.	ICPL 84074	20	15	0.0	0.0	0.0
9.	ICPL 84077	86	53	2.3	0.0	1.1
10.	Pusa 85	24	39	87.5	87.1	87.3
11.	Pant 102	45	87	93.3	86.2	89.8
12.	Pant 103	75	101	94.6	97.0	95.8
13.	Pant 104	89	75	91.0	90.6	90.8
14.	PT 14	37	92	59.4	83.6	71.5
15.	PT 20	52	54	80.7	48.1	64.4
16.	TT 5	89	103	97.7	95.1	96.4
17.	TT 6	58	85	86.2	94.0	90.1
	BDN 1	14	30	92.8	90.0	91.4

Table 43. Reaction of ACT 2 entries to SM at ICRISAT Center during 1985-86.

S.No.	Entry	Total plants		Percent disease		Mean
		R I	R II	R I	R II	
1.	AGS 498	52	62	100.0	100.0	100.0
2.	AKT 1	72	59	100.0	100.0	100.0
3.	AKT 6	84	128	100.0	100.0	100.0
4.	C 11	42	41	100.0	100.0	100.0
5.	G 78-3	84	93	100.0	100.0	100.0
6.	MA 162	30	38	86.6	71.0	78.8
7.	MTH 8	72	87	100.0	100.0	100.0
8.	MTH 9	66	102	100.0	100.0	100.0
9.	MTH 11	108	78	100.0	100.0	100.0
10.	MRG 66	102	122	100.0	100.0	100.0
11.	PT 17	63	114	92.6	92.9	92.8
12.	PT 18	73	50	100.0	98.0	99.0
13.	No. 148	73	53	100.0	100.0	100.0
	BDN-1	54	53	100.0	100.0	100.0
	(SM check)					

Table 44 Reaction of ACT 3 entries to SM at ICRISAT Center during 1985-86.

S.No.	Entry	Total plants		Percent disease		Mean
		R I	R II	R I	R II	
1.	AGS 522	49	66	100.0	98.4	99.2
2.	Bahar	39	54	0.0	0.0	0.0
3.	DA 15	20	32	0.0	3.1	1.5
4.	ICPL 146	63	104	1.5	0.9	1.2
5.	ICPL 161	47	94	87.2	91.4	89.3
6.	ICPL 360	61	72	100.0	97.2	98.6
7.	ICPL 366	71	83	2.8	3.6	3.2
8.	KA 1	12	11	8.3	9.0	8.7
9.	KA 25-1	36	29	88.8	86.2	87.5
10.	KA 28	30	52	93.3	96.1	94.7
11.	KA 32	12	18	0.0	5.5	2.7
12.	MA 2	58	61	55.1	53.6	54.6
13.	MA 95-2	55	60	29.0	25.0	27.0
14.	MA 97	18	21	5.5	9.5	7.5
15.	MA 128-2	24	48	20.8	10.4	15.6
16.	MA 165	35	33	60.0	42.4	51.2
17.	MA 166	26	14	3.8	0.0	1.9
18.	MA 167	78	64	6.4	7.8	7.1
19.	PDA 1	13	17	46.1	11.7	28.9
20.	PDA 9	45	37	31.1	54.0	42.5
21.	PDA 10	34	37	2.9	2.7	2.8
22.	PDA 83-3	25	22	88.0	72.7	80.3
23.	PT 20	71	64	28.9	73.4	51.1
24.	T 7	36	35	72.2	65.7	69.0
25.	KA 73-1	16	20	0.0	15.0	7.5
	BDN 1			92.8	90.0	91.4
	(SH check)					

Table 45. Reaction of HPSWVKY entries to SM at ICRISA Center during 1985-86.

S.No.	Entry	Total plants		Percent disease		Mean
		R I	R II	R I	R II	
1.	ICPL 227	63	76	0.0	6.5	3.2
2.	ICPL 332	103	108	100.0	100.0	100.
3.	ICPL 333	73	70	100.0	100.0	100.
4.	ICPL 335	35	66	0.0	0.0	0.
5.	ICPL 342	68	86	0.0	0.0	0.
6.	ICPL 343	86	102	0.0	0.0	0.
7.	ICPL 345	111	94	0.0	0.0	0.
8.	ICPL 8356	58	83	100.0	100.0	100.
9.	ICPL 8357	107	112	100.0	100.0	100.
10.	ICPL 8358	89	39	100.0	100.0	100.
11.	ICPL 8362	79	82	0.0	0.0	0.
12.	ICPL 8363	87	69	2.2	1.4	1.
13.	ICPL 84001	62	80	100.0	100.0	100.
14.	ICPL 84002	73	78	98.6	96.1	97.
15.	ICPL 84008	44	98	100.0	98.9	99.
16.	ICPL 84011	66	61	7.6	29.5	18.
17.	ICPL 84016	93	91	100.0	100.0	100.
18.	ICPL 85061	133	52	100.0	100.0	100.
19.	ICPL 85062	73	92	100.0	100.0	100.
20.	ICPL 85063	94	99	1.0	0.0	0.
21.	ICPL 85064	79	60	1.2	0.0	0.6
	BDN-1	84	70	98.8	97.1	98.
	(SM check)					

Table 46. Reaction of LPSMVR entries to SM at ICRISAT Center during 1985-86.

