RP/02085

Pulse Pathology Progress Report-10

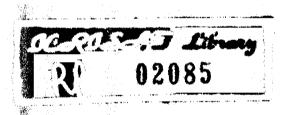
Pulse Pathology (Chickpea) Report of Work

(June 1979 - May 1980)



International Crops Research Institute for the Semi-Arid Tropics
ICRISAT Patancheru P.O.
Andhra Pradesh 502 324 . India

This report has been prepared to share the information with scientists having interest in chickpea improvement. This is not an official publication of the Institute and should not be cited.



YLN

PULSE PATHOLOGY SUB-PROGRAM (CHICKPEA)

STAFF

Dr. Y.L. Nene	• •	Leader, Pulses Improvement Program and Principal Plant Pathologist
Dr. M.V. Reddy	• •	Plant Pathologist S-2
Dr. M.P. Haware		Plant Pathologist S-2
Mrs. Sheila Vijayakumar		Research Technician
Mr. J. Narayan Rao		Technical Assistant
Miss E. Deena	• •	Technical Assistant (From April 1979)
Mr. K. Prabhakar Reddy		Field Assistant
Mr. G. Musala Reddy	• •	Field Assistant
Mr. A. Chandar		Secretary I
Mr. R. Narsing Rao		Stenographer
Mr. Mohd. Sharfuddin Khan		Driver-cum-General Assistant II
Mr. M.M.S. Ali Baig		Driver-cum-General Assistant I

PULSE PATHOLOGY SUB-PROGRAM (CHICKPEA)

LIST OF APPROVED PROJECTS (1978-1980)

Sub-program Leader : Y. L. Hene

No.	Title	Project Scientist	Cooperators
CP-Path-1	Studies on Fusarium wilt of chickpea	M.P. Haware	J. S.C. Sethi C.L.L. Gowda O. Singh
CP-Path-2	Studies on stem and root rots of chickpeas	M.P. Haware	J. S.C. Sethi C.L.L. Gowda O. Singh
CP-Path-3	Studies on chickpea stunt and other viral diseases	M.V. Reddy	J.P. Verma (HAU, Hissar) J. Kumar C.L.L. Gowda O. Singh W. Rmed
CP-Path-4	Studies on Ascochyta blight	M.V. Reddy	O. Singh J. Kumar K.B. Singh (ICARDA)
CP-Path-5	International chickpea disease nurseries	Y.L. Nene	M.P. Haware M.V. Reddy L.J.G. van der Maesen

CONTENTS

				Page Nos.
PF	ROJECT: CP-PATH-1(78) : STUDIES OF CHIC		FUSARIUM	WILT
I.	SUMMARY	• •	• •	1
II.	INTRODUCTION	• •	• •	1
III.	FIELD SCREENING FOR WILT RESISTANCE	• •	• •	2
	A. Colletotrichum blight-promising 1	in ės	• •	2
	B. Stunt-promising lines	••	••	2
	C. Ascochyta blight-promising lines	• •	• •	2
	D. Germplasm	• •		2
	E. Breeders' material	••		2
IV.	FURTHER STUDIES ON RACES	••	• •	3
v.	SURVIVAL OF THE WILT FUNGUS	• •		4
	A. Longevity	••		4
	B. Longevity as influenced by depth	of burial	• •	5
VI.	STUDIES ON INOCULUM POTENTIAL	• •	••	6
VII.	INHERITANCE STUDIES	• •	• •	9
vIII.	YIELD TRIAL	• •	• •	11
PF	ROJECT: CP-PATH-2(78) : STUDIES OF CHIC		4 AND ROOT	ROTS
ı.	SUMMARY			14
II.	INTRODUCTION			14
III.	DRY ROOT ROT (RHIZOCTONIA BATATICOLA)		15
IV.	SCREENING TECHNIQUE			15
	1. Multiplication of inoculum			16

					Page Nos
	2. Rai	sing of seedlings	••	• •	16
	3. Inc	oculation	••	••	16
	4. Inc	ubation	• •	• •	16
	5. Die	ease rating (Dry root :	rot)	• •	16
		t-promising lines screebatatioola (dry root re	•	••	. 17
V.	FIELD SCR	EENIN G	• •	• •	17
	A. Wilt-p	romising lines from 19	78-79	• •	17
	B. Entrie	s received from other	locations	• •	18
	1. New	Delhi	••	••	18
	2. Kan	pur	• •	• •	18
	3. Gur	daspur	••	••	18
	4. Bad	napur	••	••	18
	C. GIET		••		-18
	D. GCVT		• •	• •	18
VI.		ISOLATIONS FROM WILTED, FROM MULTIPLE DISEASE		••	19
VII.	SURVIVAL IN SOIL	OF SCLEROTIUM ROLFSII	(COLLAR ROT PATHO	XGEN)	20
VIII.	MISCELLAN	EOUS STUDIES	••	••	21
	A. Screen	ing against Colletotric	chum blight	••	21
	1. Lab	oratory screening	• •	• •	21
	2. Fie	ld Screening		• •	22
	3. Pat	hogenic behaviour of $\mathcal{C}.$	dematium and C.	capsici	22
	B. Leaf d	iseases	••	••	23
	l. Bot	rytis grey mold			23

			Page Nos.
(a) Pathogenicity			23
(b) Symptoms			23
2. Stemphyllium leaf spot			23
3. Alternaria leaf spot			24
PROJECT: CP-PATH-3(78) : STUDIE VIRAL	S ON CHICKPEA DISEASES	STUNT	AND OTHER
I. SUMMARY	• •		25
II. INTRODUCTION	• •		26
III. SCREENING NURSERY	• •	• •	26
IV. SCREENING FOR RESISTANCE	• •	• •	26
A. Germplasm	• •		26
B. Advanced germplasm lines	••	• •	27
1. 1976-77 Selections	• •	• •	27
2. 1977-78 Selections			27
3. 1978-79 Selections	• •	• •	28
C. Crossing block entries	• •	• •	29
1. Advanced lines	• •	••	29
2. Observations in 1979-80 cross	sing block nurses	·y	29
D. Ascochyta blight-resistant lines	5	• •	31
1. Advanced lines	• •		31
2. Preliminary screening			31
E. Wilt and root rot resistant line	8		33
1. Advanced lines	••	• •	33
2. Preliminary screening	••	••	33
F. ICCC materials	• •	• •	33

			P	age Nos
	G. GIET lines			33
	H. GCVT lines			36
	I. Ascochyta blight-promising lines :	from Gurdaspur		36
	J. Multilocation testing	••		36
	K. F2 materials			36
v.	EFFECT OF PLANTING DATES			37
VI.	EFFECT OF PLANT POPULATIONS			37
	A. Sprayed conditions			38
	B. Unaprayed conditions			38
	OJECT: CP-PATH-4(78): STUDIES			41
		• •	• •	
	INTRODUCTION	••	• •	42
III.	SCREENING TECHNIQUE	• •	• •	42
IV.	SCREENING FOR RESISTANCE	••	• •	42
	A. Kabuli germplasm	••	• •	42
	B. Repeat screening of germplasm	• •		43
	C. Materials from other locations	• •		44
	1. ICARDA promising lines	••	• •	44
	2. Gurdaspur promising lines	••	• •	44
	3. GIET entries	••	• •	44
	4. GCVT entries	••	••	47
	D. Inheritance study	• •	••	47
	1. Parents and P1 material	• •	• •	47

				Page Nos.
	2. F ₂ materials	• •	• •	47
	3. Collection of seed from re	sistant plants	• •	48
	E. Collection of new isolates	• •	• •	49
v.	DISEASE CONTROL	• •	• •	49
	A. Effect of fungicides on spore	germination		49
VI.	MECHANISM OF RESISTANCE	•••	4 •	49
	A. Leaf exudates	• •	• •	49
	 Effect of exudates from su moderately resistant lines 	•	 mation	51
	Effect of different diluti susceptible and moderately on spore germination			51
	3. pH of the exudates at diff	erent times of t	he day	51
	4. pH at different ages	• •		51
	B. Leaf extracts	••	• •	51
	C. Acid mixture	••	••	55
	D. Role of yeast cells	••	• •	55
VII.	SEED MULTIPLICATION	••	• •	56
vIII.	SUMMARY OF WORK DONE AT ICARDA	••	••	56
IX.	SUMMARY OF THE REPORT ON TRIPS T MOROCCO, AND TUNISIA BY DR. M.V.	•	••	57
Pf	ROJECT: CP-PATH-5(78) : INTE NURS	ERNATIONAL CHI SERIES	CKPEA DIS	SEASE
I.	SUMMARY			59
II.	INTRODUCTION			59
TTT	TCRRWN 1979-80			60

			Page Nos
	A. List of countries and	cooperators	60
	B. Entries	••	63
ıv.	TOURS	••	64
v.	CONSULTANT		65
VI.	SECOND INTERNATIONAL CHIC TRAINING COURSE	KPEA PATHOLOGY	• 65
	APPENDIX-1	••	68

I. SUMMARY

- 1. Of the 2628 additional germplasm accessions acreened, 133 were found promising against the wilt by showing less than 20% mortality. These will be tested again.
- 2. None of the stunt-promising lines was found resistant.
- 3. Only one line ICC-3 was found promising against the wilt out of 19 Colletotrichum-blight-promising lines.
- Ascochyta blight-promising line ICC-3935, was also found promising against the wilt.
- 5. Evidence was obtained to confirm the existence of at least 3 physiological races of F. oxysporum f. sp. oiosri.
- 6. The wilt fungus is able to survive up to 27 months in infected roots and stems buried in soil. It was isolated after 12 months from host tissues buried up to 24-inch depth in soil. The experiments are continuing.
- 7. About 3000 propagules of F. oxysporum f. sp. ciceri per gram of soil was found essential to get 100 percent mortality in the susceptible cultivar, JG-62.
- 8. Help was extended to breeders in the studies on inheritance of wilt resistance.
- 9. In a yield trial with 14 resistant cultivars and 4 check cultivars (for yield) conducted at Hyderabad, cultivars JG-74, NEC-790, and Annique performed well. JG-74 gave highest yield.

II. INTRODUCTION

The project became operative from January 1978 with the following objectives:

- 1. Study survival and spread of the pathogen (Fusarium oxysporum f. sp. ciceri),
- 2. Study the situation on pathogenic races, if any,
- 3. Further improve screening techniques, and
- 4. Screen germplasm/breeding material for resistance.

Work carried out during 1979-80 is reported here.

III. FIELD SCREENING FOR WILT RESISTANCE

Material was planted in a wilt-sick plot (BT-6C) in 4 meter rows, 75 cm apart. Susceptible check JG-62 was planted after every 2 test rows. Periodic observations on wilt incidence were recorded. The materials screened were:

Colletotrichum-blight-promising lines	19
Stunt-promising lines	6
Ascochyta blight-promising lines	141
Germplasm	2628

A. Colletotrichum-blight-promising lines

Of the 19 lines tested all, except ICC-3, showed more than 20% wilt.

B. Stunt-promising lines

None was promising.

C. Ascochyta blight-promising lines

Of the 141 lines tested, ICC-3935 was found promising.

D. Germplasm

The following 133 accessions showed less than 20% wilt.

```
ICC-154, 184, 240, 268, 301, 573, 585, 594, 647, 773, 805, 857, 859, 871, 884, 925, 933, 967, 1019, 1038, 1039, 1101, 1129, 1132, 1155, 1234, 1246, 1250, 1279, 1288, 1292, 1296, 1297, 1298, 1314, 1316, 1319, 1330, 1393, 1397, 1403, 1405, 1434, 1435, 1437, 1441, 1448, 1449, 1451, 1477, 1491, 1516, 1550, 1567, 1587, 1594, 1597, 1599, 1606, 1624, 1649, 1663, 1664, 1688, 1694, 1710, 1712, 1716, 1718, 1719, 1723, 1753, 1755, 1756, 1758, 1795, 1820, 1901, 1939, 1984, 1987, 2031, 2032, 2034, 2036, 2037, 2039, 2041, 2061, 2089, 2135, 2243, 2246, 2250, 2253, 2263, 2304, 2484, 2519, 2520, 2547, 2580, 2595, 2797, 2800, 2831, 3067, 3072, 3075, 3076, 3095, 3208, 3219, 3273, 3274, 3328, 3407, 3448, 3449, 3451, 3457, 3458, 3463, 3470, 3494, 3504, 3508, 3536, 3537, 3538, 3545, 3650, and 3768.
```

All these lines will be retested next year in a wilt-sick plot.

E. Breeders' material

The material from F_2 through F_9 generations was planted in a wilt-sick plot (M-5). Progenies considered superior by the pathologist had

been advanced by the breeders by individual plant selections or they were bulked. The materials screened were:

F ₂ generation	171 populations
F ₃ generation	1302 progeny rows
F ₄ generation	638 progeny rows
F5-F8 generation	209 progeny rows
Back cross F2	536 progeny rows
r ₆ (bulk)	36 entries
F7-F8 (bulk)	31 entries) 2 replications
F ₅ -F _q (bulk)	11 entries
F ₂ generation	398% single plant selections
F ₃ generation	653
F ₄ generation	385
F5-F8 generation	93
Back cross F ₂	150

 F_6 , F_7 - F_8 and F_4 - F_9 (bulks) will be retested. In addition 203 lines were bulked in F_3 - F_8 generation.

IV. FURTHER STUDIES ON RACES

Studies during the last two years on 5 isolates F. oxysporum f. sp. ciceri collected from Hyderabad, Jabalpur, Kanpur, Hissar, and Gurdaspur provided us with the evidence that there are at least 3 physiological races of the chickpea wilt fungus (see Pulse Pathology (Chickpea) Report of Work 1978-79). The reaction of these isolates on 10 chickpea cultivars was generally consistent.

The isolates from Hyderabad, Kanpur, and Gurdaspur gave distinct reactions and were used in further study. Thirteen resistant and 2 susceptible lines (JG-62, C-104) to the Hyderabad isolate were used.

The inoculum was multiplied on sand-maize meal medium (9:1) in 250 ml flasks for 14 days. One hundred g of inoculum was mixed in a plastic pot (15-cm dia) containing 2 kg of autoclaved soil (Vertisol) sand mixture (1:1). All the plastic pots were washed in running water, dipped in 54 CuSO₄ solution and then air dried before use.

Seedlings for each cultivar were raised in autoclaved sand for a week, removed and then transplanted, 5 in each pot. Twenty seedlings were tested against each isolate. Non-inoculated checks for each cultivar were kept. Pots were irrigated with sterilized water and utmost care was taken to avoid cross contamination.

A critical look at the results in Table 1 indicates that out of 13 resistant lines at Hyderabad; BG-212, P-1265, and P-1353 were susceptible to the Kanpur isolate and P-165 showed moderate susceptibility. C-104 which is susceptible to the Hyderabad and Kanpur isolates was resistant to the Gurdaspur isolate. BG-212 and WR-315 and JG-62 were moderately susceptible to the Gurdaspur isolate; others were resistant.

These results have confirmed our earlier observations that these 3 isolates are different in their pathogenic behaviour and are distinct races.

Table 1. Reaction of chickpea cultivars to three isolates of Pusarium oxysporum 1. sp. cicsria

	Rea	ction to isola	ite ¹
Cultivars	Hyderabad	Kanpur	Gurdaspur
JG-62	s	S	М
WR-315	R	R	М
C-104	S	S	R
BG-212	R	S	M
DA-1	R	R	R
P-165	R	M	R
P-289	R	R	R
P-517	R	R	R
P-678	R	R	R
P-1265	R	S	R
P-1270	R	R	R
P-1353	R	S	R
P-4116-1	R	R	R
P-60 9 9	R	R	R
NEC-790	R	R	R

Readings were taken 60 days after inoculation (experiment was repeated); 20 seedlings for each cultivar were used.

