



Prevalence and potential for aflatoxin contamination in groundnuts and peanut butter from farmers and traders in Nairobi and Nyanza provinces of Kenya

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ABSTRACT

Objective: Most of the peanut butter marketed in Nairobi is processed in cottage industry and its aflatoxin contamination status has not been documented. This study was therefore conducted to determine the status of aflatoxin contamination in groundnuts and peanut butter in Nairobi and Nyanza.

Methodology and results: Eighty two fresh samples comprising raw and roasted groundnuts and peanut butter were obtained from market outlets and cottage processors in Nairobi and Nyanza regions. The marketers and processors were asked for information on the source of groundnuts. The incidence of *Aspergillus* section *Flavi* was determined using standard laboratory methods. Defective nuts in raw groundnuts were determined by manual sorting. Aflatoxin analysis was done using competitive ELISA technique. Groundnuts in Nairobi were imported from Malawi while those Nyanza were grown in the region. The fungal species isolated from the samples were: *Aspergillus flavus* (L and S strains), *A. parasiticus*, *A. niger*, *A. tamari*, *A. alliaceus*, *A. caelestus* and *Penicillium* spp. The percentage of defective nuts among all unsorted groundnuts ranged from 0.0% to 26.3%. The mean percent defective nuts was higher for Nairobi samples than Nyanza. Aflatoxin levels in all samples ranged from 0 to 2377.1 µg/kg. The mean aflatoxin level was higher for raw samples from Nairobi than Nyanza. The source of groundnuts and defective nuts were positively associated with aflatoxin levels.

Conclusions and application of findings: The source of groundnuts and presence of defective nuts were identified as the main factors influencing increased aflatoxin contamination in the cottage industry. Mechanisms for inspection and certification of imported groundnuts should be put in place accompanied by effective monitoring for compliance to set aflatoxins standards. All the market players should sort their groundnuts before selling or processing in order to reduce aflatoxin contamination of peanut butter.

Key words: Aflatoxin, cottage industry, groundnut, peanut butter.

INTRODUCTION

Groundnut (*Arachis hypogaea* L.) is an oilseed produced all over the world and is also consumed in form of snacks such as salted nuts, confectionary sauce, and peanut butter (Campos-Mondragón *et al.*, 2009). Botanically, groundnut is a legume although it is widely identified as a nut and has similar nutrient profile with tree nuts (Ros, 2010). Peanut butter is made by grinding dry roasted groundnuts into a paste (Mutegi *et al.*, 2009). Groundnuts that are used as raw materials for peanut butter processing are liable to colonization by fungal molds during handling, storage and transportation, exposing them to the risk of contamination with aflatoxin (Polixeni & Panagiota, 2008; Mutegi *et al.*, 2012). Groundnuts can be contaminated with aflatoxin pre- and post-harvest and the risk of contamination increases along the marketing chain due to poor handling practices (Kladpan *et al.*, 2004; Kaaya *et al.*, 2006). The main aflatoxin producing fungi in groundnuts are *Aspergillus flavus*, *A. parasiticus* and *A. nomius* (CAST, 1998) which mostly infect groundnuts as a complex (Varga *et al.*, 2012). Mutegi *et al.* (2012) reported *A. flavus* L-strain, *A. flavus* S-strain, and *A. niger* as the major fungal pathogens infecting groundnuts and groundnut products from Busia and Homa Bay districts in Western Kenya. *Aspergillus flavus* and aflatoxin have been reported in groundnuts in Sudan (Omer, 2001; Omer *et al.*, 2001) and Egypt (Abdelhamid, 1990) and in peanut butter in Sudan (Eshafie *et al.*, 2011). *Aspergillus flavus*, the main producer of aflatoxin, only grows in groundnuts when the moisture content exceeds 9% and has optimum growth conditions of between 25 and 30°C, and water activity of 0.99 with a minimum of 0.83_{a_w}, while production of aflatoxin occurs optimally at 25°C and 0.99_{a_w} with a minimum of 0.87_{a_w} (Ribeiro *et al.*, 2006). The warm and humid environmental conditions in Africa are ideal for growth of *A. flavus* making aflatoxin contamination of food, including groundnuts, a widespread problem across the continent (Gordon, 2003; Bankole *et al.*, 2006; Wagacha & Muthomi, 2008). Aflatoxins are among the most prevalent carcinogens (Ribeiro *et al.*, 2006). Acute exposure of humans to aflatoxins leads to outbreak of a disease condition known as