S.No.	Entry	Total plants		Percent disease		Mean
		R I	R II	R I	R II	
1.	ICPL 84072	75	-	13.3	-	13.3
2.	ICP 8121	72	106	5.5	0.0	2.7
3.	ICPL 83143	108	88	43.5	36.3	39.9
	BDN 1 (SM check)	84	70	98.8	97.1	98.0

Table 47. Reaction of ACT-83 selections to sterility mosaic and wilt (BIL 78) during 1985-86 at ICRISAT Center.

S. Pedigree No.	R I			R II			Average	
	Total plants	% SM	% wilt	Total plants	% SM	% wilt	% SM	% wilt
1. MA-97(BHU)-1-SB0-SM	28	0.0	86.8	30	0.0	46.6	0.0	66.7
2. PDA-2-S10	42	0.0	97.6	27	0.0	55.5	0.0	76.5
3. PDA-7-S10	37	2.7	43.2	20	0.0	85.0	1.3	64.1
4. MA-2-S10	47	0.0	51.0	36	5.5	72.2	2.7	61.6
5. MA-97	30	0.0	93.3	24	0.0	87.5	0.0	90.4
6. DA-15	45	0.0	15.5	44	0.0	47.7	0.0	31.6
7. ICPL-311	38	0.0	2.6	39	0.0	10.2	0.0	6.4
8. PDA-3	32	0.0	96.8	28	0.0	75.0	0.0	85.9
9. DA-13	33	0.0	100.0	31	0.0	90.3	0.0	95.1
10. ICPL-358	36	0.0	66.6	27	0.0	48.1	0.0	57.3
11. MA-95-2-S10	35	5.7	97.1	29	3.4	100.0	4.5	98.5
12. BDN-1	75	72.0	58.6	35	91.4	22.8	81.7	40.7
13. T-21	63	84.1	74.6	53	90.5	60.3	87.3	67.4
14. GV-3	134	97.5	69.4	118	83.0	61.0	87.7	65.2
15. ICP-10976 (Res. check)	49	0.0	32.6	40	0.0	35.0	0.0	33.8
16. ICP-7867 (Res. check)	32	0.0	21.8	62	0.0	16.1	0.0	18.9
17. BDN-1 (SM susc. check)	154	100.0	-	155	96.1	-	98.0	-

Table 48. List of pigeonpea breeding materials screened for SM resistance during 1985-86 at ICRISAT Center.

Maturity	Materials	No. of entries	Reps.
Early	Multilocation trial entries	108	2
	Retesting and purification of ICPL's	127	2
	Station trial entries	292	1
	F4 and F5 bulks	3	1
	Vegetable F3 bulks	10	1
	EPWYT entries	5	1
	TOTAL	545	
Medium	BDN 1 BC3F4 and C 11 BC3F4 SPS (resistant)	138	1
	BDN 1 BC3F4 and C 11 BC3F4 SPS (susceptible)	122	1
	C 11 and BDN 1 backcross bulks	2	1
	MPSHWRY entries	5	1
	F7 bulks	9	1
	F3 bulks (from Test No.86, 84k)	11	1
	F3 bulks	33	1
	Advanced lines from trials	287	1
	Advanced lines test for multiplication	33	1
	TOTAL	640	
Late	F2 bulks	1	1
	Advanced lines	34	1
	TOTAL	35	1
Male steriles, hybrids and high protein lines	MS 3783, 7086, and 7035 lines	13	1
	MSBDN 1 and C 11 progenies	200	1
	MS 4A lines	42	1
	SM resistant hybrids	18	1
	Yield test lines	27	1
	HPL genetic stocks	65	1
	TOTAL	365	

Table 49. Reaction of MAHYCO lines to sterility mosaic at ICRISAT Center during 1985-86.

Pedigree	Total plants	SH plants	Percent SH
MPPL 1	24	24	100.0
2	6	6	100.0
3	10	10	100.0
4	16	All plants wilted	
5	19	19	100.0
6	13	3	23.0
7	19	19	100.0
8	17	0	0.0
9	19	All plants wilted	
10	21	1	4.7

9. Multilocation testing

The list of cooperators and locations of ICAR-ICRISAT Uniform Trial for Pigeonpea Sterility Mosaic Resistance (IIUTPSMR) is given in table 51. The reaction of these entries at six different locations is given in table 52. Several lines such as ICP 7035, 7234, 10976, 10984, 11049, 11207, BSMR 225 and BSMR 235, showed less than 10.0 percent SM across locations (Badnapur, ICRISAT Patancheru, Kanpur, Kumarganj, Pantnagar, Pudukkottai).