V. SURVIVAL OF THE WILT FUNGUS

A. Longevity

An experiment was initiated in March 1978 to find out how long F. oxysporum f. sp. oiceri survives in different plant parts of chickpea. In this experiment, roots with 5-cm stem base from naturally infected plants were buried in 45-cm pots (bottom removed) in soil. The pots themselves were buried in soil in a way that top of the pots was in level with soil surface. All the roots were weighed before burial. Four roots were carefully removed after every 3 months from the pots, dried and weighed. After washing in running water, the tissues were surface-sterilized in 2.5% sodium hypochlorite for 2 to 3 minutes and isolations were attempted. Pathogenicity was checked.

In the last year's report, we reported that the fungus could be isolated from these roots for 15 months. The isolations were con-

bR = Resistant (0-20% wilt); M = Moderately susceptible (21-50% wilt); S = Susceptible (51-100% wilt).

tinued further and the data indicated that the fungus could be isolated from disintegrating tissues even after 27 months. This indicates that the chickpea wilt fungus can survive in the soil in host tissues for more than 2 years in the absence of host. The experiment is continuing (Table 2).

The wilt fungus could be isolated from all the plant parts of chickpea. We reported (see Pulse Pathology (Chickpea) Report of Work 1978-1979) that the fungus could not be isolated from the leaflets after 2 months when stored in laboratory or buried in soil. After 12 months it could not be isolated from any part of the plant stored at room temperature but it could be isolated from stem and root tissues buried in soil even after 27 months.

Table 2. Survival of F. oxysporum f. sp. oiceri in buried roots

	Original weight	Weight at the time of iso-	
Date of isolation	(g)	lation (g)	Isolation
10-6-1979	5.54	1.06	+
(reported earkier)	5.42	1.05	+
_	8.02	1.58	+
	8.45	1.48	+
10-9-1979	17.16	0.71	+
	4.20	0.36	+
	4.83	0.25	+
	2.01	0.15	+
10-12-1979	8. 56	0.19	+
	5.8 2	0.19	+
	4.32	0.25	+
	2.97	0.80	+
10-3-1980	6.65	0.33	+
,	11.33	0.73	+
	3.8 8	0.35	+
	3.9 8	0.20	+
10-6-1980	7.35	0.06	+
	2.19	0.02	+
	3.8 2	0.03	+
	3.24	0.06	+

Roots were buried on 10 March 1978.

B. Longevity as influenced by depth of burial

Roots from chickpea wilted plants were collected in March 1979. They were air-dried and made into small pieces of 20 to 25 mm. Each

sample consisted of 10 pieces and placed in nylon mesh after weighing. Diseased samples were kept at various depths in earthen pot (45 cm) in soil which itself was buried in soil after removal of bottom. Top of the pot was in level with soil. After every 3 months samples from one pot were assayed for chickpea wilt pathogen. The experiment is planned for 5 years.

F. oxysporum f. sp. oiosri could be isolated from host tissue buried up to 24" depth in soil after 12 months. Tissues buried in soil at 6" to 24" depth are disintegrating faster as compared to the tissues near the surface of the soil (Table 3).

Table 3. Survival of F. oxysporum f. sp. ciceri in host tissues buried at different depths of soil^a

	_	Observation					
Date of isc	olation	Surface	6*	12"	18"	24"	
1-9-1979	1	2.42	2.68	2.69	2.36	2.66	
	2	1.95	2.06	2.35	2.10	1.85	
	3	+	+	+	+	+	
1-12-1979	1	2.70	2.25	2.59	2.75	2.68	
	2	1.53	0.96	1.33	1.38	0.98	
	3	+	+	+	+	+	
1-3-1980	1	2.71	2.19	2.55	2.48	2.32	
	2	1.69	1.35		only in traces		
	3	+	+	+	+	+	
1 -6-19 80	1	2.58	2.42	2.34	2.47	2.49	
	2	1.05	0.39	0.47	only in traces	0.12	
	3	+	+	+	+	+	

l Original weight of root pieces

VI. STUDIES ON INOCULUM POTENTIAL

Density of inoculum distributed throughout the soil of a field having the same cropping and cultivation history will show an inoculum potential. The infectivity characteristics of such a population can be assessed by suitable sampling method. Such studies should be able to predict the disease risk that will attend the planting of a susceptible crop. There is a critical level of inoculum potential necessary for initiation of a progressive infection in a host plant. Frequent obstacle encountered by a root disease worker is the task of reproducing the disease by inoculation. The failures can be ascribed to an

² Weight of root pieces at the time of isolation

³ Isolation results (+ yes; - no)

Roots were buried in soil on 1 June 1979.

inoculum potential inadequate to establish infection. Thus the information on inoculum potential in case of chickpea wilt will be useful to create the disease under artificial condition as well under natural condition in the field.

Soil was collected from the field (Vertisol) which did not have the history of chickpea plantings. To ensure further that it was free from F. oxysporum f. sp. oiceri, highly susceptible cv. JG-62 (collected from healthy plants) was sown in the soil in pot and watched for 40 days. All the plants were healthy. Seeds were treated with 2.5% sodium hypochlorite for 2 minutes.

The soil (10 kg) was filled in 30-cm earthen pots in a net-house. Inoculum grown on sand-maize meal medium for 15 days at 25°C was mixed in the soil at different rate. For each level of inoculum, 3 pots were used. Inoculum was mixed thoroughly in the soil. After 4 days soil samples were collected from the surface of each pot to determine the Fusarium propagules. Immediately after removing soil samples, 10 seeds of the susceptible cultivar JG-62 were sown in each pot. Final observations on wilt incidence were recorded 40 days after sowing (Table 4a).

Table 4a. Effect of the quantity of inoculum added to soil on wilt incidence in chickpea

Inoculum added in soil (g/kg)	Propagules/g ^a of soil	Percent ^b wilt	No. of days for wilting
100.0	73,000	100	17
50.0	43,000	100	17
33.3	21,500	100	17
25.0	13,500	100	17
20.0	14,000	100	19
16.6	13,000	100	19
14.2	12,000	100	19
12.5	6,000	100	19
11.1	5,500	100	19
10.0	4,500	100	19
No inoculum	0	0	-

Average of 4 replications

Soil was also collected from different fields of ICRISAT Center including wilt-sick plots. From each field 100 kg of soil was collected, mixed thoroughly and filled in 3 earthen pots (10 kg/pot). From these pots soil samples were taken to determine Fusarium propagules. The pots were then sown with JG-62 and observations on wilt incidence were recorded.

Final observation taken 40 days after sowing

With 10 g/kg of inoculum (on SPM) added to the soil the mortality of chickpea was 100%. The number of inoculum propagales were 4500/g soil (Table 4a). This experiment was repeated twice and the mortality obtained at the lowest inoculum level (10 g/kg) in soil was complete. Therefore it was decided to conduct the experiment with lower dose of inoculum (Table 4b).

Table 4b. Effect of quantity of inoculum added to the soil on the wilt incidence in chickpea

Inoculum added in soil (g/kg)	Propagules/gm ^a of soil	Percent b wilt	No. of days
25.0	11,000	100	14
20.0	10,500	100	14
16.6	11,000	100	20
14.2	8,000	100	20
12.5	8,500	100	20
11.1	6,000	100	20
10.0	4,500	100	20
5.0	4,000	100	20
2.5	3,000	100	20
No inoculum	0	0	-

Average of 4 replications

These studies (Table 4b) indicated that the lower dose of inoculum (2.5 g/kg) was sufficient to cause 100% wilt in susceptible chickpea cultivar. Only the time required for wilting was 20 days against 14 days at 20 g/kg inoculum level.

The propagules of Fusarium oxysporum f. sp. ciceri in wilt-sick plots (BT-6, M-5, and B-5) ranged from 4000 to 7000/g of soil (Table 5). Our observations during early 2 to 3 years indicated that JG-62 showed 100% mortality within 30 days after planting in wilt-sick plots. In pots, 10 g of inoculum when mixed to the soil gave nearly 4,500 propagules/g of soil. At this level of inoculum, JG-62 gave 100% mortality.

These results indicate that our wilt-sick plots carry the sufficient inoculum potential (certain number of inoculum propagules) in soil, that is when susceptible cultivar like JG-62 is planted it gets killed uniformly in definite time period.

b Final observation taken 40 days after sowing

Table 5. Incidence of chickpea wilt in soil collected from different fields at ICRISAT

S1. No.	Field	Soil type	Crop during 1978-79	Fusarium propagules/ q of soil	Percent wilt	No. of days for wilting
1.	BT-6 A	Vertisol	Chickpea	7,000	100	19
2.	BT-6 C-1	Vertisol	Chickpea	5,500	100	21
3.	BT-6 C-2	Vertisol	Chickpea	4,000	100	25
4.	M-5 A	Vertisol	Chickpea	6,000	100	21
5.	M-5 B	Vertisol	Chickpea	5,000	100	27
6.	B-5 A	Vertisol	Pigeonpea	500 ر5	100	29
7.	B-5 B	Vertisol	Pigeonpea	4,500	100	27
8.	BT- 5	Vertisol	Sorghum	0	0	-
9.	BW-1	Vertisol	Pigeonpea + Maize	0	0	-
10.	R-2	Alfisol	Pigeonpea	0	0	•
11.	RA-9	Alfisol	Pigeonpea	0	0	•

Serial nos. 1-5 are wilt-sick plots

VII. INHERITANCE STUDIES

We are cooperating with ICRISAT chickpea breeders in screening the breeding material to get information on inheritance of wilt resistance. The screening is done in wilt-sick pots in a net house.

In each pot (45 cm) 10 seeds of test line were grown along with susceptible cv. JG-62. The surviving plants were allowed to grow and seed from individual plants was collected. The results are presented in Table 6 and will be interpreted by the breeders.

Table 6. Study on the inheritance of chickpea wilt resistance

Sl.		No. of	No. of wilted	Percent
No.	Particulars	plants	plants	wilt
1	2	3	4	5
	BC-1 F ₂			
	K-4 x (NEC-802 x K-4)			
1.	Selection No.3	2	2	100
2.	5	7	7	100
3.	6	2	2	100
4.	7	No germ	ination	
5.	8	ň		
6.	9	1	1	100

contd.

Table 6. Contd.

1	2		3	4	5
7.	Selection No.10		1	1	100
8.	11	No	germination		
9.	12		3	3	100
10.	13		2	2	100
11.	14	No	germination		
12.	15		8	8	100
13.	17	No	germination		•
14.	19		1	1	100
15.	21	No	germination		
16.	23		3	3	100
17.	25	No	germination		
18.	26		5	5	100
19.	28	No	germination		
20.	30		2	2	100
21.	32		4	4	100
22.	34		7	7	100
23.	36		3	3	100
24.	37		2	2	100
25.	39		1	1	100
26.	40		2	2	100
27.	43		4	4	100
28.	44		4	4	100
29.	45		1	1	100
30.	46		4	4	100
31.	47	No	germination		
32.	48		4	4	100
33.	49		3	3	100
	BG-203 x (BG-203 x WR-315)				
34.	Selection No.31		4	1	25.00
35.	42		2	1	50.00
36.	33	1	.4	8	57.14
37.	34	1	.1	2	18.18
38.	35		6	3	50.00
39.	36		.7	5	29.41
40.	37		.0	8	80.00
41.	38		6	2	33.33
42.	39		.0	5	50.00
43.	40	1	.0	3	30.00
44.	43		3	0	0.00
45.	44		4	1	25.00
46.	45		8	4	50.00
47.	46		4	2	50.00
48.	47		4	1	25.00
49.	48		1	1	100.00
50.	49		.2	4	33.33
51.	50		5	2	40.00
52.	51		.0	8	80.00
<u>53.</u>	52	1	.7	9	52.94

contd.

Table 6. Contd.

 2	3	4	5
Selection No.54	11	3	27.27
55	6	5	83.33
56	13	2	15.38
57	8	ī	12.50
58	17	8	47.05
60	8	2	25.00
Parents	-	_	
<u>K-4</u>	10	10	100.00
NEC-802	10	4	40.00
BG-203	10	6	60.00
WR-315	10	1	10.00
1977-78 diallel F1 WR		_	
CPS-1 x P-1353	5	2	40.00
CPS-1 x P-6099	5	0	0.00
CPS-1 x P-436	5	5	100.00
P-1353 x P-6099	2	1	50.00
P-6099 x P-436	В	8	100.00
1977-78 diallel F2 WR			
CPS-1 x P-1353	88	44	50.00
CPS-1 x P-6099	93	28	30.10
CPS-1 x P-436	83	62	74.69
P-1353 x P-6099	49	19	38.77
P-1353 x P-436	81	61	75.30
P-6099 x P-436	73	69	94.52
Others			
F ₁ WR-315 x P-1353	10	1	10.00
F2 WR-315 x P-1353	65	26	40.00
Parents			
CPS-1	10	0	0.00
P-1353	6	0	0.00
P-6099	10	1	10.00
P-436	9	9	100.00
WR-315	10	0	0.00

VIII. YIELD TRIAL

In order to study the yield potential of wilt resistant lines, a field trial was conducted at ICRISAT Center and at Hissar sub-center with the help of breeders. Sixteen wilt resistant lines and 4 checks were included. The trial was planted in 4 replications in M-4 field. Each plot had 4 rows of 4 m length at 30 cm apart. 40 seeds were sown in each row. The results are presented in Tables 7 and 8.

Table 7. Yield testing of wilt resistant chickpea lines at Hyderabad (arranged in ascending order)

Cultivar name-	Average yield/	Yield/hectare	Average yield	
Cultivar names	plot (g)	(calculated)(kg)	plant (g)	
H-208 (check)	87.5	182.0	1.4	
BG-203 (check)	192.5	400.4	1.9	
P-289	270.0	561.6	2.0	
P-1265	337.5	702.0	2.5	
P-165	360.0	749.8	2.7	
P-517	365.0	759.2	2.8	
P-4116-1	405.0	842.4	3.6	
BG-212	441.2	917.6	3.4	
P-678	447.5	930.8	3.3	
P-1270	452.5	941.2	3.4	
P-1353	460.0	956.8	3.4	
CPS-1	470.0	977.6	3.5	
WR-315	477.5	993.2	3.7	
BDN-9-3 (check)	505.0	1050.4	4.3	
P-6099	512.5	1066.0	4.0	
NEC-790	601.2	1250.4	5.0	
Annigeri (check)	663.7	1380.4	5.1	
JG-74	682.5	1419.6	5.2	
No. of treatments	18. No. of rep	lications 4		
Average yield of	the			
experiment (plo	t) 429.0 g	(plant) 3.4	
CV	23.0		21.1	
LSD (0.05)	140.0		1.0	

Table 8. Yield testing of wilt resistant chickpea lines at Hissar (arranged in ascending order)

Cultivar names	<pre>Average yield/ plot (g)</pre>	Yield/ha (calculated) (kg)	Cultivar names	Average yield/ plot (g)	Yield/ha (calculated) (kg)
P-1265	51.25	106.7	WR-315	138.75	288.6
Annigeri	57.50	119.6	JG-74	188.75	392.6
BDN-9-3	90.00	187.2	NEC-790	240.00	499.2
P-517	95.00	197.6	P-4116-1	246.00	511.6
P-1270	105.00	218,4	P-6099	250.00	520.0
CPS-1	115.00	239,2	BG-212	280.00	582.4
P-165	130.00	270.4	P-289	280.00	582.4
P-1353	136.25	283.4	H-208	303.75	631.8
P-678	137.50	286.0	BG-203	505.00	1050.4

Average yield of the experiment (plot) 186.09 g CVN 61.86 LSD (0.05) 163.37 Of the 18 lines tested, NB-203, H-208, NDM-9-3, and Annigeri were the checks. Among the top high yielding cultivars; JG-74, and MEC-790, were the wilt resistant lines. JG-74 was the highest yielder among 18 cultivars at Hyderabad. Annigeri and JG-74 were also significantly superior over others on the basis of yield/plant. Incidentally H-208 and BG-203 were poor in yield performance (yield/plot and yield/plant).