aflatoxicosis contributing significantly to the disease burden in Africa (Mehan *et al.*, 1991; Gordon, 2003). Chronic effects of aflatoxins result from continuous exposure to relatively low levels of toxins over a prolonged period (Mehan *et al.*, 1991). Some of the common effects of chronic aflatoxicosis include impaired food conversion, slower rates of growth, and a decrease in absorption of various micronutrients (Jolly *et al.*, 2007). Aflatoxin contamination of groundnuts therefore poses a risk to human health and has been identified as a major constraint to trade in Africa (Mutegi *et al.*, 2009). In a recent study (Mutegi *et al.* 2012), 37% of groundnuts and their products including peanut butter and peanut flour sampled from Nairobi, Nyanza and Western Kenya did not meet the 10 µg/kg total aflatoxin limit set by the Kenya Bureau of Standards (KEBS, 2007). Peanut butter is one of the main products of groundnut processing (Campos-Mondragón *et al.*, 2009) which is used as a sauce or spread in many households in Kenya (Mutegi *et al.*, 2010). Most of the peanut butter consumed in Nairobi is produced in the cottage industry (Mutegi *et al.*, 2010). If raw groundnuts are contaminated with aflatoxins, there is a high risk of exposure to the consumer through consumption of peanut butter processed from such groundnuts. Practices such as poor storage and handling within the peanut butter cottage industry could contribute to further aflatoxin contamination of peanut butter.

There are many cottage industries in Nairobi that produce peanut butter (Mutegi *et al.*, 2010). High aflatoxin levels up to 22 µg/kg in groundnut products such as roasted nuts and peanut butter have been reported in Nairobi (Mutegi *et al.*, 2010). Previous studies have not investigated the status of aflatoxin contamination in the peanut butter cottage industry in Nairobi. The objectives of this study were therefore to: (i) establish the aflatoxin contamination of various products from the peanut butter cottage industry in Nairobi and Nyanza; (ii) establish the aflatoxin contamination of raw groundnuts from market outlets that are used to process peanut butter in the cottage industry in Nairobi.

MATERIALS AND METHODS

Study sites: The study was conducted in Nairobi and Nyanza (Kisii, Rongo, Homabay and Ndhiwa districts) provinces of Kenya. Nairobi is a major trade center for groundnuts with many cottage industries that process peanut butter. Demand for peanut butter in Nairobi is high. Nyanza province is the main groundnut producing region in Kenya and it would therefore be expected to be the main source of raw groundnuts for processing peanut butter in Nairobi, hence, it was chosen as a study site.

Sample collection: Twelve peanut butter cottage processors and 22 groundnut traders in Nairobi were involved in the study. Additionally, groundnut samples were taken from 24 farmers and traders from four districts in Nyanza. A list of 22 peanut butter cottage processors in Nairobi was provided by Kenya Bureau of Standards (KEBS), the standards regulatory body in Kenya. The 22 processors were contacted with 12 consenting to participate in the study thereby constituting the study group within the peanut butter cottage industry in Nairobi. The cottage processors provided information on where they sourced their raw groundnuts, data which was used as the basis of identifying the traders. Nyamakima and Gikomba were identified as the major groundnut markets in Nairobi.

Sampling of groundnuts and groundnut products: A total of 82 peanut samples were randomly taken from different market outlets which were purposively selected in Nairobi and Nyanza regions as follows: 24 raw groundnut samples from retailers in Nyanza region; 23 unsorted raw groundnut samples from wholesalers in Nyamakima market in Nairobi and 11 sorted raw groundnut samples from wholesalers in Gikomba market in Nairobi. Five (5) raw groundnuts, 8 roasted groundnuts, and 11 peanut butter samples were obtained from the cottage processors in Nairobi. For wholesalers who stocked large groundnut lots (> than 40 bags), a 1 kg representative sample was obtained by sampling different portions of a consignment. At least nine subsamples were obtained from a 100 kg sack of groundnuts using a sampling probe. Three subsamples were obtained from the top, middle, and bottom portions of the sack, respectively. The subsamples were thoroughly mixed to form a representative sample from each wholesaler. For the cottage processors whose batches were small (handling less than 300 kg of groundnuts per month), each batch of groundnuts was mixed thoroughly for homogeneity before drawing a 1 kg sample. The samples were delivered to the laboratory for analysis in Kraft papers. For peanut butter, two packaged containers weighing 400 grams each were randomly

picked from a batch. All the samples were stored for a week at 4°C in their original form before analysis.