C. PHYTOPHTHORA BLIGHT

The main emphasis is on identification of sources of resistance to P3 isolate of the fungus. Screening of the germplasm is carried out in pots in the glass house. There is no separate programme for breeding for resistance to blight alone at present. Screening in the field for blight resistance is carried out in a multiple disease nursery where wilt and SM are also present. As soon as sources of resistance to P₃ isolate are identified, a breeding program will be undertaken.

1. Multilocation testing

The lines that are found promising in the pot screening to P2 isolate and field tolerant lines are tested in ICAR-ICRISAT Uniform Trial for pigeonpea Phytophthora Blight Resistance (IIUTPPBR). The list of cooperators and locations are given in table 53. The reaction of the lines at different locations is given in table 54. At ICRISAT this nursery was planted in the multiple disease nursery. As most of the lines died due to wilt, their reaction to blight could not be obtained. Most of the lines showed susceptibility at 4 of the locations where proper screening was carried out. The line KPBR 80-1-4 showed <20% blight at Varanasi and DPR Kanpur.

D. MULTIPLE DISEASE RESISTANCE

Screening for combined resistance to the three major diseases, wilt, SM and phytophthora blight has been in progress for the past several years. Several germplasm accessions with resistance to two diseases have been identified. Breeding for resistance to wilt and SM has also made good progress especially in the medium maturity group. Progress on breeding for combined resistance to phytophthora blight wilt and blight and SM has been slow because of lack of good sources of resistance to all the isolates of blight fungus.

1. Wilt + SM

A 0.4 ha plot in Alfisol (RM 3C and RM 8E) is being used for screening for combined resistance to wilt and SM. The field is highly sick for wilt and also high incidence of SM was created.

a) Germplasm selections

Single plant selections made from 13 germplasm accessions during 1984-85 season were retested. The data is presented in table 55. All the lines were free from SM. Two lines PR 5149 and PI 397630 also showed <10% wilt.

b) Breeding materials

The list of breeding materials screened for combined resistance to wilt and SM is presented in table 56. The detailed results have been provided to the breeders.

c) Multilocation testing (IIUTPMDR-A, 1985-86)

This is the first year of conducting ICAR-ICRISAT Uniform Trial for Pigeonpea Multiple Disease Resistance. This is a collaborative trial between ICRISAT and ICAR. This trial consists of two sets. Set A with lines having combined resistance to wilt and SM and set B with lines having combined resistance/tolerance to SM and phytophthora blight.

The list of cooperators and locations of IIUTPMDR-A is given in table 57. The reaction of the lines at different location is given in table 58. Two lines; ICP 8860 and ICP 8869 showed <20% wilt and SM across 4 locations.

2. Wilt + SM + Phytophthora blight

a) Selections from cultivars

Eight resistant SPS made from some cultivars in this nursery last season were retested. The results presented in table 59 show that no selection was resistant to wilt and SM.

b) Heliothis tolerant lines

Thirty two lines received from pulse entomology as promising to Heliothis were tested in the multiple disease nursery. SM incidence in this trial was less and the screening for this disease cannot be considered effective (Table 60 and 61). For wilt three lines; ICP 4769, ICP 6831 and ICP 7198 showed less than 20.0 percent disease.

c) ICRISAT breeding materials

The list of pigeonpea breeding materials screened in the multiple disease nursery is given in table 62. The details have been provided to breeders.

d) Multilocation testing IIUTPMDR-B (1985-86)

The list of cooperators and locations of IIUTPMDR-B (SM and blight resistance) is given in table 63 and the results in table 64. Though some lines were resistant to SM across locations, none was resistant to phytophthora blight.

Table 31. List of cooperators and locations of
IIUTPSMR 1985-86.

1.	Dr. K.K. Zote, Badnapur
2.	Dr. T.B. Anilkumar, Bangalore
3.	Dr. K.S. Amin, DFR, Kanpur
4.	Dr. R.P. Gupta, Kumargunj
5.	Dr. Gurdip Singh, Ludhiana
6.	Dr. H.S. Thripati, Panthnagar
7.	Dr. G. Arjunan, Pudukottai
8.	Mr. N.J. Bindre, Rahuri
9.	Dr. S.C. Agarwal, Sehore
10.	Dr. V.B. Chauhan, Varanasi
11.	Dr. M.V. Reddy, and Dr. Y.L. Nene, ICRISAT, Patancheru

Table 52. ICHISAT-ICAR Uniform Trial for Pigeonpea Sterility Mosaic Resistance (1983-86).

