

Molecular characterization of pigeonpea [Cajanus cajan (L.) Millspaugh] composite collection



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Pigeonpea [*Cajanus cajan* (L.) Millspaugh]

- Pigeonpea is diploid (2n = 22) and primarily grown for food (dry seeds or green vegetables)
- Sixth most important food legume crop and good source of protein; enriches soil; provides fodder and fuel wood; and arrests soil erosion
- Grown in over 82 countries worldwide as a field and/or backyard crop
- As a regular crop, grown only in 19 countries on 4.6 million ha producing 3.5 million tons of grain (FAOSTAT, 2005).

Pigeonpea composite collection

- Developed considering the phenotypic diversity present in the entire collection held at the ICRISAT genebank (Upadhyaya et al., 2005).
- Collection consists 1000
 accessions comprising minicore (Upadhyaya et al.,
 2006), comparator minicore
 and trait-based accessions
 along with 62 accessions of
 seven wild *Cajanus* species.



Genetic diversity in composite collection

Plant material

DNA extracted from randomly selected 12 plants per accession following a high-throughput procedure and pooled together to capture the within accession variation.

Selection of SSR markers

- Pigeonpea SSR markers being less polymorphic, all the available markers at ICRISAT initially screened on 15 diverse accessions (8 cultivated and 7 wild) that are included in the composite collection.
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- Thirty-three SSR markers showed polymorphism between at least two of the tested accessions.
- A series of artificial pools having different proportions of DNA from two genotypes developed and tested with 33 SSR markers for polymorphism.
- The coefficient of correlations analyzed between different proportion of alleles

recorded (corresponding to 12 plants per accession) and proportion of genomic DNA used for the corresponding accession.

Twenty SSR markers (Table 1)
with highly significant
correlations (r² > 0.9) identified.

selected for genotyping the composite collection						
CCB1	PGM109	PGM45	CCB10			
CCB9	PGM82	PGM102	PGM16			
PGM3	PKS21	PKS30	PKS18			

PKS25

PKS26

PGM5

PGM10

CCB7

CCB8

Molecular characterization

• PCR components of all 20 SSR markers optimized following Taguchi method (Taguchi, 1986) as described in Cobb and Clarkson (1994)

PGM101

PGM106

• Fluorescent-based multiplex genotyping system used to generate five multiplexes of four markers each

- Capillary electrophoresis with an automated system (ABI 3700) used to separate the amplified PCR products
- SSR fragment sizes called to two decimal places using the Genotyper v3.7 software
- Observed allelic data binned into discrete genetic units using the Allelobin program (http://www.icrisat.org/gt-bt/biometrics.htm) developed at ICRISAT based on the least squares algorithm of Idury and Cardon (1997)
- Markers produced allele sizes expected, based on the repeat motif of each of the SSR markers
- Less than 5% missing data (ie, marker x genotype) in the dataset.

Data analysis

- Data analysis completed on 12 SSR markers
- DARwin 5.0 Structure program (Perrier et al., 2003) used to determine the population structure of the composite collection
- Forty-five accessions with high missing values excluded from data analysis
- Principal coordinate analysis done considering the biological status of the crop (cultivated and wild)
- Cervus 2.0 software used to determine the allele frequencies and PIC values.

Results

- Preliminary analysis detected 144 alleles, ranging from 5 (PGM109) to 24 (CCB8) with a mean of 12.0 alleles per locus (Table 2)
- Mean Polymorphic Information Content (PIC) value was 0.345, ranging from 0.09 to 0.53, (Table 2).
- Phylogenetic tree constructed based on biological status revealed that both cultivated and wild accessions formed distinct clusters (Fig. 1)

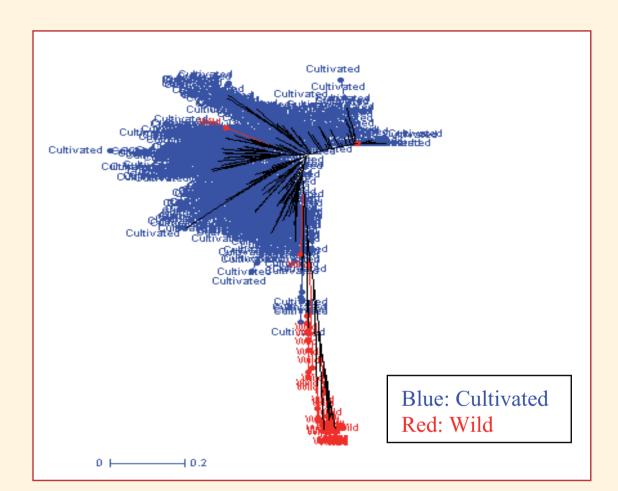


Fig. 1. Tree diagram of 955 accessions with 12 SSR markers.

 All the accessions belonging to seven wild Cajanus species grouped together in a separate cluster except few, which grouped with the cultivated accessions.

Table 2. Information on 12 SSR primers							
Repeat Unit	Quality Index	No. of Accessions Genotyped	No. of Alleles	PIC Values			
(CA)10	0.26	1000	20	0.46			
(CT)22	0.46	951	18	0.53			
(GAA)5G (GAA)5	0.42	976	11	0.50			
(AC)7	0.14	984	8	0.43			
(CT)16	0.24	1000	17	0.47			
(AAG)13	0.31	1000	11	0.33			
(CTT)8	0.18	1000	5	0.44			
(CT)30	0.27	961	24	0.47			
(CT)6TT (CT)2	0.23	997	8	0.13			
(GAA)6	0.23	948	8	0.13			
(AGA)5	0.22	986	7	0.09			
(TC)8	0.26	962	7	0.17			
	Repeat Unit (CA)10 (CT)22 (GAA)5G (GAA)5 (AC)7 (CT)16 (AAG)13 (CTT)8 (CTT)8 (CT)30 (CT)6TT (CT)2 (GAA)6 (AGA)5	Repeat Quality Unit Index (CA)10 0.26 (CT)22 0.46 (GAA)5G 0.42 (GAA)5 (AC)7 0.14 (CT)16 0.24 (AAG)13 0.31 (CTT)8 0.18 (CTT)8 0.18 (CT)30 0.27 (CT)6TT 0.23 (CT)2 (GAA)6 0.23 (AGA)5 0.22	Repeat Unit Index Genotyped (CA)10 0.26 1000 (CT)22 0.46 951 (GAA)5G 0.42 976 (GAA)5 (AC)7 0.14 984 (CT)16 0.24 1000 (AAG)13 0.31 1000 (CTT)8 0.18 1000 (CT)30 0.27 961 (CT)6TT 0.23 997 (CT)2 (GAA)6 0.23 948 (AGA)5 0.22 986	Repeat Unit Quality Index No. of Genotyped Genotyped Alleles (CA)10 0.26 1000 20 (CT)22 0.46 951 18 (GAA)5G (GAA)5 0.42 976 11 (AC)7 0.14 984 8 (CT)16 0.24 1000 17 (AAG)13 0.31 1000 11 (CTT)8 0.18 1000 5 (CT)30 0.27 961 24 (CT)6TT (CT)2 0.23 997 8 (GAA)6 0.23 948 8 (AGA)5 0.22 986 7			

Future plan

- Further analysis of data with remaining eight markers in progress to fully understand the genetic diversity and population structure of the composite collection
- Results from genotypic data will be used to identify a reference set of 300 diverse accessions for future use.

References

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