At Hissar, BG-203 was significantly superior over other cultivars in yield performance. Low yield in general was because of the poor plant population at Hissar.

PROJECT: CP-PATH-2(78): STUDIES ON STEM AND ROOT ROTS OF CHICKPEA

I. SUMMARY

- 1. Rhisoctonia bataticola, the dry root rot fungus, survives in the soil on infected tissues for at least 24 months.
- 2. Blotting paper technique was developed to identify resistance to the dry root rot of chickpea. Clear differences between susceptible and resistant lines of chickpea were observed. Chickpea wilt-resistant cultivars were screened using the blotting paper technique. The infected roots were scored on 1 to 9 scale.
- 3. Following this technique ICC-554 and -6926 showed very little infection to roots and were considered resistant to the dry root rot.
- 4. Of the 168 lines tested, 39 lines showed 3 rating (slight infection) to the dry root rot.
- 5. During the year under report 354 wilt promising lines were sown in a multiple disease sick plot. Of these, 195 lines showed less than 10% wilt/root rots.
- 6. Nine entries from Kanpur were found promising. Of them PPK-1 has been included in the International Chickpea Root Rots/Wilt Nursery-GL-779 from Gurdaspur was found promising. MCK-4, 9, 10, 15, 31, 40, 43, 51, 74, and 83 from New Delhi showed less than 20% mortality in the multiple disease sick plot.
- 7. From the GIET entries, ICCC-18, P-324, JG-2260, JG-1259 were promising. From GCVT entries, ICCC-10 was found resistant.
- 8. The multiple disease sick plot had the following pathogens, F. oxysporum f. sp. ciceri, Rhizoctonia bataticola, Sclerotium rolfsii, R. solani, F. solani and white root rot fungus in that order.
- 9. Sclerotia of Sclerotium rolfsii (collar rot fungus) lost their viability in 3 months when buried in soil at 5 cm depth. While those stored in laboratory were viable.

II. INTRODUCTION

The project was initiated in January 1978 with the following objectives:

- Collect more precise information on the prevalence of stem and root rots in the chickpea growing areas,
- 2. Study the etiology of pathogens leading to the understanding of epiphytology of these diseases, and
- 3. Develop efficient techniques to screen for resistance.

After the chickpea wilt, dry root rot caused by Rhisostonia bataticola is widely prevalent in chickpea growing areas in the semi-arid tropics. During the year under report we have made progress in developing a laboratory screening technique for this disease and attempted to screen wilt-promising lines. For field screening we had to depend on natural incidence in the multiple disease sick plot.

The studies were also initiated on the surviving ability of sclerotia of Sclerotium rolfsii (the collar rot pathogen).

III. DRY ROOT ROT (RHIZOCTONIA BATATICOLA)

Survival of Rhizoctonia bataticola on host debris

Since April 1978 we were attempting the isolations of *R. bataticola* from the infected stems and roots buried in Vertisol-filled earthen pots and from the material kept in laboratory. Isolations were attempted every month starting from April 1979 on the CMR medium described by Meyer et al (Phytopathology 68: 613-620, 1973).

The composition of the medium is as follows:

Polished rice	10	g
Agar	20	g
Chloroneb	300	mg
Mercuric chloride	7	mg
Rose bengal	90	mg
Streptomycin sulphate	40	mg
Potassium penicillin	60	mg

Polished rice is boiled for 5 min in one litre water and strained through cheese cloth. Agar is added and the medium is autoclaved. The remaining ingredients are mixed after autoclaving and the pH adjusted to 6.0 with lactic acid.

The results obtained so far indicate that the fungus is able to survive in infected tissues for at least 24 months. The experiment is continuing.

IV. SCREENING TECHNIQUE

During the last two years several experiments were conducted to develop a screening technique which will be reliable and efficient in showing different degrees of resistance/susceptibility among chickpea cultivars. It was observed that high temperature (28 to 35°C) and water stress are most important factors in the disease development. Stage of plant growth (5 to 7-day-old seedlings) and use of fresh culture (5-day-old) are important in creating the disease under artificial conditions.

The following technique has been developed.

Blotting paper technique

1. Multiplication of inoculum

Inoculum is multiplied on potato-dextrose broth, 100 ml in 250 ml flasks for 5 days at 25°C. Growth of the fungus (mycelial mat) from 2 flasks is removed and blended in Waring blendor for a minute with 100 ml of sterile distilled water.

2. Raising of seedlings

Seed of each test lines is surface disinfected with 2.5% solution of sodium hypochlorite (5 min) before sowing in autoclaved sand. Seedlings of 5-day-old are carefully removed from sand, washed in running water and used for inoculation. Normally 30 seedlings are inoculated for each cultivar. BG-212 and Chafa which are highly susceptible are used as checks.

3. Inoculation

Roots of the seedlings are dipped into the inoculum. The seedlings are then spread on fold-blotting paper in such a manner that roots along with cotyledons are covered. Paper towels are moistured before use.

4. Incubation

Inoculated seedlings in paper towel are then incubated at 35°C for 8 days. Paper towels are moistured daily. After 8 days of incubation seedlings are examined for the extent of root damage. The results are reported below.

5. Disease rating (Dry root rot)

We are rating the seedlings on the basis of 1-9 rating where 1 is healthy and 9 is completely rotted root. The interpretation of the scale is as follows:

- 1 Clean root, no infection
- 3 Infection slight, small lesions on few roots
- 5 Infection moderate, lesions on 50 percent roots
- 7 Infection severe, extended lesions on about75 percent roots, shoots remain green
- 9 Completely rotted roots, extended lesions on all roots, shoots show yellowing and drying

The wilt promising lines were screened in the laboratory using paper towel technique. The disease reaction was rated as follows.

6. Wilt-nomising lines screened against Rhisoctonia bataticola (Dry root rot)

- 2 ICC-554, 6926
- 3 ICC-444, 537, 999, 1910, 1913, 1918, 2086, 2461, 2874, 3181, 3392, 3428, 4716, 4902, 4994, 5901, 6081, 6366, 6411, 6455, 6501, 6570, 6608, 6668, 6687, 6772, 6816, 6840, 6939, 7681, 8971, 9018, 9023, 9042, 10466, 10500, 10539, 10630, 11550.
- 5 ICC -338, 438, 519, 1376, 1443, 1611, 2354, 2450, 2566, 2616, 2774, 2858, 3439, 4552, 4847, 5006, 6384, 6474, 6502, 6743, 6800, 6874, 6815, 7254, 7489, 8585, 8979, 9025, 9032, 9033, 9042, 9112, 10803, 10823, 11531.
- 7 43, 202, 267, 391, 858, 1338, 1450, 2862, 2872, 2935, 2950, 4519, 4843, 4850, 6344, 6383, 6817, 6880, 7248, 8222, 8612, 8933, 8988, 9030, 9043, 10130, 10809, 11551, 11552.
- 9 229, 434, 595, 658, 867, 2089, 2354, 2616, 2660, 2803, 2883, 2917, 2943, 3058, 3117, 3354, 3513, 3528, 3531, 3533, 3539, 3782, 4485, 4994, 6098, 6381, 6385, 6386, 6411, 6440, 6488, 6491, 6494, 6671, 6680, 6711, 6730, 7254, 7336, 7481, 8166, 8446, 8454, 8499, 8622, 8980, 8982, 8985, 9006, 9021, 9028, 9032, 9034, 9035, 9036, 9039, 9040, 9041, 9055, 9103, 9117, 10104, 10394

Susceptible check BG-212 (ICC-11088)

V, FIELD SCREENING

We have developed a multiple disease sick plot wherein different soil-borne pathogens have been encouraged to build up through incorporation of dead plant debris every year. Chickpea lines found promising against the wilt were planted in multiple disease sick plot. In addition, GCVT and GIET entries were planted. This year we received the entries from New Delhi, Kanpur, and Gurdaspur which were also planted in this plot.

A. Wilt-promising lines from 1978-79

This year 354 wilt promising lines were sown in a multiple disease sick plot. They were sown in 2 rows of 4 meters. After every 2 test rows JG-62 (susceptible check) was sown. Of these, 195 lines showed less than 10% wilt and root rots. These were:

ICC-102, 104, 182, 229, 267, 338, 391, 434, 438, 444, 449, 460, 516, 519, 554, 595, 606, 658, 858, 867, 999, 1338, 1376, 1450, 1611,

1891, 1910, 1913, 1918, 2072, 2083, 2086, 2089, 2104, 2200, 2328, 2337, 2354, 2450, 2461, 2566, 2616, 2660, 2664, 2774, 2812, 2835, 2858, 2862, 2883, 2950, 3058, 3099, 3103, 3181, 3310, 3354, 3392, 3396, 3415, 3428, 3439, 3513, 3528, 3531, 3534, 3539, 3684, 3782, 4129, 4485, 4519, 4552, 4716, 4843, 4847, 4850, 4902, 4918, 4920, 4994, 5006, 5864, 5901, 6081, 6098, 6366, 6381, 6383, 6384, 6385, 6411, 6440, 6455, 6460, 6474, 6480, 6488, 6489, 6491, 6494, 6501, 6502, 6570, 6608, 6630, 6643, 6668, 6671, 6687, 6711, 6730, 6743, 6761, 6772, 6774, 6800, 6815, 6816, 6817, 6874, 6880, 6926, 6939, 7111, 7248, 7254, 7336, 7481, 7489, 7681, 8166, 8170, 8222, 8612, 8622, 8933, 8971, 8979, 8980, 8982, 8985, 8988, 8999, 9001, 9003, 9006, 9018, 9021, 9023, 9025, 9028, 9029, 9030, 9031, 9032, 9033, 9034, 9035, 9037, 9039, 9041, 9042, 9043, 9055, 9085, 9103, 9112, 9127, 10104, 10130, 10382, 10384, 10394, 10397, 10399, 10492, 10500, 10514, 10517, 10536, 10537, 10539, 10630, 10662, 10802, 10803, 10809, 10823, ICCC-10, DA-1, RAVP-52, GW-9, F-496, and BG-228.

B. Entries received from other locations

1. New Delhi

Of the 100 lines received from Dr. H.K. Jain, Director, IARI, New Delhi, none was found resistant. However, MCK-4, 9, 10, 15, 31, 40, 43, 51, 74, 83 showed less than 20% wilt and root rots.

2. Kanpur

Of the 23 wilt-resistant lines (at Kanpur), KN-4, KW-5, KW-2B, KW-17B, BA-1, GW-6, GW-3-1, PPK-1, and PPK-2 showed less than 10% wilt and root rots.

3. Gurdaspur

Of the 7 lines, GL-779 showed less than 10% wilt and root rots.

4. Badnapur

Of the 9 lines received, none was found promising.

C. GIET

Of the 35 entries planted, ICCC-18, P-324, JG-2260, and JG-1259 were promising.

D. GCVT

Of the 12 entries planted, only ICCC-10 showed less than 10 percent mortality.

VI. PERIODIC ISOLATIONS FROM WILMED/DRIED PLANTS COLLECTED FROM MULTIPLE DISEASE SICK PLOT

To monitor the presence of different root pathogens from October through Pebruary (chickpea season at Hyderabad) we made periodic isolations from diseased plants. The results have been presented in Table 9.

Table 9. Periodic isolations from wilted/dried plants of chickpea collected from multiple disease sick plot^a

Date of collection	F. oxysporum f.sp. ciceri	R. bata- ticola	S. rolfsii	R. solani	F. solani	White root rot fungus
26-11-1979	43.85	4.38	37.71	3.50	0.87	
14-12-1979	47.60	24.41	10.46	4,65	3.48	5.81
4-1-1980	65.00	25.00	_	6.00	1.00	-
25-1-1980	60,00	35.00	-	•	3.00	•
10-2-1980	37.00	62.00	-	-	•	•

^aFigures are percentage of isolations

The wilt fungus F. oxysporum f. sp. oiosri was predominant all through the season except in February when the day temperature rose close to 30° C (Table 10). That time R. bataticola became more dominant. S. rolfsii (Collar rot) was very much active in the early stage of plant growth. Other pathogens present were, R. solani, F. solani and the white root rot fungus.

Table 10. Ambient temperature data from November 1979 through February 1980

Standard week	Dates	Average Maximum	temperature (C) Minimum	Rainfall (mm)
44	29 October-4 November 1979	30.2	19.7	1.0
45	5 November-11 November	29.0	19.8	61.0
46	12 November-18 November	27.3	20.3	10.0
				•
47	19 November-25 November	29.7	17.7	0.4
48	26 November-2 December	28.1	19.0	7.7
49	3 December-9 December	27.9	15.2	0.0
50	10 December-16 December	27.5	14.5	0.0
51	17 December-23 December	28.2	17.7	0.0
52	24 December-31 December	27.4	12.2	0.0
1	1 January-7 January 1980	28.7	16.4	0.0
2	8 January-14 January	28.0	14.6	0.0
3	15 January-21 January	28.4	13.8	0.0
4	22 January-28 January	30.1	15.8	0.0
5	29 January-4 February	30.1	16.0	0.0
6	5 February-11 February	31.1	17.5	0.0
7	12 February-18 February	32.0	17.7	0.0
8	19 February-25 February	33.7	18.2	0.0
9	26 February-4 March	35.0	17.7	4.0

These isolations gave us a clear picture of the performance of wilt-promising lines planted in multiple disease sick plot against at least 2 pathogens, S. rolfsii and R. batatioola.

VII. SURVIVAL OF SCLEROTIUM ROLFSII (COLLAR ROT PATHOGEN) IN SOIL

Selerctium rolfsii causes the collar rot in chickpea during seedling stage with high soil moisture. The disease appears whenever the chickpea field is irrigated. The fungus forms the black, small mustard-seed-like sclerotia in culture as well as on host tissues. The sclerotial bodies are resistant and it is presumed that the fungus survival depends on these bodies. An experiment was conducted to know the ability of the fungus to survive in the soil and in laboratory through the sclerotia.

The fungus was isolated from diseased chickpea plants during December 1979 on Potato-dextrose-agar (PDA). Pathogenicity was proved by inoculating 10-day-old chickpea seedlings (JG-62) grown in plastic pots (15 cm) containing autoclaved soil. Sclerotia developed on PDA were placed at the base of the plants. Pots were watered adequately. Within 10 days after inoculation diseased plants were observed. Fungus was reisolated. It is being multiplied and maintained on PDA.

S. rolfsii was grown on PDA in petri plates for 15 days at 25°C. Sclerotia were harvested from medium, aseptically air-dried and kept in sterilized petri plates. Since sclerotia had to be buried in soil, the surface disinfection after their removal from soil was essential. For this purpose an experiment was conducted to study the effect of surface disinfectants on germination of sclerotia. Mercuric chloride solution (1:1000) and sodium hypochlorite (2.5%) were used to disinfect the sclerotia. After disinfection they were kept on PDA in petri plates and incubated at 25°C for 10 days.

There was no adverse effect on germination of sclerotia treated with mercuric chloride (1:1000) for 8 to 10 seconds and washed subsequently twice with sterilized water and sodium hypochlorite (2.5%) for 2 to 3 minutes. Also the study indicated the viability of sclerotia harvested from medium (Table 11).

Table 11. Effect of surface disinfection on the germination of sclerotia of S. rolfsii

Treatment	No. of sclerotia kept on PDA	No. of sclerotia germinated	Percent germination	
Mercuric chloride (1:1000)	20	20	100	
Sodium hypochlorite (2.5%)	20	20	100	
Sterilized water	20	20	100	

In our experiments we decided to use sodium hypochlorite as surface disinfectant.

Sclerotia (nearly 30) were enclosed in small nylon mesh pieces and buried in Vertisol at 5 cm depth in pots. These pots were kept open so as to expose to field conditions. One lot of sclerotia was kept at room temperature (25-28°C) in petri plates. The viability of sclerotia was tested every month (Table 12).