Entry	Percent SM					No. of locations with <20 % SM
	Badrupur	ICHISAT	Kanpur	Kumarganj	Pantnagar	Pudukottai
ICP 6997	8	3	13	6	0	25
ICP 7035	0	0	0	0	0	0
ICP 7234	0	0	0	0	0	18
ICP 7234	4	1	5	7	0	100
ICP 7898	11	23	2	10	0	38
ICP 8094	9	3	2	0	0	13
ICP 8862	0	0	0	0	0	0
ICP 10976	3	0	50	23	2	80
ICP 10978	3	0	0	19	0	20
ICP 10979	0	0	0	18	0	0
ICP 10982	2	0	0	18	0	0
ICP 10984	10	1	3	13	5	25
ICP 10991	0	4	9	6	0	17
ICP 10993	0	5	45	22	0	89
ICP 10996	6	0	0	0	0	0
ICP 11049	3	0	0	4	0	4
ICP 11204	0	0	2	4	0	42
ICP 11206	0	0	0	2	3	25
ICP 11207	0	0	0	0	0	10
ICP 11211	6	0	0	0	0	50
BSMR 160	-	-	-	11	0	5
BSMR 225	0	4	5	4	0	0
BSMR 235	0	0	2	10	0	0
BSMR 251	0	1	2	20	0	78
BSMR 258	0	2	7	18	0	18
BSMR 268	0	0	2	26	0	75
BSMR 277	0	0	0	17	0	50
BSMR 287	0	0	6	48	0	75
BSMR 311	0	0	4	48	0	4
BSMR 602	0	0	3	13	0	67
BWR 190	3	0	0	1	0	59
ICPL 84071	2	10	5	12	0	100
ICPL 8343	0	2	0	13	0	88
ICPL 83120	6	12	7	40	2	100
ICPL 848	11	16	0	6	0	76
ICPL 8324	1	0	0	18	0	50
ICPL 8308	22	23	11	16	0	8
ICP 7187	5	1	50	40	0	75
ICP 7182	100	97	87	95	0	93
(Range)		(90-100)	(70-100)	(88-100)	(0-32)	(72-100)

Varnant - Data received but SM incidence was negligible and hence not included.

Table 53. List of cooperators and locations of IIUTPPBR 1985-86

1. Dr. S.N. Pillai, Baroda
2. Dr. K.S. Amin, DPR, Kanpur
3. Dr. H.S. Thripati, Pantnagar
4. Dr. S.C. Agarwal, Sehore
5. Dr. V.B. Chauhan, Varanasi
6. Dr. M.V. Reddy, and Dr. Y.L. Nene, ICRISAT, Patancheru

Table 54. Reaction of the entries of IIUTPPBR to blight at different locations during 1985-86.

Cultivar	Percent blight			
	Pantnagar	Sehore	Varanasi	DPR Kanpur
ICP 28	100	55	55	42
ICP 113	55	84	30	27
ICP 1529	75	42	22	27
ICP 4135	45	68	23	31
KPBR 80-1-4	48	27	10	19
ICPL 161	85	23	29	27
ICPL 288	100	87	83	47
ICPL 8309	63	43	13	39
ICP 7119	100	98	100	61
(Range)		(93.2-100)		(45-64)

At Baroda the blight incidence in the susceptible check ICP 7119 was negligible thus the data is not recorded. At ICRISAT, because of severe wilt incidence, blight reaction could not be obtained.

Table 55. Reaction of 1984 SM promising germplasm selections to SM and wilt in SM and wilt nursery (RM 82) 1985-86 at ICRISAT Center.

S. No.	Pedigree	R I				R II				Average	
		Total		Percent		Total		Percent		Percent	
		Plants	SM	wilt	Percent	plants	SM	wilt	Percent	SM	wilt
1.	ICP 11934	27	0.0	63.9		9	0.0	55.5		0.0	59.7
2.	ICP 11438	24	0.0	62.5		17	0.0	47.0		0.0	54.7
3.	PR 5149	21	0.0	19.0		2	0.0	0.0		0.0	9.5
4.	PR 5145	13	0.0	23.1		9	0.0	33.3		0.0	28.2
5.	PR 5117	13	0.0	46.1		11	0.0	90.9		0.0	64.5
6.	PI 397456	16	0.0	18.8		5	0.0	40.0		0.0	29.4
7.	PR 5164	12	0.0	8.3		9	0.0	22.2		0.0	15.2
8.	ICP 11844	20	0.0	40.0		9	0.0	11.1		0.0	25.5
9.	PI 397630	5	0.0	0.0		12	0.0	16.7		0.0	8.3
10.	Sri Lanka 477-1	16	0.0	25.0		3	0.0	33.3		0.0	29.1
11.	PR-5140	14	0.0	21.4		13	0.0	38.5		0.0	29.9
12.	PR 5294	11	0.0	100.0		5	0.0	10.0		0.0	100.0
13.	PR-5118	13	0.0	30.7		18	0.0	66.6		0.0	48.6
	ICP 8463 (SM susc. check)	46	76.3	-		40	78.9	-		77.6	-
	ICP 2376 (wilt susc. check)	92	-	98.4		61	-	98.2		-	98.3

Table 56. List of pigeonpea breeding materials screened for combined resistance to wilt and SM during 1985-86 (RM 3C & RM 8E) at ICRISAT Center.