Determination of defective nuts: The presence of defective nuts in unsorted raw groundnuts was determined by manual sorting according to KEBS standards (KEBS, 2007). The 1kg sample obtained from the market was mixed thoroughly to ensure homogeneity. A 250g sub-sample was drawn and all the defective nuts sorted by hand and weighed. The proportion of defective nuts per sample was calculated as the weight of defective nuts (g) divided by 250g, and multiplied by 100. The defects were further classified as broken, discolored, or shriveled.

Microbial assay: Fungal species were isolated on the semi selective modified dichloran rose Bengal (MDRB) agar (Horn & Dorner, 1998). The medium was prepared by mixing 10 g glucose, 2.5 g peptone, 0.5 g yeast extract, 1 g KH₂PO₄, 0.5 g MgSO₄·7H₂O, 20 g agar and 25 mg Rose Bengal in 1 liter of distilled water. The medium was autoclaved for 20 minutes at 121°C and 15 psi and cooled in a water bath to 50°C. To inhibit bacterial growth and ensure the medium was semi-selective for *Aspergillus* section *Flavi*, 10 ml of 4 mg/l dichloran (in acetone), 10 ml of 40 mg/l streptomycin (in distilled water) and 10ml of 1 mg/l chlorotetracycline (in distilled water) were added to the medium through a sterile 0.25 µm syringe filter after cooling to 50°C. The medium was then poured into 90 mm-diameter sterile Petri plates and allowed to settle overnight in a sterile hood before use. One gram of ground sample or paste was weighed into a sterile dilution tube and 9ml of sterile peptone water added to make the first dilution. The sample suspension was then vortex-mixed to ensure homogeneity. A second dilution was prepared by drawing 1 ml from the first dilution and adding it into 9ml of sterile peptone water. Then, 0.1ml of each sample dilution was inoculated onto the MDRB agar medium in six replicates per sample for each dilution, and incubated for 6 days at 30°C.

Reference cultures of *Aspergillus* section *Flavi* obtained from the Centre for Agricultural Bioscience International (CABI), Plant Pathology Laboratory, were cultured on the MDRB medium at the time the samples were plated. The reference cultures included *A. flavus* (S and L strains), *A. parasiticus*, *A. tamarii*, *A. alliaceus*, and *A. caelestus*. The shape, margins, elevation, pigmentation, texture, and pattern of growth of the fungal colonies were used in identifying the *Aspergillus* species and for *A. flavus*, distinguishing between the L and S strains. A tally counter was used to count the total number of colony forming units (CFU) per Petri plate. The number of colonies of *A.*

flavus (S and L strains), *A. parasiticus*, *A. tamarii*, *A. alliaceus*, and *A. caeleus* was recorded and their population determined per gram of groundnuts. The population of *Penicillium* spp. and other fungal species was also recorded for each sample.

Determination of aflatoxin levels: A representative 200g sub-sample was drawn from each homogenous sample and ground into a fine powder using a dry mill kitchen grinder (Kanchan Multipurpose Kitchen Machine, Kanchan International Limited, Mumbai, India). Grinding was not required for peanut butter paste samples. The powder (or paste) was then triturated in 70% methanol (v/v 70 ml absolute methanol in 30 ml distilled water) containing 0.5% w/v potassium chloride in a blender, until thoroughly mixed. The extract was transferred to a conical flask and shaken for 30 min at 300 rpm. The extract was then filtered through Whatman No. 41 filter paper and diluted 1:10 in phosphate buffered saline containing

500 µl/l Tween 20 (PBS-Tween) and analyzed for aflatoxin level based on Enzyme Linked Immunosorbent Assay, ELISA (Waliyer & Reddy, 2009). The aflatoxin results were classed based on EU (≤ 4 µg/kg) and KEBS (≤ 10 µg/kg) standards.

Statistical analyses: Data were subjected to analysis of variance (ANOVA) using the SAS general linear model (GLM) procedure (Statistical Analysis System, The SAS Institute, Cary, NC, USA). The Tukey's studentized range (Honestly Significant Difference, HSD) test was used as post ANOVA technique for comparison of pairs of means. The t-test procedure was used to compare two means. Skewed data were transformed to \log_{10} before data analysis and separation of means. Aflatoxin levels were compared with the maximum safe limits based on European Union (EC, 2010) and KEBS (KEBS, 2007) standards.