Table 12. Viability of Sclerotium rolfsii in soil and in laboratory

From soil				From		
Date of isolation	No. of sclerotia	No. of sclerotia germinated	Percent germi- nation	No.of sclerotia	No. of sclerotia germinated	Percent germi- nation
13-4-1980	20	20	100	20	18	90
13-5-1980	25	23	92	25	25	100
13-6-1980	25	0	0	25	25	100

Sclerotia were buried in soil or kept in the laboratory on 13-3-1980.

The studies indicated that sclerotia removed from soil after 3 months lost their viability while those from laboratory were viable with 100% germination on PDA.

VIII. MISCELLANEOUS STUDIES

A. Screening against Colletotrichum blight

1. Laboratory screening

Germplasm lines that were found promising under natural conditions against the Colletotrichum blight last year (1978-79) were tested for their reaction using Isolation Plant Propagator, as being done in case of Ascochyta blight.

Colletotrichum dematium isolated from chickpea at ICRISAT was used. Inoculum was multiplied on potato-dextrose-broth (PDA) (100 ml in 250 ml flask) for 15 days at 25°C. The growth from 2 flasks was mixed in 500 ml sterilized distilled water and that served as inoculum. Promising lines selected from field (with 5 rating on a 9-point scale) were grown in a plant propagator. Ten-day-old seedlings were sprayed with the inoculum and covered for 5 days with plastic covers. In addition 19 Ascochyta blight-promising lines were screened.

The disease developed within 4 days, killing most of the lines. Two lines which showed 1 (ICC-4925) and 5 (ICC-8923) rating on 9 point

scale after 4-day incubation period also succumbed to disease ultimately. All 19 lines resistant to Ascochyta blight were also susceptible. A list of susceptible lines (rating 9) is given below:

Colletotrichum blight-promising lines: ICC-3, 1611, 1903, 1909, 2223, 2267, 2619, 3032, 3230, 4925, 4939, 4948, 5035, 5098, 5107, 5119, 5127, 5165, 6213, 6261, 6319, 6671, 6743, 6805, 6819, 6901, 7287, 7359, 7722, 8284, 8542, 8920, 8923, 8927, 9150, and 10259.

Ascochyta blight-promising lines: ICC-150, 280, 377, 931, 1009, 1465, 1903, 1911, 2153, 2156, 2160, 2237, 3259, 3277, 3330, 6067, 7513, 7514, 7520, and Pb-7 (check).

2. Field screening

The Microbiology sub-program had sown 416 chickpea entries from the germplasm in Alfisol (R-2) on 31 August 1979 to study nodulation. These cultivars were evaluated during October-November for the Colletotrichum blight resistance.

Most of these cultivars were affected by blight during early October and disease was uniform in the field. Within 2 months after sowing most of the cultivars succumbed to disease.

There was no cultivar with 1-3 rating. Lines, ICC-341, 693, 893, 8920, and K-1170 showed a rating of 5.

3. Pathogenic behaviour of C. dematium and C. capsici

C. dematium and C. capsici are the two species reported to cause chickpea blight in India. C. dematium was isolated from infected leaves of chickpea collected at ICRISAT, Hyderabad and also from summer plantings at Taparwaripura (Kashmir) in 1978. C. capsici was isolated in 1975 from infected leaves of chickpea collected from Coimbatore. Both species were identified by the Commonwealth Mycological Institute, London.

To study their pathogenic behaviour, 3 chickpea cultivars; JG-62 Pb-7, and 850-3/27 were inoculated artificially using a Isolation Plant Propagator.

The fungi were grown separately on potato-dextrose broth (100 ml medium in 250 ml flask) for 15 days at 25°C. The growth from 2 flasks was removed and mixed in 250 ml sterilized water. The inoculum was sprayed on 10-day-old seedlings and seedlings covered with plastic tops to provide humidity. There were 5 seedlings in each pot and 5 pots were sprayed with each fungus. The reactions were recorded on a 9-point scale progressively after every 3 days (Table 13).

The experiment indicated that C. demation was aggressive and killed all the seedlings in 3 cultivars within 9 days. The disease intensity in seedlings inoculated with C. capsioi was low even after 12-day incubation.

OCROS	AT Library
1 29	02085

Table 13. Reaction of three chickpea cultivars to species of Colletotrichum

Cultivar		C. dematisen		C. capeici				
	3d	6d	94	124	3d	6d	9d	128
JG-62	5	7	9	-	3	5	5	7
Pb-7	5	7	9	-	3	5	7	9
850-3/27	3	7	9	-	•	5	7	7

B. Leaf diseases

1. Botrytis grey mold

The disease is prevalent in chickpea growing areas of Uttar Pradesh and Bihar. It has also been reported from Australia.

The culture of the fungus was collected from Kanpur during a visit on 7 February 1980.

(a) Pathogenicity

The fungus was multiplied on potato-dextrose-broth at 25°C for 15 days in 250 ml flask containing 100 ml medium. The growth from one flask was blended with 100 ml sterilized water in a blendor and 10-day-old seedlings (JG-62) grown in Isolation Plant Propagator were sprayed with the inoculum suspension and covered for increasing humidity.

(b) Symptoms

The symptoms on leaves were observed 2 days after inoculation. The leaflets were discoloured, discoloration started along the margin or at the tip of the leaflet. There was a dropping of terminal branch. On 5th day, the leaflets turned grey in colour. Fungal growth could be seen on infected tissues. The infection spread further to branches and main stem and rotting was evident. No definite lesions were seen. The infected portions in humid condition produced a lot of erect hairy sporophores and one-celled hyaline conidial mass.

The fungus was successfully reisolated. It is being maintained in pure culture on PDA.

2. Stemphyllium leaf spot

Stemphyllium species was isolated from the diseased chickpea leaves brought from Dholi (Bihar) during the last week of March 1980. The pathogenicity of the fungus was proved. The disease spots were identical to those observed in field. The Stemphyllium leaf spot is reported to be prevalent in Bihar and West Bengal.

3. Alternaria leaf spot

Another leaf blight pathogen Alternaria alternata was brought in pure culture from chickpea diseased specimens brought from Sehore (Madhya Pradesh). The disease was severe in December 1979 in that area. The pathogenicity of the fungus was proved. The fungus is being maintained in pure culture in the laboratory.

PROJECT: CP-PATH-3(78): STUDIES ON CHICKPEA STUNT AND OTHER VIRAL DISEASES

I. SUMMARY

- 1. Work on chickpea stunt was mainly confined to screening of germplasm and breeding materials for resistance.
- 2. Screening was done in a plot at Hissar in the Haryana State of India, where the natural incidence of the disease is high. To enhance the natural incidence of the disease, advance planting of hosts of pea leaf-roll virus and its aphid vectors and susceptible chickpea cultivar was undertaken to create a 'reservoir' and have 'spreader' hosts, respectively.
- 3. Out of a total of 1398 germplasm lines planted at Hissar by the Genetic Resources Unit of ICRISAT, 27 and 22 lines with 0-7% and 8-15% incidence, respectively were identified.
- 4. Of the 21 germplasm lines that were selected during 1976-77 season and tested for the past 3 years, 18 showed less than 10% infection and were considered to be highly promising.
- 5. Of the 22 germplasm lines that were selected during 1977-78 season and tested for the past 2 seasons, five lines showed less than 10% infection and were considered to be promising.
- 6. Of the 153 lines that were selected in the last season, 19 and 41 lines showed no or less than 10% infection, respectively.

 These will be tested further.
- 7. Of the 38 advanced crossing block entries which were tested last year also, 11 and 15 lines showed no and less than 10% infection, respectively and were considered promising.
- 8. Of 37 advanced Ascochyta blight-resistant lines, 9 and 14 lines showed no and less than 104 infection, respectively.
- 9. In the preliminary screening only one line out of the 37 Ascochyta blight-resistant lines tested was found promising.
- 10. Some of the wilt and root rot resistant lines were found promising to chickpea stunt.
- 11. Three ICCC lines showed less than 10% infection for two seasons.
- 12. Nine out of 36 GIET lines tested were found to show no infection.

 Fourteen lines had less than 10% infection.
- 13. Two of the 12 GCVT lines tested showed less than 10% infection.
- 14. Twenty-two lines that were found promising during the last 3 seasons were sent for multilocation testing.

- 15. Seven F_2 populations involving resistant parents were screened and selections made.
- 16. Early plantings were found to develop more infection than the later plantings.
- 17. Low plant populations (8 plants/m²) both under sprayed and unsprayed conditions had higher disease incidence than at higher plant populations (33 and 67 plants/m²).

II. INTRODUCTION

During the year under report, work on chickpea stunt was mainly confined to the screening of germplasm lines and breeding materials for resistance. Lines that were found resistant during the last two seasons were sent for multilocation testing.

III. SCREENING NURSERY

Screening of the materials was carried out in a 0.75 ha plot at Hissar on the campus of Haryana Agricultural University in northern India. One month before planting of the test materials (15 September 1979) a mixture of the important hosts of pea leaf-roll virus (PLRV) and aphid vectors (Peas, Broadbeans, Alfalfa, Berseem, Brassica sp., Cowpeas, Beans, Lentil, Chickpea, Groundnut, Mungbean, Urdbean) was planted in and around the field to serve as reservoirs for both the virus and vectors. The field was ridged 0.75 cm apart. The distance between two rows of 'reservoir' hosts was 9 m. Fifteen days later, WR-315, a susceptible cultivar of chickpea was planted between these rows to serve as spreader rows. Planting of the test materials was done 15 days later and WR-315 was planted after every two test rows to serve as 'indicator rows' (Fig. 1). Observations on the incidence of disease were recorded twice, first 3 months after planting and second at the time of maturity.

IV. SCREENING FOR RESISTANCE

A. Germplasm

A total of 1398 lines planted by the Genetic Resources Unit of ICRISAT at Hissar were observed for the incidence of stunt. Twenty-seven lines with 0-7% and 22 with 8-15% incidence were identified. The lines with 0-7% incidence were; ICC-614, 624, 628, 660, 827, 881, 947, 1029, 1022, 1023, 1216, 1404, 1416, 1447, 1578, 1821, 1868, 1962, 1972, 1974, 2254, 2258, 2264, 2265, 2304, 2306, and 8930. The lines with 8-15% incidence were; ICC-845, 1044, 1061, 1130, 1421, 1563, 1576, 1581, 1817, 1867, 1971, 1973, 1977, 2009, 2019, 2175, 2227, 2229, 2273, 2292, 2294, and 2307.

The seed of the above lines was collected and will be tested in the stunt screening nursery next season.

B. Advanced germplasm lines

1. 1976-77 selections

The results of screening of twenty-one lines that were selected in 1976-77 season and screened in the subsequent two seasons are presented in Table 14. One line, ICC-3133 did not show any infection. Of the remaining 20, except three, ICC-1003, -4869, and -8252 which showed more than 10% all others showed less. These lines have now been tested for 4 seasons and considered to be highly promising.

Table 14. Results of advanced screening of chickpea stunt resistant lines (1976-77 selections) to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

Lines	Total plants	Infected plants	Percent infection
ICC-1003	31	6	19.3
-2233	36	2	5.5
-2 385	45	1	2.2
-2430	44	3	6.8
-2925	29	1	3.4
-3034	36	2	5.5
-3133	42	0	0.0
-3718	34	3	8.8
-3735	34	1	2.9
-4869	39	4	10.2
-6433	47	2	4.2
-6934	41	3	7.3
-8252	39	4	10.2
-10490	49	2	4.0
-10495	47	1	2.1
-10508	46	1	2.1
-10586	47	2	4.2
-10587	36	2	5.5
- 10592	45	2	4.4
-10594	50	3	6.0
-10800	45	1	2.2

2. 1977-78 selections

The results of screening of 22 germplasm lines that were selected during 1977-78 season and screened in the last season also are presented in Table 15. One line, ICC-2356 did not show any infection.

Four lines, ICC-613, 2617, 7003, and 10597 showed less than 10% infection and these lines will be checked in the next season again. Others showed higher infection.

Table 15. Results of advanced screening of chickpea germplasm lines (selected in 1977-78 season) to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

Lines	Total plants	Infected plants	Percent infection	
ICC-613	51	3	5.8	
-2336	38	13	34.2	
-2341	34	4	11.7	
-2352	35	10	28.5	
-2356	39	0	0.0	
-2362	35	15	42.8	
-2367	36	9	2 5 .0	
-2369	38	6	15.7	
-2617	25	1	4.0	
-3637	38	13	34.2	
-3782	33	14	42.4	
-4094	45	10	23.2	
-6371	33	5	15.1	
-6457	40	7	17.5	
-66 34	39	16	41.0	
-68 96	38	9	2 3. 6	
-70 03	50	4	8.0	
-87 86	45	7	1 5. 5	
-88 56	36	12	33.3	
-8867	35	12	34.2	
-8897	41	6	14.6	
-10597	50	2	4.0	

3, 1978-79 selections

Screening of 153 lines was carried out. Summarised results are presented in Table 16. Nineteen lines, ICC-575, 577, 678, 690, 767, 787, 981, 1067, 1876, 1881, 2090, 2226, 2277, 2534, 2546, 2572, 2604, 2713, and 5012 did not show any infection. Forty-one lines showed 10% or less infection. These were ICC-130, 159, 248, 279, 403, 526, 539, 591, 599, 685, 705, 706, 735, 773, 774, 817, 1112, 1113, 1126, 1404, 1510, 1563, 1564, 1882, 1893, 1963, 2039, 2089, 2092, 2108, 2191, 2212, 2228, 2236, 2267, 2276, 2289, 2292, 2388, 2516, and 2521.

Table 16. Summarised results of preliminary screening of chickpea germplasm lines (selected in 1978-79 season) to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

Percent	infection	No. of lines	Percent lines
	0.00	19	12.4
0.01	10.00	41	26.7
10.01	20.00	31	20.2
20.01	30.00	18	11.7
30.01	40.00	16	10.4
40.01	50.00	14	9.1
50.01	60.00	7	4.5
60.01	70.00	3	1.9
70.01	80.00	4	2.6
80.01	90.00	0	0.0
90.01	99.00	0	0.0
100.00		0	0.0

Total lines 153

C. Crossing block entries

1. Advanced lines

The results of screening of 38 entries from the crossing block selected in 1977-78 season and screened in the last year's stunt nursey are presented in Table 17. Eleven lines, NEC-2368, E-235, RS-11, Coll. -327, P-1774, P-2202-2, P-4353-1, G-24, G-130, G-543, and Pant G-115 did not show any infection. Fifteen lines, NEC-472, NEC-550, NEC-701, NEC-746, NEC-1135, NEC-2296, BG-482, F-61, F-370, P-1072, P-1781, P-2019-1, T-3, Coll.-238, and ICCC-5 showed less than 10% infection. These lines have now been tested for 3 seasons and are considered to be promising.

2. Observations in 1979-80 crossing block nursery

High natural incidence of the disease was observed in the crossing block nursery raised by the breeders. Early and spaced planting of the nursery seemed to have encouraged high natural incidence. Observations were made to identify the most susceptible and promising lines. The lines that showed high susceptibility (> 50% incidence) were Phule-G-3, Phule-G-4, Annigeri, P-9800, P-2591, Giza, No.501, C-2201, P-2530, 35168, 1400, K-1170, K-1174, K-1184, K-1189, K-1258, K-1286, K-1481, K-56567, P-9847, BDN-9-3, NEC-139, NEC-329, K-1480, Kourosh, L-534, NEC-10, NEC-694, NEC-1604, NEC-1640, NEC-1663, Caina, CPS-1 Ponaflar, and NEC-2148. The lines that showed no incidence or less than 10% infection were ICC-4, H-208, 850-3/27, 73111-8-2-B-BP x

 $(850-3/27 \times H-208)$, JN-485, JN-2292, GL-629, HMS-8, HMS-10, L-559, 7378-18-5-2H-BP \times (L-550 \times H-208), Coll.-327, T-3, NBC-2296, NEC-2404 and No.501. These lines will be tested in the next year's screening nursery.