Maturity	Material	Entries	Reps.
Medium	NPSHWRY	10	1
	F3 bulks population (test No.87.84k)	5	1
	ICPL 227 SPP's	84	1
	Vegetable DT yield test bulks	76	1
	Vegetable NDT yield test bulks	55	1
	TOTAL	230	
Late	LPAY entries	10	1
Male steriles	MS NPWR-15 BC5F2s	7	1
	WRMS 3783 BC2F4s	18	1
	TOTAL	25	

Table 57. List of cooperators and locations of IIUTPMDR-A 1985-86 (Wilt + SM resistance).

1. Dr. K.K. Zote, Badnapur
2. Dr. K.S. Amin, DPR, Kanpur
3. Dr. G. Arjunan, Pudukkottai
4. Mr. N.J. Bindre, Rahuri
5. Dr. M.V. Reddy, and Dr. Y.L. nene, ICRISAT, Patancheru

Table 58 Reaction of the entries of IITUPMHR-A (wilt and SM resistance) at different locations during 1985-86.

Percent disease										
S. No.	Entry	Sadnapur		Kanpur		Pudukkottai			ICRISAT Hyderabad	
		Wilt	SM	Wilt	SM ²	FB	Wilt	SM	Wilt	SM
1.	ICP 8860	9.4	4.7	0.0	0.0	9.5	0.0	17.9	12.5	0.0
2.	ICP 8861	18.3	0.0	-	0.0	100.0	0.0	27.3	5.0	1.5
3.	ICP 8862	30.3	0.0	0.0	0.0	75.0	0.0	6.6	57.3	3.5
4.	ICP 8867	9.8	0.0	0.0	0.0	0.0	0.0	16.6	41.6	1.1
5.	ICP 8869	5.6	0.0	0.0	7.7	50.0	0.0	0.0	6.8	6.8
6.	ICP 10960	0.0	0.0	0.0	7.7	100.0	0.0	7.0	41.1	30.0
7.	ICP 11289	15.6	0.0	-	0.0	100.0	0.0	78.5	30.5	1.2
8.	ICP 11290	57.9	0.0	0.0	0.0	33.3	0.0	65.6	41.5	0.0
9.	ICP 11291	51.4	2.2	0.0	0.0	36.3	0.0	28.1	23.3	3.0
10.	ICP 11296	12.5	0.0	100.0	-	-	0.0	0.0	59.6	0.0
11.	ICP 11297	0.0	0.0	-	9.1	100.0	0.0	0.0	23.5	0.0
12.	ICP 11298	40.0	0.0	-	8.3	100.0	0.0	14.9	34.8	0.0
13.	BSMR 68	10.3	0.0	0.0	0.0	62.5	0.0	0.0	29.3	2.9
14.	BSMR 540	2.4	0.0	0.0	0.0	45.0	0.0	22.5	27.8	0.0
15.	BSMR 294	6.5	0.0	0.0	0.0	50.0	0.0	16.6	25.9	0.0
16.	BSMR 544	4.2	0.0	0.0	0.0	40.0	0.0	25.0	30.3	0.0
17.	BSMR 736	10.9	0.0	0.0	5.5	62.5	0.0	30.7	32.7	0.0
18.	BSMR 268	94.7	2.3	0.0	0.0	10.0	0.0	50.0	80.0	0.0
19.	BSMR 520	6.8	0.0	0.0	0.0	12.5	0.0	39.3	23.8	1.6
20.	BSMR 595	96.2	0.0	0.0	0.0	66.6	0.0	50.0	87.1	0.0