RESULTS AND DISCUSSION

Sources and types of groundnuts used in the cottage industry in Nairobi: Groundnuts used in the peanut butter cottage industry in Nairobi were bought from wholesalers within the city who in turn sourced the groundnuts from Malawi. The demand for groundnuts and groundnut products in Nairobi is high and therefore the traders said that relied on imported groundnuts for their supply since it was more sustainable than sourcing locally. Groundnuts produced in Nyanza were traded in the region and did not reach the Nairobi wholesale markets and consequently the traders reported that they imported groundnuts from Malawi, in the study period. This is consistent with previous reports that groundnuts traded in Nairobi are hardly produced in Kenya but are purchased from neighboring countries (Mutegei *et al.*, 2012). The groundnut traders in Nairobi comprised of wholesalers and retailers. The groundnut wholesalers were found in Nyamakima and Gikomba markets; with the latter doubling up as a retail market where groundnuts were sorted before selling. The raw groundnuts in Gikomba had been sourced from wholesalers in Nyamakima market. Other retail markets were found in Burma, Maringo, Kawangware, and Kikuyu. Different types of groundnuts were sampled from the cottage industry and markets in Nairobi (Fig. 1). The local names of the groundnut types, as used by the traders were: Pink regular, Pink large, Red regular, Red small and Red mixed. Pink regular and Red regular were the

predominant types (45%) of groundnuts both in Nairobi and Nyanza markets.

Defective kernels in unsorted raw groundnuts: There was no significant ($p \geq 0.05$) difference in the proportion of defective nuts among the unsorted groundnut samples from markets in Nairobi and Nyanza (Table 1). Broken nuts were the most frequent (47%) type of defective nuts possibly due to poor handling practices e.g. threshing and transportation. Whitaker *et al.* (1997) reported that broken nuts are a high risk factor to aflatoxin contamination. Overall, discolored and shriveled nuts constituted 23 and 30% of the defects, respectively. The proportion of defective nuts did not meet the KEBS threshold of 2% for raw groundnuts (KEBS, 2007). Higher than the recommended proportion of defective nuts could lead to higher aflatoxin contamination levels in groundnuts and groundnut products. Turner *et al.* (2005) reported that hand sorting reduced the amount of defective nuts which translated to reduced aflatoxin levels. The researchers observed that use of simple methods such as hand sorting, proper sun-drying on mats, and storage in natural-fiber bags reduced aflatoxin exposure in West Africa villages by more than 50%. A similar study by Galvez *et al.* (2003) developed a manual sorting procedure where raw groundnuts were blanched at 140°C for 25 minutes in preheated roasters, cooled, de-skinned manually and then sorted. The procedure reduced aflatoxin levels in groundnuts from 300 µg/kg to undetectable levels.



Figure 1: Major groundnut types traded in Nairobi and Nyanza markets in Kenya. (A) Red regular, (B) Red mixed, (C) Red small, (D) Pink regular, (E) Pink large.

Table 1: Proportion (%) of defective nuts and types of defects in unsorted raw groundnut samples from markets in Nairobi and Nyanza regions.

Sample source	Sample type	Defective nuts ¹	Types of defects			n
			Broken	Shriveled	Discolored	
Cottage industry	Pink regular	13.7 ± 1.7a	48.0	30.0	22.0	3
	Red small	7.7 ± 0.0 a	50.0	27.0	23.0	1
Nairobi wholesale outlets	Pink regular	15.2 ± 1.7 a	45.5	32.3	22.2	11
	Red regular	16.8 ± 1.9 a	50.3	26.7	23.0	12
Nyanza retail outlets	Pink large	15.9 ± 0.2 a	45.0	32.1	22.9	3
	Pink regular	12.6 ± 1.9 a	47.6	28.7	23.7	9
	Red regular	12.5 ± 7.1 a	44.1	33.9	22.0	9
	Red mixed	6.6 ± 0.9 a	46.0	30.5	23.5	3
Mean		12.63 ± 1.9	47.1	30.2	22.8	

¹Values are means ± SE. Values accompanied by the same letter within the column are not significantly different ($p \geq 0.05$).

Population of fungal species in groundnuts and peanut butter: Seven *Aspergillus* species were isolated from groundnut and peanut butter samples (Table 2). They included *A. flavus* S strain (mean = 1230 CFU/g), *A. flavus* L strain (mean = 689 CFU/g), *A. parasiticus* (mean = 122 CFU/g), *A. niger* (mean = 81 CFU/g), *A. tamarii* (mean = 72 CFU/g), *A. alliaceus* (mean = 25 CFU/g), and *A. caelestis* (mean = 1 CFU/g). The population of *A. flavus* S strain was significantly ($p \leq 0.05$) higher in unsorted groundnuts from Nairobi markets than from Nyanza. It takes a short time for groundnuts in Nyanza to reach the market as opposed to the long time taken for groundnuts from Malawi to reach Nairobi markets. The long time can predispose groundnuts to infection by the *Aspergillus* fungi especially if handling conditions are poor for instance during transport.