Table 17. Results of advanced screening of chickpea crossing block entries to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

	Total	Infected	Percent
Lines	plants	plants	infection
NEC-177	43	12	27.9
-240	45	13	28.8
-472	48	2	4.1
-550	45	3	6.6
-555	47	6	12.7
-701	46	4	8.6
-746	45	3	6.6
-1126	43	9	20.9
-1135	46	1	2.1
-2 29 6	46	2	4.3
-2368	43	0	0.0
H-208	37	9	24.3
ห-556-1	35	5	14.2
H-208 x Pant G-114	33	5	15.1
H-556-1	42	5	11.9
BG-203	35	4	11.4
BG-482	28	2	7.1
C-235	39	0	0.0
C-104	37	5	13.5
F-61	39	1	2.5
F-370	38	1	2.6
F- 378	3 5	5	14.2
RS-11	45	0	0.0
K-468	48	5	10.4
Coll238	37	1	2.7
Coll 327	44	0	0.0
P-1092	39	2	5.1
P-1774	40	0	0.0
P-1781	34	2	5.8
P-2019-1	40	1	2.5
P-2202-2	41	0	0.0
P-4353-1	36	0	0.0
T-3	34	2	5.8
G-24	42	0	0.0
G-130	42	0	0.0
G-543	35	0	0.0
Pant G-115	37	0	0.0
ICC-5	43	3	6.9

D. Ascochyta blight resistant lines

1. Advanced lines

The results of screening of 37 lines that were found promising to Ascochyta blight in the propagator screening and that were screened in the last year's stunt screening nursery are presented in Table 18. Nine lines, ICC-539, 838, 1005, 1012, 1272, 1583, 1911, 4939, and 4989 did not show any infection. Fourteen lines, ICC-666, 667, 693, 954, 1003, 1006, 1024, 1202, 1283, 1407, 1504, 2294, 3330, and 4935 showed less than 10% infection. Performance of these lines needs to be confirmed for one more year.

Table 18. Results of advanced screening of chickpea Ascochyta blight resistant lines to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

	Total	Infected	Percent		Total	Infected	Percent
Lines	plants	plants	infection	Lines	plants	plants	infection
ICC-118	19	6	10.6	1∝-1149	16	5	31.2
-272	18	5	27.7	-1202	24	1	4.1
-539	10	0	0.0	-1219	37	4	10.8
-567	4	1	25.0	-1272	32	0	0.0
-666	17	1	5.8	-1283	26	2	7.6
-667	38	1	2.6	-1329	14	8	57.1
-693	41	1	2.4	-1407	41	2	4.8
-838	26	0	0.0	-1504	30	2	6.6
-843	23	3	13.0	-1583	8	٥	0.0
-903	27	4	14.8	-1911	31	0	0.0
-904	18	2	11.1	-2264	26	3	11.5
-954	20	1	5.0	-2294	34	3	8.8
-1003	23	1	4.3	-3330	63	1	1.5
-1005	11	0	0.0	-4935	30	2	6.6
-1006	10	1	10.0	-4939	23	0	0.0
-1012	28	0	0.0	-4989	29	0	0.0
-1017	14	2	14.2	-6067	33	6	18.1
-1024	37	2	5.4	-7513	19	5	26.3
-1078	37	4	10.8	, , ,		•	

2. Preliminary screening

Thirty-seven lines that were found promising to Ascochyta blight in the propagator screening were tested for their reaction to stunt. The results are presented in Table 19. Only one line, ICC-3587 did not show infection but the plant number was low and it needs further checking. All the other lines had more than 10% infection indicating that none of them is promising.

Table 19. Results of preliminary screening of Ascochyta blight-resistations (propagator screening) to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

	Total	Infected	Percent	
Lines	lants	plants	infection	
ICC-3377	32	5	15.6	
-3378	20	13	65.0	
-3432	27	18	66.6	
-3495	20	5	25.Q	
-3496	9	3	37.5	
-3497	16	8	50 .0	
-3509	5	3	60.0	
-3573	21	7	33.3	
-3577	19	4	21.0	
-3578	18	11	61.1	
-3580	21	7	33.3	
-3581	34	9	26.4	
-3582	6	3	50. 0	
-3585	10	2	20.0	
- 3586	21	9	42.8	
-3587	5	0	0.0	
-37 37	23	21	91.3	
-3738	28	9	32.1	
-3739	9	6	66 .6	
-3740	30	7	23.3	
-3744	7	6	85.7	
-4751	9	2	22.2	
-476 2	29	21	72.4	
-4765	20	8	40.0	
-49 07	22	7	31.8	
-5 25 2	18	4	22.2	
-633 0	29	10	34.4	
-6354	25	15	60.0	
-6843	14	7	50.0	
-6 85 6	19	8	42.1	
-7 56 0	29	17	58.6	
-7563	32	15	46.8	
-7589	32	7	21.8	
-7611	22	8	36.3	
-7633	20	5	25.0	
-7664	18	9	50.0	
-7674	17	10	58.8	

E. Wilt and root-rot resistant lines

1. Advanced lines

The results of screening of 5 of the wilt and root-rot resistant lines that showed less than 10% in the last year's screening are presented in Table 20. One line, ICC-7254 was found free from infection, but its plant number was very low and needs further checking. Another line, ICC-3426 showed less than 10% and the remaining 3 had higher infection.

Table 20. Results of advanced screening of chickpea wilt and root-rot resistant lines to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

Lines	Total plants	Infected plants	Percent infection
1œ-391	23	6	26.0
-1450	5	3	60.0
-2860	17	3	17.6
-3426	16	1	6.2
-7254	3	0	0.0

2. Preliminary screening

The results of screening of 46 wilt and root-rot resistant lines are presented in Table 21. Two lines; ICC-516 and 1891 did not develop any infection but in both the cases the plant number was very low and therefore their performance needs further checking. Seven lines; ICC-2089, 6880, 8982, 10104, ICCC-10, G-543, and GG-669 showed less than 10% infection.

F. ICCC materials

The results of screening of 9 ICCC materials are presented in Table 22. These lines were tested in the last season also and were found to show less than 10% infection. Six lines ICCC-2, 3, 5, 9, 11, and 12 had less than 10% infection in the second screening also.

G. GIET lines

The results of screening of 35 GIET lines are presented in Table 23. Nine lines; H-76-105, ICCC-19, JG-1261, ICCC-17, GNG-88, BG-402, BG-239, GG-588-2509, and GNG-84 did not show any infection. Another 14 lines had less than 10% infection.

Table 21. Results of preliminary screening of chickpea wilt and root rot resistant lines to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

	Total	Infected	Percent
Lines	plants	plants	infection
ICC-102	14	4	28.5
-229	7	2	28,5
-267	27	9	33.3
-338	10	3	30.0
-434	8	5	62.5
-516	5	0	0.0
-519	8	2	25.0
-554	30	7	23.3
-867	9	2	22.2
-1891	1	0	0.0
-1910	10	2	20.0
-2072	6	6	100 .0
-2089	29	2	6.8
-2104	20	9	45.0
-2461	31	6	19.3
-256 6	10	5	50. 0
-26 60	14	5	35.7
-2812	6	3	50.0
-2835	13	5	38.4
-2883	13	2	15.3
- 3099	27	8	29.6
-3103	14	3	21.4
-33 96	28	12	42.8
- 34 39	17	6	35.2
-25 39	25	10	40.0
- 36 84	27	10	37.0
-45 19	24	4	16.6
-5 8 64	24	13	54.1
- 59 01	21	4	19.0
-60 98	15	8	53.3
-68 80	27	2	7.4
-7111	22	5	22.7
-7248	23	8	34.7
-7254	27	10	37.0
-76 81	36	14	38.8
-89 33	32	18	56.2
-89 82	42	4	9.5
-9001	24	8	33.3
-10104	41	1	2.4
-10394	40	10	25.0
1cc-10	38	2	5.2
G-543	30	2	6.6
GG- 588	28	3 3	10.7
GG-663	19	3	15.7
GG-669	33	1	3.0
GG-688	37	12	32.4

Table 22. Results of advanced screening of ICCC materials to pea leafroll virus (stunt) at Hissar during 1979-80 season

Particular	Total plants	Infected plants	Percent infection
ICCC-2	3	o	0.0
-3	23	2	8.6
-4	28	12	42.8
- 5	34	1	2.9
-7	27	8	29.6
-9	21	1	4.7
-11	2	•0	0.0
-12	38	3	7.8
-13	34	4	11.7

Table 23. Results of preliminary screening of GIET lines to pea leafroll virus (stunt) at Hissar during 1979-80 season

	Total	Infected	Percent		Total	Infected	Percent
Lines	plants	plants	infection	Lines	plants	plants	infection
76 105	24	•		DC: 403	16	•	12.6
H-76-105	24	0	0.0	BG-403	16	2	12.5
1CCC-14	28	1	3.5	BG-234	14	1	7.1
ICCC-15	30	1	3. 3	H-772	16	1	6.2
HMS-6	22	2	9.0	BG-404	16	1	6.2
HMS-23	31	3	9.6	BG-240	14	1	7.1
JG-1258	22	1	4.5	BG-402	18	0	0.0
ICCC-19	26	0	0.0	BDN-9.3	11	3	27.2
BG-406	23	4	17.3	BG-239	18	0	0.0
BG-405	26	2	7.6	BG-236	14	2	14.2
BG-401	30	. 4	13.3	GG-588-2509	13	0	0.0
JG-1261	29	0	0.0	BG-237	10	4	40.0
ICCC-18	32	1	3.1	GNG-15	14	5	35.7
1CCC-16	31	5	16.1	H-77-61	10	2	20.0
BG-235	27	2	7.4	P-324	-	-	-
ICCC-17	17	0	0.0	JG-2260	15	1	6.6
H208	32	3	9.3	CNG-84	16	0	0.0
GNG-88	14	0	0.0	JG-1259	-	-	•
RSG-40	13	2	15.3				

Observations not recorded.

H. GCVT lines

The results of screening of 12 GCVT lines are presented in Table 24. Mone of them was free from infection. Two lines; ICCC-10 and ICCC-13 had 10 or less than 10% infection. Others had more infection.

Table 24. Results of preliminary screening of GCVT lines to pea leafroll virus (stunt) at Hissar during 1979-80 season

Lines	Total plants	Infected plants	Percent infection
ICCC-4	25	12	48.0
RAV-52	15	5	33.3
BDN-9.3	9	3	33.3
ICCC-6	32	8	25.0
Phule-2	33	17	51.5
ICCC-9	26	3	11.5
RAV-54	25	8	32.0
ICCC-10	37	1	2.7
BG-220	27	7	25.9
rccc-13	30	3	10.0
Phule G-4	23	18	78.2
Phule G-1	21	15	71.4

I. Ascochyta blight-promising lines from Gurdaspur

Nineteen lines that were found promising to Ascochyta blight at Gurdaspur were screened (Table 25). Four lines, C-235, G-588, G-609, and G-679 did not show any infection. Another four lines; BG-216, G-543, G-570, and GG-677 had less than 10% infection.

J. Multilocation testing

Twenty-two of the promising lines were sent to Chile, Sudan, The Netherlands (only 5), New Zealand and New Delhi, India for testing. The results from the Netherlands showed that all the 5 lines were susceptible. Results from other places are awaited.

K. F2 materials

In collaboration with breeders, seven F₂ populations involving resistant parents were screened. Resistant and agronomically good looking plants were selected and bulked. The F₃ bulks will be tested next season.

Table 25. Results of preliminary screening of chickpea lines found promising to Ascochyta blight at Gurdaspur to pea leaf-roll virus (stunt) at Hissar during 1979-80 season

Lines	Total plants	Infected plants	Percent infection
BG-216	28	1	3.5
C-235	32	0	0.0
G-543	12	1	8.3
G-549	17	6	35.2
G-570	32	2	6.2
GG-575	28	. 4	14.2
GG-578	31	8	25,8
GG-580	28	4	14.2
GG-588	26	0	0.0
GG-589	36	6	16.6
GG- 609	31	0	0.0
GG-612	19	2	10.5
GG-677	26	1	3.8
GG-679	30	0	0.0
GG-684	27	19	70.3
GG-685	25	12	48.0
GG-686	16	5	31.2
H-75-35	26	3	11.5
1cc-11	7	2	28.5

V. EFFECT OF PLANTING DATES

Experiment to find out the effect of planting date on the disease incidence was continued for the third year. The planting dates were 20-9-1979, 30-9-1979, 15-10-1979, 30-10-1979, 15-11-1979, and 30-11-1979. Experimental design was RBD with 3 replications. The cultivar used was WR-315 and the plot had five five-meter rows.

Observations in the first week of February indicated higher disease incidence in the earlier dates of planting compared to later dates. Final observations could not be relied because of general drying of the plants due to some soil problem that affected the stands.

VI. EFFECT OF PLANT POPULATIONS

In an experiment conducted in collaboration with Physiology and Entomology Subprograms, the effect of plant population on the incidence of disease in 3 cultivars varying in the degree of susceptibility was studied both under sprayed and unsprayed conditions. The number of plants per m² studied were 8, 33, and 67 and the cultivars were G-130, 850-3/27 and WR-15. The design was split plot with 4 replications. Final observations were recorded at the time of maturity.

A. Sprayed conditions

The results are presented in Table 26. The disease incidence was more in low plant population (8 plants/ m^2) in all the three cultivars when compared to higher populations (33 out of 67 plants/ m^2). The differences among the varieties were negligible.

B. Unsprayed conditions

The results are presented in Table 27. The results were similar as under sprayed conditions.

Effect of plant population on the incidence of pea leaf-roll virus (stunt) in chickpea under insecticide spray conditions (1979-80, Hissar) Table 26.