21.	BSMR 161	15.4	0.0	0.0	0.0	100.0	0.0	3.9	25.6	0.0
22.	BSMR 225	44.6	0.0	-	0.0	100.0	0.0	10.0	54.8	7.6
23.	BSMR 277	36.0	0.0	0.0	0.0	66.6	0.0	0.0	14.6	7.1
24.	BSMR 285	79.5	2.6	5.6	9.1	44.4	0.0	91.7	52.7	1.4
25.	BSMR 235	69.6	0.0	0.0	10.0	24.9	0.0	11.1	56.1	0.0
26.	BSMR 231	87.5	4.5	0.0	9.1	16.6	0.0	87.5	79.1	0.0
27.	BSMR 258	76.7	0.0	0.0	11.2	83.3	0.0	16.6	77.5	0.0
28.	BSMR 160	52.9	0.0	-	0.0	100.0	0.0	0.0	51.6	2.6
29.	BSMR 311	35.3	2.4	0.0	0.0	0.0	0.0	11.1	79.0	0.0
30.	BSMR 602	20.6	0.0	0.0	0.0	55.0	0.0	16.6	72.0	44.3
31.	BSMR 1	26.3	0.0	-	0.0	100.0	0.0	60.6	15.1	0.0
32.	BSMR 2	45.6	2.5	50.0	7.1	50.0	0.0	50.0	93.2	18.0
33.	BWR 67	32.9	51.4	0.0	0.0	0.0	0.0	60.5	80.6	93.8
34.	BWR 97	2.3	0.0	16.6	33.3	66.6	0.0	63.4	26.0	100.0
35.	BWR 135	32.6	40.0	0.0	18.1	20.0	0.0	71.5	77.0	64.6
36.	BWR 153	5.3	11.8	-	0.0	100.0	0.0	75.0	32.6	28.4
37.	BWR 154	16.7	0.0	0.0	11.1	30.0	0.0	30.0	10.9	0.0
38.	BWR 159	8.3	3.6	0.0	0.0	0.0	0.0	16.6	14.3	25.4
39.	BWR 175	7.8	0.0	0.0	0.0	34.2	0.0	0.0	35.4	0.0
40.	BWR 190	43.9	5.3	0.0	20.6	50.0	0.0	43.2	61.8	11.9
41.	BWR 198	14.8	7.5	0.0	30.0	100.0	0.0	25.0	49.5	9.1
42.	BWR 217	12.9	29.7	0.0	50.0	48.0	0.0	78.2	56.3	87.8
43.	BWR 234	8.2	0.0	0.0	27.7	80.0	0.0	49.9	31.0	100.0
44.	BWR 259	24.3	0.0	0.0	0.0	68.8	0.0	0.0	29.5	0.0
45.	BWR 245	29.5	74.3	0.0	27.2	71.4	0.0	47.4	34.6	98.1
46.	BWR 301	5.0	0.0	0.0	14.2	35.4	0.0	4.2	40.7	0.0
47.	BWR 321	29.0	8.8	50.0	0.0	75.0	-	-	26.4	32.0
48.	BWR 322	26.9	5.1	-	20.0	37.7	0.0	33.6	47.7	18.0
49.	BWR 332	8.9	4.8	-	20.0	100.0	0.0	26.9	38.5	48.0
50.	BWR 369	17.7	2.4	0.0	6.6	87.5	0.0	35.0	22.3	6.0
51.	BWR 370	13.1	100.0	0.0	85.7	71.4	0.0	84.0	46.4	100.0
52.	BWR 250	6.4	5.6	0.0	5.8	83.8	0.0	39.1	33.1	100.0
53.	ICP 2370	100.0	-	19.6	-	55.1	32.7	-	98.9	100.0
	(wilt check)			(0-100)		(0-100)	(20-75)		(97-100)	
54.	ICP 7182	-	100.0	-	78.1	-	83.6			
	(SM check)				(45-100)		(50-100)			