Penicillium spp. were isolated in high frequency in most of the samples (Mean = 19026 CFU/g). Other fungi isolated from the groundnuts included *Fusarium* spp., *Trichoderma* spp., *Rhizopus* spp. and *Mucor* spp. The presence of these other fungi in large amounts is a clear indication of poor handling, e.g. poor hygiene and storage, which exposes the nuts to conditions favorable for fungal growth. However, the occurrence of a mixture of fungal organisms is deemed to reduce mycotoxin production (Mirocha et al., 1974; Raper et al., 1965). Some *Fusarium* strains can produce mycotoxins, such as fumonisins, trichothecenes and zearalenones, which are associated with mycotoxicoses in humans or domestic animals (Antonio et al., 2003). Toxicogenic *Penicillium* species produces mycotoxins such as ochratoxins, citrinin, and patulin (Antonio et al., 2003).

Therefore the presence of *Penicillium* in groundnuts poses a serious problem to human health. The occurrence of *Rhizopus* spp. and *Mucor* spp. can be used as indicators of potential presence of aflatoxins (Kane et al., 2006). No fungal pathogens were isolated from roasted groundnuts which could be attributed to heat processing during roasting. During roasting of groundnuts for peanut butter production, nuts are exposed to dry heat at temperatures of 120 – 150°C and above (Okello et al., 2010). However, fungal pathogens were isolated from peanut butter which is made immediately after the roasting step. This could be attributed to recontamination during processing. Indirect contamination is common in oilseeds and cereals in Africa and is the main entry point of mycotoxins in human dietary systems (Smith & Moss, 1985).

Incidence of *Aspergillus* section *Flavi* in groundnuts and peanut butter: The incidence of *Aspergillus* section *Flavi* in various samples ranged from 0 to 80% (Table 3). Unsorted raw groundnuts sampled from Nairobi were the most infected: *A. flavus* L strain (69.5%), *A. flavus* S strain (65.0%), and *A. parasiticus* (30.0%). Raw groundnuts from the cottage industry recorded the highest incidence (80%) of *A. flavus* S strain followed by unsorted groundnuts from Nairobi wholesale market (65%). *Aspergillus flavus* L strain was the most frequently isolated (69.5%) pathogen from unsorted groundnut samples from Nairobi wholesale market. The incidence of *A. flavus* (S and L strains), and *A. parasiticus*, the three main aflatoxin producers, was higher in raw groundnuts sampled from markets in Nairobi than from Nyanza. Additionally, the incidence of *A. flavus*

(S and L strains) was higher in unsorted groundnuts than in sorted groundnuts from Nairobi wholesale market. This could be attributed to the fact that unsorted groundnuts are more prone to infection by aflatoxigenic fungi than sound kernels (Whitaker et al., 1997). Sorting groundnuts removes defective nuts and is effective in reducing aflatoxin contamination (Turner et al., 2005; Galvez et al., 2003). Members of *Aspergillus* section *Flavi* mainly *A. flavus* and *A. parasiticus* are the major producers of aflatoxin in groundnuts (Passone et al., 2010). *Aspergillus flavus* produces only B-type aflatoxins and it is the second leading cause of human aspergillosis (Hedayati et al., 2007; Pasqualotto & Denning, 2008; Krishnan et al., 2009) while *A. parasiticus* produces both B- and G-type aflatoxins (Ehrlich et al., 2004; Frisvad et al., 2005).

Table 3: Incidence (%) of *Aspergillus flavus* (L and S strains) and *A. parasiticus* in groundnut and peanut butter samples from Nairobi and Nyanza provinces, Kenya.

Sample source	Sample description	<i>A. flavus</i> L strain	<i>A. flavus</i> S strain	<i>A. parasiticus</i>	Total (n)
Cottage industry	Raw nuts	80.0	40.0	40.0	5
	Roasted nuts	0.0	0.0	0.0	8
	Peanut butter	9.0	9.0	0.0	11
Nairobi wholesale	Unsorted nuts	65.0	69.5	30.0	23
	Sorted nuts	54.5	54.5	27.0	11
Nyanza markets	Unsorted nuts	33.0	12.5	17.0	24
Mean		40.3	30.9	19.0	