	Spacing	?	Total	Total plants	**	Ir	Infected plants	d pla	nts	A.	ercent	Percent infection	uo	
Cultivar	(plants/m ²)	22	2	8	2	R	2	83	2	R.1	23	2	*	Yean
6-130	œ	124	156	162	152	21	14	15	12	16.9	8.9	9.5	7.8	10.7
	33	631	554	843	542	13	7	4	72	2.0	1.2	♦.0	4.0	1.9
	49	986	882	926	887	113	24	0	15	1.3	2.7	0.0	1.6	1.4
850-3/27	œ	152	162	189	145	19	13	20	7	12.5	0.8	10.5	8.	9.9
	33	524	268	565	518	70	16	9	19	3.8	2.8	1.0	3.6	2.8
	19	009	604	612	504	32	16	80	13	5.3	5.6	1.3	2.5	5.9
WR-315	æ	147	151	197	164	17	0 0	23	17	11.5	ري. س	11.6	10.3	9.7
	33	641	603	558	504	11	9	15	16	1.7	0:	2.6	3.1	2.1
	67	988	762	801	572	5	56	15		0.5	3.4	1.8	1.9	1.9

Table 27. Effect of plant population on the incidence of pea leaf-roll virus (stunt) in chickpea under unsprayed conditions (1979-80, Hissar)

	Spacing		Total	plant	.5	L	nfect	ed pla	ants		Perce	nt info	ection	
Cultivar	(plants/m ²)	Rl	R2	R3	R4	Rl	R2	R3	R4	Rl	R2	R3	RA	Nean
G-130	8	271	278	289	284	12	28	8	26	4.4	10.0	2.7	9.1	6.6
	37	1081	1198	1070	858	29	31	20	9	2.6	2.5	1.8	1.5	2.0
	67	1514	1340	1460	1013	10	22	12	7	0.6	1.6	0.8	0.6	0.9
850-3/27	8	278	290	241	235	29	12	18	6	10.4	4.1	7.4	2.5	6.1
	33	998	1148	1095	889	55	60	22	10	5.5	5.2	2.0	1.1	3.4
	67	1711	860	1290	1069	14	32	30	6	0.8	3.7	2.3	0.5	1.8
WR-315	8	268	286	300	273	17	14	19	5	6.3	4.9	6.3	1.8	4.8
	33	1084	1118	1145	802	12	22	21	16	1.1	1.9	1.8	2.0	1.7
	67	1455	1190	1347	1279	49	14	22	9	3.3	1.1	1.6	0.7	1.7

PROJECTICP-PATH-4(78): STUDIES ON ASCOCHYTA BLIGHT

I. SUMMARY

- Work at ICRISAT Center was mainly confined to screening of the kabuli germplasm lines in Isolation Plant Propagators. At ICARDA Center, Aleppo, Syria, extensive field screening of germplasm and breeding materials was carried out.
- 2. A total of 743 additional kabuli germplasm lines were screened and 116 lines showing 5 or less rating were identified.
- Of the 195 lines that showed 5 or less rating in the last year's screening, 104 lines showed consistent reaction in a repeat screening.
- 4. None of the 7 desi germplasm lines that were found resistant at ICARDA Center, Syria, was found resistant to the IARI-isolate of the blight fungus indicating the possibility that the two isolates are different.
- 5. None of 19 lines that were found promising under natural conditions at Gurdaspur in Punjab state of India was found resistant to the IARI-isolate of the fungus indicating the possibility that the two is-lates are different.
- 6. None of the 47 GIET and GCVT entries tested was found promising.
- 7. Some F₁ and F₂ materials involving resistant lines were screened to help the breeders in understanding the inheritance of resistance.
- 8. Of the five fungicides tested Dihane M-45 and Difolatan completely inhibited spore germination in vitro tests.
- 9. No difference was found in the germination of spores of Ascochyta rabiei in exudates of susceptible and some moderately resistant lines. There was also not much difference in the exudate pH. The pH of the exudates was lower at one month age compared to early stages.
- 10. There was no difference in the germination of spores of A. rabiei in the extracts of susceptible and moderately resistant lines tested.
- 11. Germination of spores of A. rabiei at higher concentration of a mixture of organic acids (1.3%) consisting of malic acid, oxalic acid and citric acid was inhibited.
- 12. Yeast cells in leaf exudates seemed to inhibit spore germination by increasing the pH of the exudates.

- 13. Seed of 56 promising lines was multiplied for multilocation test through the Chickpea International Ascochyta Blight Nursery (CIABN) as well as for supplying to cooperators.
- 14. A large number of germplasm lines and breeding materials was screened at ICARDA and several promising materials were identified.
- 15. Ascochyta blight and pea leaf-roll virus were found to be the major disease problems of chickpea in Algeria, Morocco, and Tunisia.

II. INTRODUCTION

Work on Ascochyta blight at ICRISAT Center was mainly confined to screening of kabuli germplasm in Isolation Plant Propagators. Lines received from other locations were also tested. The role of leaf exudates in disease resistance was further studied. Extensive field screening of germplasm and breeding materials was carried out at ICARDA Center, Aleppo, Syria.

III. SCREENING TECHNIQUE

As in the previous years, screening was carried out using Isolation Plant Propagators. In the preliminary screening for each germplasm entry, 10 seeds were planted in a single pot and were inoculated with spore suspension (IARI-isolate, 40,000 spores/ml) when the seedlings were 10-15 days old. Observations on incubation period, percent plants infected and killed were recorded. The disease severity was scored on a 9-point scale two times; first 10 days after inoculation and second 10 days later (recovery rating). The lines that showed 5-rating and less were rescreened with a larger plant population for confirmation. With each lot of screening, a susceptible check (Pb-7) was included.

IV. SCREENING FOR RESISTANCE

A. Kabuli germplasm

Screening of the additional germplasm lines was continued to identify additional promising lines. A total of 743 lines was screened. The summary of results indicating the percent of lines in each category of the scale used are presented in Table 28. Only one line; ICC-8078 showed a rating of 1. Sixteen lines; ICC-8030, 8084, 8085, 8086, 8131, 8463, 8820, 8821, 9147, 9149, 9150, 9258, 9278, 9284, 9367, and 9402 showed a rating of 3.

Additional 16 lines; ICC-9198, 9202, 9203, 924-, 9251, 9252, 9253, 9259, 9260, 9275, 9277, 9285, 9288, 9369, 9374, and 9403 showed a rating of 4. A total of 83 lines showed a 5-rating. These were ICC-7904, 7907, 7909, 7926, 7928, 7932, 7966, 79993, 8007, 8011, 8029, 8042, 8043,

8050, 8062, 8067, 8071, 8072, 8076, 8087, 8088, 8089, 8090, 8091, 8092, 8093, 8094, 8095, 8096, 8097, 8098, 8099, 8127, 8132, 8134, 8137, 8148, 8175, 8203, 8205, 8206, 8207, 8210, 8223, 8286, 8507, 8678, 8818, 8819, 8822, 8823, 8913, 8914, 8915, 8920, 8925, 8926, 8927, 8929, 8934, 8935, 8942, 8945, 8949, 8952, 8957, 8960, 9142, 9143, 9146, 9211, 9245, 9254, 9255, 9256, 9264, 9265, 9283, 9319, 9326, 9368, 9379, and 9394. Others showed 6 or higher rating. Lines with 5 or less rating will be rechecked.

B. Repeat screening of germplasm

A total of 195 germplasm lines that showed 5 or less rating in the last year's propagator screening were tested again. The summarised results are presented in Table 29. A total of 104 lines showed 5 or less rating. One line, ICC-3582, showed 1-rating. Two lines, ICC-3698 and 3724 showed 2-rating. Sixteen lines; ICC-3141, 3296, 3346, 3377, 3576, 3578, 3585, 3586, 3597, 3599, 3916, 3917, 6840, 6843, 6847, and 7676 showed 3-rating.

A total of 43 and 42 lines showed 4 and 5 ratings, respectively. The lines that showed 4-rating were ICC-3252, 3269, 3270, 3304, 3334, 3376, 3378, 3386, 3387, 3573, 3577, 3581, 3589, 3592, 3594, 3598, 3606, 3634, 3737, 3915, 3918, 3919, 4826, 4827, 4855, 4856, 4857, 4861, 4864, 5124, 6045, 6314, 6354, 6887, 6888, 7559, 7562, 7567, 7589, 7592, 7609, 7655, and 7773.

The lines that showed 5-rating were; ICC-3102, 3254, 3259, 3268, 3394, 3509, 3580, 3593, 3725, 3738, 3854, 3855, 3927, 4784, 4785, 4787, 4882, 4899, 5046, 5106, 5119, 5122, 5123, 6235, 6312, 6345, 6837, 6838, 7198, 7198, 7212, 7243, 7246, 7249, 7251, 7287, 7359, 7353, 7633, 7638, 7653, 7668, and 7718.

Table 28. Summarised results of screening of chickpea kabuli germplasm lines for resistance to Ascochyta blight in Isolation Plant Propagators at ICRISAT during 1979-80

Rating on 9-point scale	No. of entries	Percent entries
,	1	0.1
1	-	
2	0	0.0
3	16	2.1
4	16	2.1
5	83	11.1
6	16	2.1
7	46 8	62.9
8	38	5.1
9	104	13.9
_		

743

Total

Table 29. Summarised results of the repeat screening of chickpea germplasm lines for resistance to Ascochyta blight in Isolation Plant Propagators at ICRISAT during 1979-80

Rating on 9-point scale	No. of entries	Percent entries
1	1	0.5
2	2	1.0
3	16	8.2
4	43	22.0
5	42	21.5
6	43	22.0
7	34	17.4
8	9	4.6
9	5	2.5
	Total 195	

C. Materials from other locations

Materials received from ICARDA and the Indian National Programs were tested for their resistance to Ascochyta blight.

1. ICARDA promising lines

Seven Desi germplasm lines that were found resistant at ICARDA during 1978-79 screening were tested against the IARI-isolate. The results are presented in Table 30. Except ICC-5889 which showed a rating of 6, all others showed 8 or 9 rating. These results suggest that there may be different races of A. rabiei.

2. Gurdaspur promising lines

Nineteen lines sent by Dr. Gurdip Singh of Punjab Agricultural University, Ludhiana, India, were screened. These were found promising against the blight at Gurdaspur in Punjab under natural conditions during 1978-79 season. The results are presented in Table 31. All the lines showed a rating of 7. The difference in the reaction to the two isolates may be again due to difference in the race situation.

3. GIET entries

A total of 35 entries included in the Gram Initial Evaluation Trial (GIET) were screened. The results are presented in Table 32. None of them was found promising. All showed 6 or higher rating.

Table 30. Results of propagator screening of lines found promising at Syria

					Rating on	9-point scale
S. No .	ICC No.	Total plants	Percent infection	Percent killed	10th day after inoculaion	20th day after inoculation (Recovery rating)
1.	1256	48	100.0	0.0	8	8
2.	2459	43	95.3	16.2	6	8
3.	4111	59	0 .0	0.0	9	9
4.	4112	51	0.0	0.0	• 7	8
5 .	4113	56	0.0	0.0	7	8
6.	5889	33	96.9	21.2	5	6
7.	8227	47	0.0	0.0	7	8

Table 31. Results of screening of chickpea lines found resistant to blight at Gurdaspur (India) to IARI-isolate in Isolation Plant Propagator at ICRISAT, Hyderabad (1979-80)

			, , , , , , , , , , , , , , , , , , , ,		Rating on 9-	point scale
S.No.	Particular	Total plants	Percent infection	Percent killed	15th day after inoculation	20th day after inoculation
5.NO.	Partitular	pranca	III I GC C TOII	KILIUU	Inocuracion	MOCULACION
1.	BG-216	22	95.4	0.0	4	7
2	G-235	21	100.0	0.0	5	7
3.	G-543	18	100.0	0.0	4	7
4.	G-549	25	100.0	0.0	5	7
5	GG-570	19	100.0	0.0	4	7
6.	GG-575	21	100.0	0.0	5	7
7 .	GG-578 ·	22	100.0	0.0	5	7
8.	GG-580	23	95.6	0.0	4	7
9.	GG-588	22	100.0	0.0	5	7
10.	GG-589	22	100.0	0.0	5	7
11.	GG-609	20	100.0	0.0	5	7
12.	GG-612	22	100.0	4.5	5	7
13.	GG-677	20	100.0	0.0	5	7
14.	GG-679	19	100.0	0.0	5	7
15.	GG-684	24	100.0	0.0	5	7
16.	GG-685	19	100.0	0.0	5	7
17.	GG-686	20	80.0	0.0	4	7
18.	H-75-35	20	90.0	0.0	4	7
19.	ICCC-11	18	100.0	0.0	5	7

Table 32. Results of screening of GIET entries to Ascochyta blight in Isolation Plant Propagator at ICRISAT during 1979-80

					Rating on	9-point scale
					10th day	20th day after
			Percent		after	inoculation
Sl.		Total	infec-	Percent	inoculation	(Recovery
	Particular	plants	tion	killed		rating)
1.	BG-240	16	100.0	93.7	9	9
2.	BG-404	34	100.0	97.0	8	9 •
3.	H-772	28	100.0	85.7	8	8
4.	ICCC-17	34	100.0	67.6	9	7
5.	BG-235	27	100.0	92.5	8	8
6.	ICCC-16	31	100.0	93.5	9	9
7.	ICCC-18	15	100.0	93.3	9	9
	P-324	25	100.0	96.0	9	9
	BG-236	29	100.0	86.2	8	9
	H-77-61	40	100.0	95.0	8	9
11.	GNG-15	24	100.0	87.5	9	8
12.	BG-237	33	100.0	0.0	8	6
13.	GG-588-2509	28	100.0	32.1	7	7
14.	1CCC-14	15	100.0	100.0	9	9
15.	H-208	29	100.0	100.0	9	9
	BG-406	33	100.0	90.9	9	9
17.	ICCC-19	5	100.0	60.0	9	9
18.	BG-402	33	100.0	93.9	9	9
19.	JG-2260	25	100.0	84.0	9	8 ,
20.	GNG-84	31	100.0	96.7	9	9
21.	JG-1259	25	100.0	96.0	9	9
22.	RSG-40	29	100.0	96.5	8	9
23.	BG-239	27	100.0	62.9	9	8
24.	GNG-88	31	100.0	74.1	9	8
25.	ICCC-15	30	100.0	73.3	9	8
26.	HMS-6	19	100.0	78.9	6	8
27.	HMS-23	32	100.0	34.3	7	8
28.	JG-1258	27	100.0	70.3	7	7
29.	H-76-105	23	100.0	56.5	7	7
30.	BDN-9-3	35	100.0	85.7	9	9
31.	JG-1261	31	100.0	80.6	9	8
32.	BG-401	25	100.0	12.0	7	6
33.	BG-234	26	100.0	7.6	7	7
34.	BG-403	29	100.0	48.2	7	7
35.	BG-405	31	100.0	45.1	7	6
36.	Pb-7 (check		100.0	62.8	9	8
	ED-1 (CHACK	, ,,	100.0	02.0	J	6

4. GCVT entries

Twelve entries involved in the Gram Coordinated Varietal Trial (GCVT) were screened. The results are presented in Table 33. None of them was found promising. All showed 7 or more rating.

Table 33. Results of screening of GCVT entries to Ascochyta blight in Isolation Plant Propagator at ICRISAT during 1979-80

				Rating on	9-point scale
Particular	Total plants	Percent infec- tion	Percent killed	10th day after inoculation	20th day after inoculation ^a
1000-6	12	100.0	91.6	8	8
BDN-9-3	34	100.0	98.2	9	7
RAV-52	42	100.0	100.0	9	9
ICCC-4	36	100.0	100.0	8	9
Phule G-1	36	100.0	52.7	7	7
Phule G-4	40	100.0	75.0	8	8
ICCC-13	35	100.0	82.8	9	8
BG-220	29	100.0	100.0	9	9
1cc-10	32	100.0	100.0	9	9
RAV-54	22	100.0	100.0	9	9
1CC-9	25	100.0	92.0	9	9
Phule-2	32	100.0	96.8	9	9

D. Inheritance study

The study was carried out in collaboration with chickpea breeding subprogram (Dr. Jagdish Kumar). In all 4 F_1 s and F_2 s involving one moderately resistant (ICC-1903), one tolerant (Pant G-114) and two susceptible parents (BG-203 and P-690) were tested

1. Parents and F1 material

The results of screening of the 4 parents and F_1 s are presented in Table 34. The F_1 s between resistant and susceptible parents remained resistant. The F_1 between the tolerant and susceptible parents remained tolerant. The F_1 between susceptible parents showed susceptibility. The results were passed on to Dr. Jagdish Kumar for further interpretations.

2. F₂ materials

The results of screening of the F_2 populations are presented in Table 35. The experiment was repeated using the crosses between the moderately resistant parent and similar results were obtained. The results were handed over to Dr. Jagdish Kumar.

Table 34. Results of screening of some chickpea parental lines and F₁s to IARI-isolate of A. rabisi in Isolation Plant Propagator at ICRISAT (1979-80)

Particular	Total plants	Percent infection	Percent killed	Rating on 9-point scale
Parents	•			
Pant G-114	9	100.0	0.0	5 - Tolerant
I∝-1903	10	100.0	0.0	3-5 - Moderately * resistant
BG-203	9	100.0	0.0	7 - Moderately susceptible
P- 690	9	100.0	0.0	7 - *
F ₁ s				
ICC-1903 x BG-203	10	90.0	0.0	3
P-690 x Pant G-114	10	90.0	0.0	5
ICC-1903 x Pant G-114	10	70.0	0.0	3
P-690 x BG-203	9	88.8	0.0	7

Table 35. Results of screening of some F₂ chickpea materials to IARI-isolate of Ascochyta rabiei in Isolation Plant Propagators at ICRISAT (1979-80)

		Total	No.	of plan	nts with	a rati	ng of
S.No.	Cross	plants	1	3	5	7	9
1.	ICC-1903 x BG-203	181	26	79	25	46	5
2.	P-690 x Pant G-114	92	2	41	25	24	0
3.	ICC-1903 x Pant G-114	208	15	75	49	65	4
4.	P-690 x BG-203	84	0	0	10	61	3

3. Collection of seed from resistant plants

Seedlings showing 5 or less rating from the crosses involving moderately resistant and tolerant parents were transplanted into bigger pots for collection of seed. Seeds from plants with 1, 3, and 5 ratings in each cross were collected separately, bulked and handed over to breeders for further advancement.