caution.

2. One replication only.

ICP 8869 showed <10% SM and wilt across locations and

ICP 8860 showed <20% SM and wilt across locations

Table 59. Reaction of some selections from pigeonpea cultivars to sterility mosaic, and wilt, in the multiple disease nursery (RP 18) during 1985-86 at ICRISAT Center.

No.	Pedigree	Total plants	Percent SM	Percent wilt
1.	ICP-88630-SVB0 (SM plants)	142	100.0	32.3
2.	ICP-88630-SVB0 (Late SM symptoms)	108	29.6	11.1
3.	ICP-8863-OP-SVB0 (more pods)	108	100.0	25.3
4.	Gujarat local-SVPB0	132	42.4	46.2
5.	BDNA-5-SVPB0	141	90.7	95.7
6.	PDA-84-2-SVPB0	123	1.5	97.5
7.	PDA-83-3-SVPB0	49	40.8	67.3
8.	BVR-175-SVPB0	133	24.8	0.0
	ICP 8863 (SM susc. check)	265	74.7	-
	ICP 2376 (wilt susc. check)	484	-	87.0

Table 60. Reaction of entomologically promising lines to sterility Mosaic and wilt in the multiple disease nursery (RP 18) during 1985-86 at ICRISAT Center.

S.No.	Pedigree	R I			R II			Average	
		Total plants	Percent SM	Percent wilt	Total plants	Percent SM	Percent wilt	Percent SM	Percent wilt
1.	ICP-8127	50	0.0	48.0	29	3.4	62.1	1.7	55.0
2.	ICP-810	44	20.4	72.7	61	13.1	90.2	16.7	44.7
3.	ICP-1903	44	18.2	100.0	21	0.0	100.0	9.1	100.0
4.	ICP-8571	40	15.0	95.0	35	0.0	94.3	7.5	94.6
5	GS-1	26	11.5	100.0	29	10.3	79.3	10.9	89.6
6.	ICP-7041	15	13.3	100.0	46	6.5	89.1	9.9	94.5
7	ICP-8010	23	0.0	86.9	55	1.8	96.4	0.9	91.6
8	ICP-7050	37	5.4	89.2	55	18.2	92.7	11.8	91.0
9.	ICP-4640	76	15.8	80.3	27	11.1	96.3	13.4	88.3
10.	ICP-7496	62	9.7	87.1	41	12.2	80.4	10.9	83.7
11.	ICP-7176-18-E2	9	11.1	55.5	10	0.0	90.0	5.5	72.7
12	ICP-8102-E1	35	0.0	68.6	30	3.3	26.7	1.6	73.0
13.	PPE-36-2	70	1.4	87.1	39	7.7	58.9	4.5	73.0
14.	ICP-1925-(1G)-2	-	-	-	-	-	-	-	-
15.	ICP-5766	55	1.8	96.4	25	20.0	80.0	10.9	88.2
16	PPE-38-1	64	7.8	92.2	40	5.0	85.0	6.4	88.6
	ICP 8863	101	27.8	-	131	17.8	-	22.8	-
	(SM susc. check)								
	ICP 2376	214	-	93.4	176	-	92.3	-	92.8
	(wilt susc. check)								

Table 61. Reaction of entomologically promising lines to sterility mosaic and wilt in the multiple disease nursery (RP 18) during 1955-56 at ICISAR Center.

S.No. Pedigree	R I				R II				Average			
	Total plants	Percent SM	Percent wilt	Total plants	Percent SM	Percent wilt	Percent SM	Percent wilt	SM	Percent wilt	SM	Percent wilt
1. ICP-6811	34	0.0	5.3	-	-	-	-	-	4.3	15.6	-	-
2. ICP-288	28	0.0	50.0	40	2.5	67.5	67.5	73.9	1.2	59.8	0.0	47.0
3. ICP-6815	5	0.0	20.0	23	0.0	0.0	0.0	16.7	0.0	11.1	0.0	0.0
4. ICP-4769	18	0.0	5.5	6	0.0	0.0	0.0	86.2	9.1	82.5	0.0	0.0
5. ICP-1059	38	7.9	78.9	29	10.3	86.2	86.2	82.6	2.5	63.8	0.0	0.0
6. ICP-8660	20	5.0	45.0	23	0.0	40.0	40.0	10.5	10.5	41.7	0.0	0.0
7. ICP-11410 (MFR-11)	53	9.4	43.4	60	11.7	96.7	96.7	7.5	7.5	96.0	0.0	0.0
8. ICP-14060	20	15.0	95.0	61	0.0	0.0	0.0	71.4	0.0	85.7	0.0	0.0
9. ICP-4307	5	0.0	100.0	7	0.0	31.9	31.9	26.0	0.0	26.7	0.0	0.0
10. MA-2	40	0.0	17.5	91	0.0	8.6	8.6	100.0	4.3	93.0	0.0	0.0
11. ICP-8669	36	0.0	86.1	35	8.6	37.8	37.8	1.4	1.4	34.4	0.0	0.0
12. ICP-5498	35	2.9	31.4	37	0.0	97.2	97.2	1.4	1.4	97.3	0.0	0.0
13. ICP-4886	39	0.0	97.4	72	2.8	20.0	20.0	2.5	2.5	18.6	0.0	0.0
14. ICP-7198	29	0.0	17.2	20	5.0	75.0	75.0	4.6	4.6	65.8	0.0	0.0
15. ICP-4070	23	4.3	56.5	20	5.0	63.3	63.3	2.7	2.7	74.7	0.0	0.0
16. QW-3	36	5.5	86.1	30	0.0	-	-	43.5	43.5	-	-	-
ICP-8863	63	61.0	-	126	26.1	-	-	-	-	-	-	-
ISM subC	-	-	-	83.4	102	-	-	79.3	79.3	-	-	-
ICP-2876	197	-	-	-	-	-	-	-	-	-	-	-

Table 62. List of Pigeonpea Breeding materials screened in the multiple disease nursery during 1985-86 (RP 18) at ICRISAT Center.

Maturity group	Materials	Entries	Reps.
Early	Multilocation trial entries	108	2
	Retesting and purification of ICPL's	22	2
	F5 bulks	2	1
	ICPL 269 SPP.	100	1
	ICPL bulks (ICPL 8327, and 269)	2	1
	TOTAL	234	
Medium	F5 SPP.	170	1
	F5 SPP. (from bulk populations)	89	1
	TOTAL	259	
Late	F3 SPP.	22	1
	Advanced lines	30	1
	F3 bulks	4	1
	TOTAL	56	

Table 63. List of cooperators and locations of IIUTPMHDR-B 1985-86 (SM + PB resistance).

1. Dr. K.S. Amin, DPR, Kanpur, Uttar Pradesh
2. Dr. H.S. Thripati, Pantnagar, Uttar Pradesh
3. Dr. G. Arjunan, Pudukkottai, Tamil Nadu
4. Dr. M.V. Reddy, and Dr. Y.L. Nene, ICRISAT, Patancheru

Table 64. Reaction of the entries of IITPMDR-B (SM and PB resistance) at different locations during 1985-86.