Aflatoxin contamination levels of groundnuts and peanut butter: There was variation in aflatoxin levels among groundnut and peanut butter samples ranging from 0.0 to 2377.1 µg/kg (Table 4). The wide variation in aflatoxin contamination in peanut butter could be attributed to contamination during processing in the cottage industry and using groundnuts that were already infected by aflatoxin-producing fungi. For instance, the processor whose peanut butter recorded the highest aflatoxin contamination did not grind roasted nuts immediately after roasting but stored them to grind later based on customer purchase orders. It was also observed that the processor had not cleaned the grinder after the previous peanut

butter production. This practice could predispose the peanut butter to increased aflatoxin levels. On the other hand, the processor from whose peanut butter aflatoxins were not detected observed hygiene in processing and was also a trained food technologist. Defective nuts are more likely to be infected with aflatoxin-producing fungi than sound nuts (Whitaker et al., 1997). The presence of defective nuts in the current study was positively correlated ($r = 0.412$, $p = 0.0092$) with aflatoxin levels. The wide variation in aflatoxin contamination in unsorted raw groundnuts from Nyamakima market can therefore be attributed to the presence of varying degree of total defective nuts among the samples. The defective nuts are high

risk components which contain higher concentration of aflatoxins. Among the samples from Nyamakima, the samples whose aflatoxin levels were below 10 µg/kg had an average of 12.43% defective nuts while those whose aflatoxin levels exceeded 10 µg/kg had an average of 17.29% defective nuts. Among the raw groundnut samples, the Red types had lower total aflatoxin levels than the Pink types, though not significant. However, there was no significant ($P \geq 0.05$) correlation between groundnut type and aflatoxin levels. Aflatoxin contamination increased along the value chain within the peanut cottage industry in Nairobi with raw groundnuts being the least contaminated followed by roasted nuts whereas peanut butter was the most contaminated. This was evidence that considerable aflatoxin contamination occurred during peanut butter processing. Improper handling practices during storage and processing have been reported to increase aflatoxin contamination in peanut butter in Sudan (Hanafi, 1987; Omer, 2001). In the present study, poor handling practices that were observed included failure to clean the grinder after peanut butter production, storing roasted nuts for long before grinding, and cooling of roasted nuts on the floor and/or outside in the open. Aflatoxin levels were significantly ($P \geq 0.05$) higher for raw groundnuts from Nairobi than from Nyanza. This implies that the source of raw groundnuts used in the peanut butter cottage industry is a factor that predisposes peanut butter to aflatoxin contamination. It is possible that raw groundnuts were handled poorly in the country of origin or aflatoxin contamination occurred during transit. This

observation was consistent with the findings of an earlier study by Mutegei *et al.* (2012) that reported higher aflatoxin levels in groundnut products sampled from Nairobi than from Nyanza. Negligence during post-harvest handling of groundnuts by market players along the market chain could lead to increase in aflatoxin contamination.

Roasting of groundnuts has been shown to reduce aflatoxin levels (Hamada & Megalla, 1982; Yazdanpanah *et al.*, 2005). The degree of reduction is dependent on heating time, temperature and the initial aflatoxin concentration (Rustom, 1997; Yazdanpanah *et al.*, 2005; Arzandeh and Jinap, 2011). Ogunsanwo *et al.* (2004) observed that roasting significantly reduced the levels of aflatoxin B₁ and G₁ in groundnuts. In the study, groundnuts were dry-roasted at 150°C for 30 minutes which resulted in a 70% and 79.8% reduction of AFB₁ and AFG₁, respectively. However, roasting did not affect aflatoxin level in the peanut butter cottage industry in Nairobi. This could be attributed to recontamination of roasted groundnuts with aflatoxin-producing fungi due to poor hygiene practices in the cottage industry.

Correlation between *Aspergillus* section *Flavi* and aflatoxin contamination: There was a positive association ($r = 0.3435$) between the incidence of *A. flavus* S strain and $r = 0.4254$ for *A. flavus* L strain and total aflatoxin level (Fig. 2). *Aspergillus parasiticus* was also positively correlated with total aflatoxin level. However, there was no significant ($p \geq 0.05$) association between groundnut types and aflatoxin level.

Table 4: Aflatoxin level (µg/kg) of groundnut and peanut butter samples from various market outlets in Nairobi and Nyanza provinces, Kenya.

Sample source	Sample description	Sample type	Aflatoxin level (µg/kg)		n
			Range	Mean	
Cottage industry	Raw nuts	Pink regular	0.0 – 52.4 ^a	18.3	3
		Red regular	-	5.0	1
		Red small	-	nd	1
	Roasted nuts	Red regular	2.4 - 297.7	54.8	8
	Peanut butter	Paste	0.0 - 2377.1	318.3	11
Nairobi wholesale Outlets	Unsorted nuts	Pink regular	0.0 - 364.7	111.2	11
		Red regular	0.0 - 276.1	89.1	12
		Sorted nuts	Pink regular	0.0 - 82.4	24.0
	Red regular	2.0 - 9.2	5.3	5	
	Red small	6.0 – 7.8	6.9	2	
Nyanza retail Outlets	Unsorted nuts	Pink large	3.7 - 128.8	71.6	3
		Pink regular	0.0 - 229.8	44.9	9
		Red regular	0.0 - 14.0	1.9	9
		Red mixed	-	nd	3

^aLimit of detection (LOD) = 0.5 µg/kg; 0.0 indicates below LOD; nd = not detected.