E. Collection of new isolates

Isolations were made from blight infected chickpea samples collected from Hissar, Haryana state of India and pure cultures of Asocchyta rabisi were obtained. The cultures were found to be pathogenic and were preserved for future use.

V. DISEASE CONTROL

As a follow up of last year's fungicide spray experiment in plant propagator, the effect of 5 fungicides on spore germination of A. rabic in vitro was studied.

A. Effect of fungicides on spore germination

Five fungicides; Bavistin, Benlate, Dithane M-45, Difolatan and Daconil were used at three concentrations; 0.1, 0.2 and 0.3%. To one drop of spore suspension (16-day-old culture) one drop of fungicide suspension was added in cavity slides which were placed in petri plates with moist filter paper. The plates were incubated at 24°C and observations were recorded after 24 hrs. The results are presented in Table 36 (repeated 4 times).

When compared to control all the five fungicides tested significantly inhibited spore germination at all the concentrations. Inhibition increased with the increase in the concentration. Two fungicides Dithane M-45 and Difolatan completely inhibited spore germination at all the concentrations tested. Bavistin was least effective. Benlate and Daconil gave complete inhibition at 0.2 and 0.3% concentrations.

VI. MECHANISM OF RESISTANCE

Studies on the role of leaf exudates and extracts in resistance in chickpeas was further explored. The effect of exudates and extracts from susceptible and some moderately resistant lines on spore germination was studied. The pH of the exudates of different lines was also studied.

A. Leaf exudates

Exudates were prepared by shaking one gram of leaf tissue collected early morning in 1 ml of sterile distilled water (standard). The suspension was given a low speed of 3000 rpm for 5 minutes to remove the dust particles. Effect of the exudates on spore germination was studied using cavity slides placed in petri plates with moist filter paper (24°C). Observations on spore germination were recorded after 24 hrs.

Table 36. Effect of different fungicides on conidial germination of Ascochyta rabiei, in vitro

						Š	190	conidi	-					Average
	Concen- No. of	No. of	conidia	lia obi	observed	796	in	ą	İ	Perc	ent ge	Percent germination	8	per cent-
Pungicide	tration	2		22	RA	Z	2	22	Z	2	2	2	Z	•86
	c	100	8	90	•	Ħ	9	33	1	34.0	40.0	35.0	•	37.3
BAVESTI	, ,	8	8 6	901	1	91	70	17	ı	10.0	20.0	17.0	ŧ	15.6
	0.3	100	100	100	ı	77	11	91	•	12.0	17.0	16.0	ŧ	15.0
4	0.1	75	•	9	8	Ŋ	0	m	•	9.9	ſ	5.0	13.3	8.3
Dentara	2.0	100	100	100	100	0	0	0	0	0.0	0.0	0.0	0.0	0.0
	0.3	001	8	100	100	0	0	0	0	0.0	0.0	0.0	0.0	0.0
14. A. S.	0	100	100	100	100	0	0	0	0	0.0	0.0	0.0	0.0	0.0
Dictions and I	0.2	100	100	100	201	0	0	0	0	0.0	0.0	0.0	0.0	0.0
	0.3	100	100	100	100	0	0	0	0	0.0	0.0	0.0	0.0	0.0
nd Colleton	0.1	001	81	100	81	0	0	0	0	0.0	0.0	0.0	0.0	0.0
	0.2	100	801	100	100	0	0	0	0	0.0	0.0	0.0	0.0	0.0
	0.3	100	100	100	100	0	0	0	0	0.0	0.0	0.0	0.0	0.0
ne och 11	0.1	001	8	40	100	0	0	-	0	0.0	0.0	2.5	0.0	9.0
***************************************	0.5	100	100	100	100	0	0	0	0	0.0	0.0	0.0	0.0	0.0
	0.3	100	100	100	100	ö	0	0	0	0.0	0.0	0.0	0.0	0.0
Control	ı	100	700	100	ı	06	8	87	•	90.0	90.0	87.0	•	89.0
						į								

al6-day old culture.

boservations recorded after 24 hours.

1. Effect of exudates from susceptible and moderately resistant lines on spore germination

The effect of exudates from three moderately resistant lines on spore germination of A. rabie: compared to exudates from susceptible Pb-7 and distilled water was studied. The results are presented in Table 37. No appreciable differences in spore germination were observed.

2. Effect of different dilutions of exudates of susceptible and moderately resistant line on spore germination

The effect of different dilutions of the exudates of susceptible cultivar Pb-7 and ICC-1256, a moderately resistant line on spore germination was studied. The results are presented in Table 38. No difference in the germination of spores was however found

3. pH of the exudates at different times of day

The pH of the exudates of susceptible and moderately resistant lines at different times of the day was measured to see if there is any difference. The results are presented in Table 39. Not much difference among the cultivars and between different times of the day was found.

4. pH at different ages

The pH of the exudates at different ages starting from 6 to 32 days was measured. The results are presented in Table 40. The pH seemed to decrease with age.

B Leaf extracts

The effect of leaf extracts from susceptible and some moderately resistant lines on the germination of spores was studied. To one drop of extract prepared by squezing the leaf tissue, one drop of spore suspension was added. Incubation and observations recorded were same as described earlier. The results are presented in Table 41. No clear trend was obtained and the germination in the extracts was more than in distilled water.

Table 37. Effect of exudates of chickpea on conidial germination of Assochata rabie:

		No of	conid	ia obs	erved	No of conidia observed No. of conidia germinated	conidi	a germ	inated	Perc	Percent germination	rainat	ion	
cultivar		2	3	23	*	B	23	R3	R4	R	2	2	z	Average
pb-7 (S)		530	300	75C	675	505	759	675	550	95.2	95.2 53.0 90.0 81.4	90.0	81.4	79.9
ICC-1256 (MR)	(MR)	350	827	725	525	342	653	575	160	7.76	97.7 79.1 79.3 30.4	79.3	30.4	71.6
ICC-4112 (MR)	(MR)	953	415	550	470	927	385	430	210	97.2	97.2 92.7 78.1 44.6	78.1	4 .6	78.2
ICC-8227 (MR)	(MR)	200	515	975	009	7	492	650	125	1.0	1.0 91.6 66.6	9.99	20.8	45.0
Control		459	100	575	•	405	06	200	•	88.2	88.2 90.0 86.9	96.9		98.3

a30 days old seedlings; I g leaf tissue/1 ml of water.

S - Susceptible; MR - Moderately resistant.

bls days old culture

S

Table 38. Effect of different dilutions of chickpea leaf exudates on conidial germination of Ascochyta rabiei

		No.	of con	idia	No.	of con	idia				
		obs	erved		ger	minated		Percen	t germi	nation	
Cultivar	Dilution	Rl	R2	R3	Rl	R2	R3	Rl	R2	R3	yverage
Pb-7 (S)	1:2	500	675	280	375	550	104	75.0	81.4	37.1	64.5
	1:4	225	650	36 0	175	57 5	110	77.7	88.4	30.5	65.5
	1:8	450	800	100	350	600	50	77.7	75.0	50.0	67.5
	1:16	400	625	100	250	525	30	62.5	84.0	30.0	58.8
ICC-1256 (MR)	1:2	425	575	315	370	400	201	87.0	69.5	63.8	73.4
	1:4	225	725	190	140	525	134	62.2	72.4	70.5	68.3
	1:8	350	400	100	225	375	40	64.2	- 93.7	40.0	66.0
	1:16	225	400	101	90	300	52	40.0	75.0	51.4	55.4

S - Susceptible; MR - Moderately resistant.

Table 39. pH of leaf exudates of different cultivars of chickpea estimated at different times of the day a

pH of the leaf exudates						
	9	hr	1	2 hr	10	5 hr
Cultivar	Rl	R2	Rl	R2	Rl	R2
Pb-7 (S)	3.5	4.8	3.7	3.9	4.1	3.7
ICC-1256 (MR)	3.7	3.5	3.7	3.5	4.3	3.4
ICC-4112 (MR)	3.3	3.9	3.5	3.8	3.5	3.7
ICC-8227 (MR)	3.8	3.8	3.8	3.8	3.8	3+4

⁹²⁻days-old seedings.

Table 40. pH of chickpea leaf exudates of different cultivars at different ages

	pH of the leaf exudates at the age of a				
Cultivar	6 days	9 days	14 days	32 days	
Pb-7 (S)	4.7	5.0	5.0	3.5	
ICC-1256 (MR)	4.6	5.2	5.0	3.7	
ICC-4112 (MR)	•	5.2	5.3	3.3	
ICC-8227 (MR)	-	5.7	5.6	3.8	

^{&#}x27;l g leaf tissue washed in 3 ml water.

Table 41. Effect of leaf extracts of different cultivars of chickpea on conidial germination

	No. of conidia observed		No. of conidia germinated		Percent germination		Aver
Cultivar	Rl	R2	Rl	R2	Rl	R2	age
Pb-7 (S)	100	100	50	50	50.0	50.0	50 .0
ICC-1256 (MR)	100	100	64	68	64.0	68.0	66 .0
ICC-8227 (MR)	100	100	38	34	38.0	34.0	36.0
Control (DW)	100	100	28	32	28.0	32.0	30.0

[&]quot;32-days-old seedlings.

bl g leaf material washed in 1 ml water.

^{5 -} Susceptible; MR - Moderately resistant.

S - Susceptible; MR - Moderately resistant; DW - Distilled water.

C. Acid mixture

The effect of different concentrations of organic acid mixture prepared by mixing malic acid, oxalic acid, and citric acid in same concentration and proportion as found in the leaf exudates on spore germination was studied (Table 42). No germination was obtained in higher concentrations and at lower concentrations slight stimulation was obtained.

Table 42. Effect of different concentrations of acid mixture on conidial germination of Assochyta rabieiab

	No. of observe	_	No. of germin	conidia ated ^c		nt nation	Aver-
Treatment	Rl R2		R1 R2		R1 R2		490
Concentrated acid mixture + spore suspension	1000	1000	0	o	0.0	0.0	0.0
10 fold dilution + spore suspension	1000	1000	0	0	0.0	0.0	0.0
100 fold dilution + spore suspension	1000	3 70	25	180	2.5	48.6	25.5
1000 fold dilution + spore suspension	1000	5 95	360	470	36 . 0	78.9	57.4
Control (only spore suspension)	10000	430	498	160	49 8	37.2	43.5

Acid mixture 1.3% (94.2% malic acid + 5.6% oxalic acid + 0.2% citric acid).

D. Role of yeast cells

During the course of studies on the effect of exudates on spore germination, contamination with yeast cells was found. Whenever contamination with yeast cells occurred, germination of spores was inhibited. It was suspected that yeast cell play some role in spore germination through their action on the acid exudates. When yeast cells (30 hrs culture multiplied on media: yeast extract 3 g + malt extract 3 g + glucose 10 g + peptone 5 g + water 1000 ml) were added to the organic acid mixture 1.3% (malic acid 94.2% + oxalic acid 5.6% + citric acid 0.2%) the pH which was 3.2 at 0 hr was increased to 6.53 after 6 hrs. It is suspected that the yeast cells inhibit spore germination by increasing pH which may be deleterious to spore germination. This was further supported by the observation where spore germination in the

b8-days-old culture.

^C24 hrs after incubation.

leaf exudate of ICC-1256 where yeast cells were added was only 4.6% against 50% germination in the control (exudate without yeast cells). This aspect needs further study.

VII. SEED MULTIPLICATION

Seeds of 56 lines that were found promising in the repeated screenings last year was multiplied for multilocation testing through Chickpea International Ascochyta Blight Nursery (CIABN) operated from ICARDA, Aleppo, Syria and for supply to cooperators.

VIII. SUMMARY OF WORK DONE AT ICARDA

A period of 3 months was spent by Dr. M.V. Reddy at ICARDA, Aleppo, Syria to help Dr. K.B. Singh, ICRISAT's Chickpea Breeder, in Ascochyta blight screening. In the later part of the stay, a brief trip to north Africa was undertaken to assess disease problems in that region and see the performance of the entries in CIABN-80. A detailed report on the work carried at ICARDA and visit to north Africa are being prepared separately and salient features of the work carried out at ICARDA are given. The work at ICARDA was conducted in close consultation and collaboration with Drs. K.B. Singh and G.C. Hawtin.

- 1. The causal agent of the fungus causing blight at ICARDA, Syria, was confirmed to be Ascochyta rabiei (Pass.) Lab.
- 2. Very high disease pressure was created artificially in an area of 5.0 ha where large amount of germplasm, breeding materials and others was screened.
- 3. Further standardization of the field screening procedure in terms of method of inoculation, date of inoculation, frequency of spreader rows, and row direction was carried out.
- 4. A total of 4789 entries in 21 trials were screened and 3 highly resistant, 156 resistant, and 335 tolerant lines/materials were identified.
- 5. The entries of CIABN-80 were screened in the plastic house by artificial inoculation and the results showed good correlation with field results.
- 6. Seed infection in a susceptible cultivar was found to be 10% and Calixin M and Benlate mixture gave very good eradication.
- 7. The leaf exudates and extracts were found to play no role in resistance in chickpea.
- 8. No difference in the stomatal count of susceptible and resistant lines studied was found.

IX. SURMARY OF THE REPORT ON TRIPS TO ALGERIA, MOROCCO, AND TUNISIA BY DR. M.V. REDDY

The trips were made in the first half of May 1980. Summaries are given below:

A. Algeria

- Present yields are low because of low plant densities, Ascochyta blight and weeds.
- Tall, blight resistant cultivars (bold seeded kabuli) for closer spacings suitable for mechanical harvesting are urgently needed.
- 3. Blight and pea leaf roll virus resistant lines identified at ICARDA/ICRISAT Centers will be very useful.
- 4. Winter planting has great potential for increasing the yields. The problems associated with winter plantings are: (i) coincidence of cereal plantings with the legume plantings, (ii) delay of rains, (iii) cold and frost damage, and (iv) weeds.
- 5. The IDRC project on grain legumes has helped in conducting agronomic trials and initiating breeding program.

B. Morocco

- 1. Morocco is one of the countries with a great potential for increased chickpea production.
- 2. The most important need is development of kabuli cultivars with Ascochyta blight resistance suitable for mechanical harvesting.
- 3. Winter chickpeas will go a long way in increasing production and bringing changes in the cropping pattern.
- 4. The blight and pea leaf-roll virus resistant lines identified at ICARDA/ICRISAT Centers will be of immediate use.

C. Tunisia

- 1. Chickpea is a major food legume crop grown and is given good attention.
- 2. Introduction of winter chickpeas in southern parts of Tunisia where rainfall is less than 400 mm has good potential. Also winter chickpeas in light soils in northern parts may be more productive.

- 3. Blight appears to be the major disease problem and resistant lines identified at ICARDA should be very usefu.
- 4. Tall, bold seeded kabuli types with blight resistance for mechanical harvesting are desirable.
- 5. At present mostly trials sent by ICARDA are being tested and there is no breeding program.
- 6. Desi chickpeas are not yet all cultivated and sending desi material either in yield trials or disease nurseries may not be of much use.