No.	Entry	Percent disease									
		Kanpur		P		Pantnagar		Pudukkottai		ICRISAT ²	
		Wilt		Blight		P. Blight		SM		Wilt	
		Trial1	Trial2	Trial1	Trial2	Trial1	Trial2	SM	Wilt	SM	Wilt
1.	ICP 11290	0.0	0.0	0.0	0.0	100.0	0.0	6.3	61.8	0.0	0.0
2.	ICP 11290	0.0	0.0	15.7	0.0	92.5	0.0	88.9	13.2	9.6	0.0
3.	ICP 11300	0.0	0.0	75.0	16.7	100.0	0.0	95.4	91.8	0.0	0.0
4.	ICP 11301	0.0	0.0	100.0	0.0	80.5	0.0	18.5	45.8	0.0	0.0
5.	ICP 11302	0.0	0.0	83.3	5.1	93.5	0.0	29.8	90.0	0.0	0.0
6.	ICP 11303	3.1	36.2	49.3	0.0	88.7	0.0	34.4	59.9	0.0	0.0
7.	ICP 11304	3.6	0.0	3.4	0.0	34.8	0.0	11.3	5.2	0.0	0.0
8.	ICP 11302	3.4	64.5	55.1	-	100.0	0.0	71.8	-	61.3	-
9.	(SM check)	(0.0)	(32.87)	(53.80)	-	-	-	(59.86)	-	(45.77)	-
10.	IPB check	0.0	0.0	88.9	25.3	99.0	0.0	7.6	-	-	-
				(87-100)	(13.47)	(99-100)	(0-19)				

1-Plant stand was poor and the results to be treated with caution
 2-Phytophthora blight screening was not possible as the stand was affected by wilt
 3-ICP 11304 only showed some promise against SM and phytophthora blight across locations.

B. WILT + HELIOTHIS

1. Wilt + Heliothis tolerant lines

Eleven Heliothis and wilt tolerant SPP of 3 germplasm lines were retested. All these lines were found resistant to wilt (Table 65). There were ICP 4769, ICP 6831 and ICP 7198.

2. Breeding materials

Seventeen F2, and F3 bulks (Table 66) and 2800 LRG 30 M2 progenies were screened for wilt resistance and the results provided to breeders.

Table 65. Advanced screening of wilt and *Heliothis* tolerant pigeonpea germplasm lines against wilt + *Heliothis* nursery 1985-86 (BH-16C) at ICRISAT Center.

S.No.	Material	Total plants	Wilted plants	Percent wilt
1.	ICP 4769-VB0-V10	214	6	2.8
2.	ICP 4769-VB0-V20	190	2	1.1
3.	ICP 4769-VB0-V30	256	6	2.3
4.	ICP 4769-VB0-V40	255	2	0.8
5.	ICP 4769-VB0-V50	92	6	6.5
6.	ICP 6831-VB0-V10	122	4	3.3
7.	ICP 6831-VB0-V20	70	0	0.0
8.	ICP 6831-VB0-V30	66	0	0.0
9.	ICP 7198-VB0-V10	60	0	0.0
10.	ICP 7198-VB0-V20	60	2	3.3
11.	ICP 7198-VB0-V30	170	20	11.8
12.	ICP 1903 E10	267	264	98.9
(susceptible to wilt but tolerant to <i>Heliothis</i>)				

ICP 6831 is agronomically desirable with spreading habit. It is of medium-late maturity type.

Table 66. Wilt incidence in F2 and F3 wilt x *Heliothis* tolerant populations in wilt + *Heliothis* nursery during 1985-86 (BH-16C) at ICRISAT Center.

S.No.	Cross No.	Generation	Total plants	Wilted plants	Percent wilt
1.	ICPX 830109	F2	2200	892	40.5
2.	ICPX 830110	F2	2224	261	11.7
3.	ICPX 830111	F2	1904	769	40.4
4.	ICPX 830111(RC4)	F2	1450	643	42.3
5.	ICPX 830112	F2	1849	1120	60.6
6.	ICPX 830113(RC4)	F2	1988	1702	85.6
7.	ICPX 830114	F2	2921	943	32.3
8.	ICPX 830115	F2	2555	1121	43.9
9.	ICPX 830116	F2	2930	1456	49.6
10.	ICPX 830117	F2	1774	365	20.6
11.	ICPX 830118	F2	3035	1368	45.1
12.	ICPX 830119	F2	1637	1504	91.9
13.	ICPX 830120	F2	2958	1296	43.8
14.	ICPX 830121	F2	1925	1678	87.2
15.	ICPX 820132-VB	F3	2429	1300	53.5
16.	ICPX 820133-VB	F3	1686	1539	91.3
17.	ICPX 83 EB 022	F2	3556	3500	98.4
(MS C 11 x ICPL 332)					