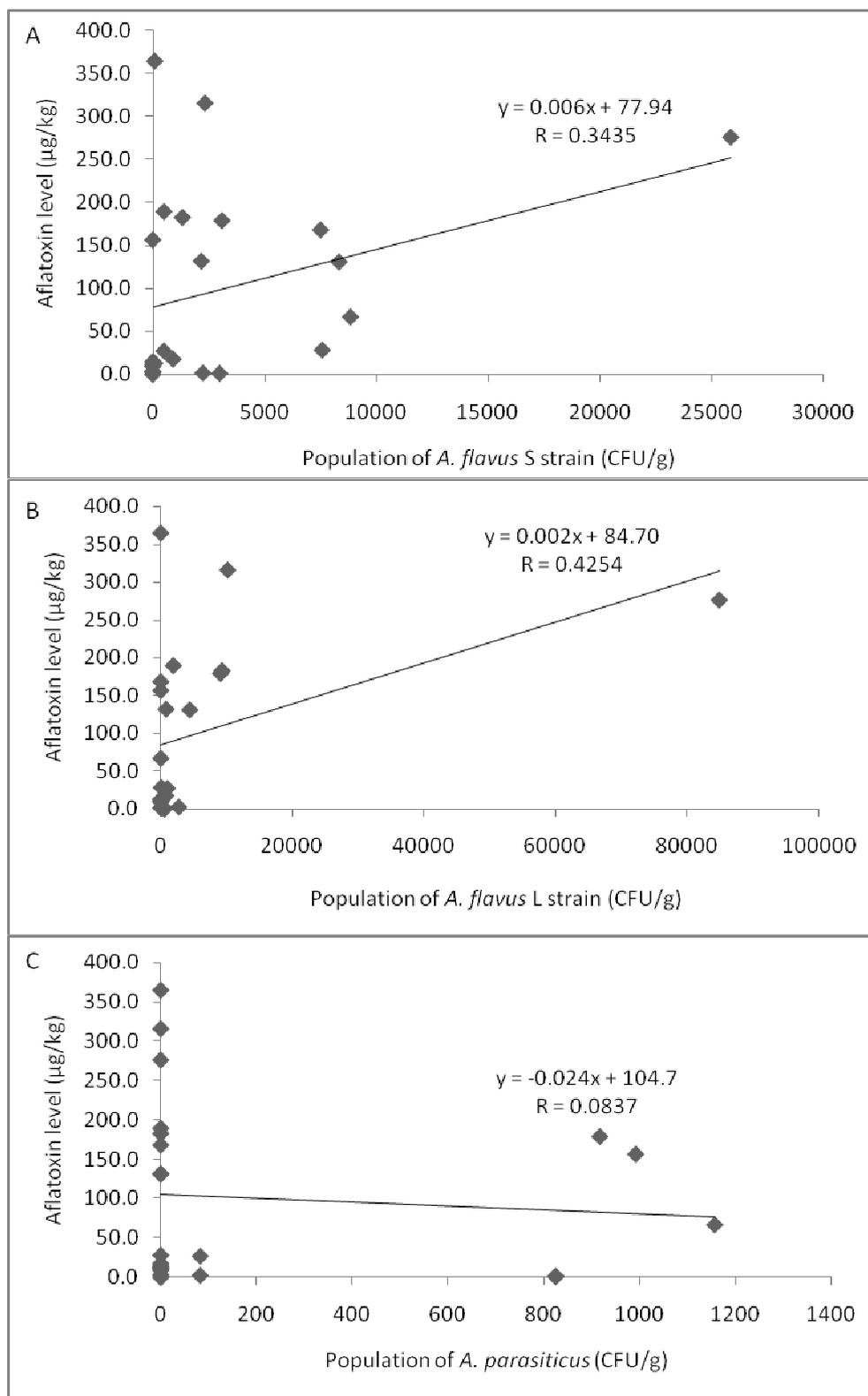


Figure 2: Scatter plot for the association between aflatoxin level (µg/kg) and *A. flavus* S strain (A); *A. flavus* L strain (B); and *A. parasiticus* (C).