PROJECT: CP-PATH-5(78): INTERNATIONAL CHICKPEA DISEASE NURSERIES

I. SUMMARY

- 1. The International Chickpea Root Rots/Wilt Nursery (ICRRWN) was sent to 33 cooperators in 19 countries. The nursery was sent for planting at 35 locations and had 56 entries. A separate report on the performance of these entries will be prepared.
- 2. Twenty entries from ICRISAT were sent to Dr. K.B. Singh, ICRISAT': Chickpea Breeder at ICARDA for inclusion in the Chickpea International Ascochyta Blight Nursery (CIABN). These entries were found promising in the 'propagator screening' at ICRISAT Center.
- Several nursery locations were visited in Algeria, India, Nepal, Syria, Tunisia, and Turkey.
- 4. A large number of scientists from several countries visited ICRISAT to see our work and exchange information.
- 5. The second International Chickpea Pathology Training Course was held in January-February 1980. Eight scientists from Egypt and India participated.

II. INTRODUCTION

The first Cooperative Chickpea Disease Nursery with 30 entries was operated during 1976-77. In January 1978 we formally initiated a project on nurseries with the following objectives:

- 1. Share promising material with cooperators in different countries,
- 2. Identify stable sources of resistance for use in breeding program at ICRISAT, and
- 3. Get a feed-back on susceptibilities of the entries to other locally serious diseases.

Since 1977-78 two separate nurseries were organised. These are (i) International Chickpea Root Rots/Wilt Nursery and (ii) International Chickpea Ascochyta Blight Nursery (ICABN). The reports for 1977-78 and 1978-79 nurseries were compiled and circulated (ICRISAT Pulse Pathology Progress Reports 4 & 7). We have now integrated our ICABN with the one operated from ICARDA and from here on it will be a joint ICARDA-ICRISAT nursery. It will be called Chickpea International Ascochyta Blight Nursery (CIABN). Results of the ICREWN 1979-80 have come from most of the locations. The results are being compiled and a separate report will be prepared.

In order to further strengthen our international activities, our staff undertook tours to different cooperating locations. We also invited well-known scientists as consultants.

III. ICRRON 1979-80

A. List of countries and cooperators

No.	Country	Cooperator(s)
1.	Afghanistan	Dr. M. Ghuffran President of Agricultural Research Research Department Ministry of Agriculture and Land Reforms Kabul
2.	Algeria	Mr. M.N. Bakhtri Regional Dryland Agronomist C/o UNDP BP: 823 Algiers
3.	Chile	Dr. Gabriel Bascur B. Programa de Leguminosas de Grano Estacion Experimental la Platina Instituto de Investigaciones Agropecuarias, Casilla 114-D Santiago
4.	Egypt	Dr. Ali Abdel Aziz Head, Grain Legume Section Field Crops Institute Agricultural Research Center Giza
5.	Ethiopia	Mr. Alemu Mengistu Plant Pathologist Agricultural Experiment Station Addis Ababa University P.O. Box 32 Debre Zeit
6.	Iraq	Mr. Issam Najjar Food Legumes Programme Directorate General of Field Crops Abu-Gharib Baghdad

No.	Country	Cooperator(s)
7,	Jordan	Dr. Hassan Gharaybeh Director Department of Agricultural Research and Extension P.O. Box 226 Amman
8.	Kenya	The Director ARD-KARI P.O. Box 30148 Nairobi
9.	Mexico	Ing. Santiago Sanchez INIA Auxiliar de Leguminosas Comeatibles Apartado Posta No.6-882 Y 6-883 Mexico 6 D.F.
10.	Nepal	Mr. R.P. Sah Assistant Agronomist (Pulses) Parwanipur Agriculture Station Birganj, Parwanipur Narayani Zone
11.	Pakistan	Dr. S.J. Hamid Assistant Plant Pathologist B-6, Al-Markaz, F-7/2 Cereal Diseases Research Institute Pakistan Agricultural Research Council Islamabad
12	Peru	The Director Centro Regional Investigacion Agraria de Norte APTDO 116 Chiclayo
13.	Argentina	Ing. Agr. Susana Garica Medina Mejoramiento de Legumbres Institute Nacional de Tecnologia Agropecuaria Estacion Experimental Regional Agropecuaria Salta Cerrillos (Salta)
14,	Sudan	Dr. Farouk Ahmed Salih Agricultural Research Corporation Hudieba Research Station P.O. Box 31

Ed-Damer

No.	Country	Cooperator(s)
15.	Syria	Dr. K.B. Singh ICRISAT Chickpea Breeder The International CEnter for Agricultural Research in the Dry Areas (ICARDA) P.O. Box 5466 Aleppo
16.	Tunisia	Dr. MLAIKI Ahmed Head, INRAT Pathology Laboratory Republique Tunisienne Ministere de L'Agriculture, INRAT Ariana, <u>Tunis</u>
17.	U.S.A.	Dr. John C. Philips Assistant Professor Crop Science Department California Polytechnic State University San Luis Obispo California 93407
18.	Yemen Arab Republic	Mr. K.M. Ahmed Pest Control Assistant (FAO) Central Agricultural Research and Training Center UNDP/FAO Project Post Box 4788, Taiz Yemen Arab Republic C/o FAO, Rome, Italy
19	India	Dr. K.K. Zote Badnapur Mr. M.D. Gupta Gwalior Dr. K. Sengupta Berhampore Dr. R.N.S. Tyagi Durgapura Dr. Gurdip Singh Ludhiana Dr. Prabhakar Shukla Kanpur
		Dr. S.R. Kotasthane

Jabalpur

No.	Country	Cooperator(s)
19.	India	Dr. J.S. Grewal New Delhi
		Dr. R.N. Singh Faizabad
		Dr. U.P. Singh Varanasi
		Dr. M. Mahmood Dholi
		Dr. B.L. Jalali Hissar
		Dr. K.C. Agarwal Raipur
		Dr. R.V. Hiremath Gulbarga
		Dr. B.G. Desai Dantiwada

Dr. Y.L. Nene/Dr. M.P. Haware

ICRISAT

B. Entries Following were the entries

S.No.	ICC No.	Pedigree	Origin
1.	102	P-79	India
2.	229	P-180-1	44
3.	267	P-212-1	94
4.	338	P-253	34
5.	434	P-319	pt .
6.	516	P-392	**
7.	519	P-394	19
8.	554	P-436-2	#1
9.	867	P-6 9 0	Ħ
10.	1891	P-1514	\$1
11.	1910	P-1542	Unknown
12.	1913	P-1546	**
13.	2072	P-1670	India
14.	2083	P-1679-2	Mexico
15.	2086	P-1683	ti
16.	2089	P-1684	t #

S.No.	ICC No.	Pedigree	Origin
17.	2104	P-1696-1	Mexico
18.	2461	P-2249	Iran
19.	2566	P-2559	Iran
20.	2660	P-2686-2	•
21.	2812	P-3036	**
22.	2835	P-3107-1	Unknown
23.	2883	P-3251	Iran
24.	3099	P-3614	#4
25.	3103	P-3617	Unknown
26.	3 3 9 6	P-4083	Iran
27.	3439	P-4116-1	**
28.	3539	P-4237	India
29.	3684	P-4321-2	Iran
30.	4519	P-6067	India
31.	5864	T-3 (Owalior)	•
32.	5901	T-32	••
33.	6081	JG-57	91
34.	6098	JG-74	*1
35.	6366	NEC-312	Iran
36.	6385	NEC-348	ti.
37.	6455	NEC-460	\$ (
38.	6494	NEC-529	m
39.	6880	NEC-1089	91
40.	6926	NEC-1166	M
41.	7111	NEC-1470	**
42.	7248	NEC-1621	India
43.	7254	NEC-1627	Pakistan
44.	7681	P-1179	India
45.	8933	WR-315	*1
46.	8982	NEC-346	Iran
47.	9001	NEC-426	Iran
48.	9117	NEC-847	м
49.	10104	P-6131	India
50.	10394	Coll.No. 129	**
51.	•	IC∞-10	ICRISAT
52.	•	G-543	India
53.	-	GG-588	*
54.	-	GG-663	**
55.	•	GG-669	99
56.	-	GG-688	**
57.	4951*	JG-62	India

^{*}Wilt susceptible check

IV. TOURS

Dr. M.V. Reddy spent 6 weeks in the laboratory of Dr. Myron K. Brakke, world-renowed plant virologist located at the University of Nebraska, USA.

Dr. Y.L. Nene participated in the IX International Plant Protection Congress in Washington, USA. He also visited the Commonwealth Mycological Institute, UK.

Dr. Y.L. Nene also visited Bangladesh at the invitation of that country's Government to study disease situation in pulse crops, particularly chickpeas. His tour report is separately available. The diseases observed in chickpeas were: Zinc deficiency, Collar rot (Solerotium rolfsii), root rot (Rhisoctonia solani), wilt (Fusarium oxysporum f.sp. ciceri), stunt (pea leaf-roll virus), root-knot (Meloidogyne spp.), and iron chlorosis. Of these zinc deficiency was widespread and the next in importance was collar rot. Other problems can be considered minor ones.

Dr. M.P. Haware spent 6 weeks at Commonwealth Mycological Institute, U.K., during September-October 1979 to participate in Refersher Course on the Taxonomy and Identification of Microfungi. He also visited Danish Government Institute of Seed Pathology for the Developing Countries, Copenhagen, on his return journey.

Dr. M.V. Reddy was deputed for a period of three months to work as a chickpea pathologist with Dr. K.B. Singh, ICRISAT's Chickpea Breeder at ICARDA. He worked specifically on the screening of chickpea for Ascochyta blight resistance. He also visited Algeria, Morocco, and Tunisia. He tour report is available separately. Chickpea diseases that he observed in these three countries were:

Algeria: Ascochyta blight, Stunt, Rust, Alfalfa mosaic virus.

Morocco: Ascochyta blight, Stunt, Rust, Stem rot, Alfalfa mosaic virus.

Tunisia: Ascochyta blight, Wilt, Stem blight, Stunt, Iron chlorosis.

Drs. M.P. Haware, M.V. Reddy, and Y.L. Nene undertook several tours in India, visiting locations where the nursery was planted.

V. CONSULTANT

Dr. L. Bos, Plant Virologist, Research Institute for Plant Protection, Wageningen, The Netherlands, was invited to serve as a consultant in pulse virology. He submitted a report and made several important recommendations for future work on pigeonpea and chickpea viruses.

VI. SECOND INTERNATIONAL CHICKPEA PATHOLOGY TRAINING COURSE

ICRISAT, with its world mandate for chickpea improvement, considered it necessary to share the knowledge on chickpea pathology with

the cooperating scientists through a course. The major objectives of this course are:

- To acquaint cooperating scientists with the procedures for

 (i) diagnosing different diseases of chickpea, (ii) identifying pathogens, and (iii) screening germplasm and breeding materials for disease resistance.
- 2. To further strengthen the links with cooperating scientists and through them with their respective countries.
- To emphasize the role of pathologists in crop improvement progra

The program of the course was drawn up to fulfil the above objectives.

As a response to the invitations sent by ICRISAT, eight scientists participated in the course. Their names and the countries have been listed below:

EGYPT

Mr. S.A.N. Omar

Giza

INDIA

Dr. R.N.S. Tyagi	Jaipur
Miss Om Gupta	Jabalpur
Mr. B.K. Sinha	Dholi
Mr. M.S. Sangwan	Hissar
Dr. Gurdip Singh	Ludhiana
Dr. R.N. Singh	Faisabad
Dr. Mahendra Pal	New Delhi

The course was initiated on January 10th. The program included following activities.

- 1. Field visits at ICRISAT Center and outside to get acquainted with characteristic symptoms of diseases and to learn about developing wilt/root rot sick plots.
- 2. Lectures on chickpea pathogens and general lectures on plant viruses.
- Laboratory sessions on pathogenic fungi and viruses of chickpea. Emphasis was on the identification of pathogens, special procedures for isolations and inoculations and laboratory/net house screening procedures.
- Lectures (followed by field visits) on germplasm, breeding for disease resistance, chickpea microbiology, and chickpea entomology.

- Field visits and discussions to get acquainted with other related activities of ICRISAT, such as cereals pathology, groundnut pathology, quarantine, etc.
- 6. Visit to the Central Plant Protection Training Institute.
- 7. Visits to farmers' fields in northern India (Delhi-Agra; Delhi-Hissar).
- 8. Visit to ICRISAT and Haryana Agricultural University chickpea fields at Hissar.
- 9. Presentation by participants summaries of the research activities at their respective research stations.
- 10. Distribution of relevant literature, books and sets of 20 colored transparencies, etc.

Staff members of the pulse pathology subprogram, who were closely associated with the course were Dr. M.P. Haware, Dr. M.V. Reddy, Dr. Y.L. Nene, Dr. J. Kannaiyan, Mrs. Sheila Viajayakumar, and Mr. Narayan Rao. 'Other ICRISAT scientists who gave lectures and/or were associated with laboratory/field activities were: Dr. P.J. Dart (Microbiology); Dr. L.J.G. van der Maesen (Genetic Resources Unit); Dr. Jagdish Kumar (Chickpea Breeding); Dr. W. Reed (Pulse Entomology); Dr. R.J. Williams (Millet Pathology); Dr. L.K. Mughogho and Dr. S.R.S. Dange (Sorghum Pathology); Dr. D. McDonald and Dr. D.V.R. Reddy (Groundnut Pathology); and Dr. K.K. Nirula (Plant Quarantine). All administrative arrangements were made by the Training Unit of ICRISAT.

A program evaluation sheet was given to each participant and very useful comments for making improvements in the course and for further cooperation among chickpea pathologists have been received. We will make sincere efforts to act on the suggestions that have been made.

One thing has disappointed us and i.e., less participation of cooperators from countries other than India, inspite of our efforts. We will in future intensify our efforts to ensure more participation from other cooperating countries.

APPENDIX-I

LIST OF PUBLICATIONS

Published

- 1. Haware, M.P., and Y.L. Nene. 1979. Non-seed borne nature of powdery mildew of chickpea. PANS 25(4): 464-465.
- 2. Haware, M.P., and Y.L. Nene. 1979. Symptomless carriers (hosts) of the chickpea wilt fungus. International Chickpea Newsletter No. 1: 8.
- 3. Haware, M.P., and Y.L. Nene. 1979. Physiologic races of the chickpea wilt pathogen. Chickpea Newsletter No. 1: 7-8.
- 4. Haware, M.P., and Y.L. Nene. 1980. Blight observed in the off-season chickpea nursery in Kashmir. Chickpea Newsletter No. 2: 12.
- 5. Nene, Y.L., and M.P. Haware. 1980. Screening of chickpea for resistance to wilt. Plant Disease 64: 379-380.
- 6. Reddy, M.V., and Y.L. Nene. 1979. A case for induced mutation in chickpea for Ascochyta blight resistance. Proc. of the Symposium on the role of induced mutations in crop improvement.
- 7. Reddy, M.V., Y.L. Nene, and K.B. Singh. 1980. Field screening of chickpeas for resistance to Ascochyta blight. International Chickpea Newsletter.
- 8. Raddy, M.V., Y.L. Nene, and J.P. Verma. 1980. Pea leaf-roll virus is the causal agent of chickpea stunt. International Chickpea Newsletter.

Accepted

- 1. Haware, M.P., and Y.L. Nene. 1980. Phoma blight A new disease of chickpea. Plant Disease.
- 2. Haware, N.P., and Y.L. Nene. 1980. Sources of resistance to wilt and root rots of chickpea. International Chickpea Newsletter.
- 3. Nene, Y.L., M.P. Haware, and M.V. Reddy. 1980. Techniques to screen for resistance to some important chickpea diseases. ICRISAT Information Bulletin.
- 4. Singh, K.B., G.C. Hawtin, Y.L. Nene, and M.V. Reddy. 1980. Resistance in chickpeas to Ascochyta blight. Plant Disease.

Communicated

1. Haware, M.P., and Y.L. Nene. 1980. Symptomless carriers of chickpea wilt fungus. Plant Disease.