Conformance of the cottage industry to regulatory standards: Aflatoxin levels in 61 and 43% of the samples (Table 5) were above the EU and KEBS regulatory thresholds of 4 and 10 µg/kg, respectively,

implying that the samples would have been rejected under the respective regulations. This proportion of unsafe products is substantial in terms of economic loss if the products were to be discarded. However,

this rarely happens in Kenya due to poor monitoring and enforcement of regulatory standards. Groundnuts from Nyanza had the highest proportion (71%) of samples that met the European safety limits. Unsorted raw groundnuts sampled from Nairobi wholesale markets and peanut butter from the cottage industry had the highest proportion (74% and 73%, respectively) of samples with aflatoxin levels above the KEBS regulatory limit of 10 µg/kg. On the hand, sorted raw groundnuts from markets in Nairobi had the highest proportion (82%) of samples with aflatoxin levels below 10 µg/kg. The high proportion of unsafe products therefore implies that consumers of groundnuts and groundnut products in Kenya are invariably exposed to unsafe levels of aflatoxin. Besides the health implications, the high levels of

aflatoxin in groundnuts are a barrier to trade. Groundnuts exported from Sudan to EU have previously been rejected owing to failure to meet aflatoxin standards (Hanafi, 1987). Thus, the purpose of reducing aflatoxin contamination levels in groundnuts should be to minimize food safety concerns and also for economic reasons (Ozay *et al.*, 2008). A high proportion (80%) of raw groundnut samples from the cottage industry had aflatoxin levels below 10 µg/kg. Whereas aflatoxin level in half of the roasted groundnut samples from the cottage industry was below the 10 µg/kg limit, contamination level of 73% of peanut butter samples was above 10 µg/kg. This showed increase in aflatoxin contamination within the peanut butter cottage industry.

Table 5: Proportion (%) of groundnut and peanut butter samples with aflatoxin levels in different categories based on EU and KEBS standards.

Sample source	Sample description	Aflatoxin level category (µg/kg)		
		≤ 4	≤ 10	> 10
Cottage industry	Raw nuts	60	80	20
	Roasted nuts	25	50	50
	Peanut butter	18	27	73
Nairobi wholesale Outlets	Unsorted nuts	22	26	74
	Sorted nuts	36	82	18
Nyanza retail outlets	Unsorted nuts	71	75	25
Mean		39	57	43

CONCLUSION

Presence of defective nuts and the source of raw groundnuts predispose peanut butter to aflatoxin contamination in the cottage industry. Farmers need to be enlightened on proper ways of handling groundnuts during harvesting and threshing to avoid breakage of the nuts in order to reduce fungal infection. Sorting should be made mandatory for the groundnut marketers with effective monitoring to ensure compliance and punitive measures should be appended to non compliance. The Kenya Bureau of Standards should strengthen monitoring for

compliance to set standards and enforce regulations in the Nairobi peanut butter cottage industry. Awareness needs to be raised among the peanut butter processors to observe safety standards in processing so as to reduce aflatoxin contamination. Mechanisms for inspection and certification of imported groundnuts should be established accompanied by regular monitoring for aflatoxin levels of the imported groundnuts to ensure that they comply with set limits.

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Table 2: Population (CFU/g peanuts) of fungal species isolated from groundnuts and peanut butter sampled from Nairobi and Nyanza provinces in Kenya.

Sample source	Sample description	<i>A. flavus</i> L strain	<i>A. flavus</i> S strain	<i>A. parasiticus</i>	<i>A. niger</i>	<i>A. tamaritii</i>	<i>A. alliaceus</i>	<i>A. caelestus</i>	<i>Penicillium</i> spp	Others	Total (n)
Cottage industry	Raw nuts	1132a	582ab	265a	17a	34a	0a	0a	64697a	26558a	5
	Roasted nut	0b	0b	0a	0a	0a	0a	0a	238c	884b	8
	Peanut butter	76b	8b	0a	0a	0a	0a	0a	323bc	3023ab	11
Nairobi wholesale	Unsorted nuts	5505a	3229a	212a	359a	398a	149a	4a	17059ab	34209a	23
	Sorted nuts	106ab	204ab	204a	61a	0a	0a	0a	3446abc	31262a	11
Nyanza retail	Unsorted nuts	559ab	111b	52a	49a	0a	0a	0a	14688bc	21954a	24
Mean		1230	689	122	81	72	25	1	16742	19648	

Means accompanied by the same letter(s) within the column are not significantly different ($p \geq 0.05$).