



Commentary

Appropriateness of biotechnology to African agriculture: *Striga* and maize as paradigms

Fred Kanampiu¹, Joel Ransom², Jonathan Gressel^{3*}, David Jewell⁴, Dennis Friesen^{1,5}, Daniel Grimanelli⁶ & David Hoisington⁶

¹CIMMYT, P.O. Box 25171, Nairobi, Kenya; ²CIMMYT, P.O. Box 5186, Lazimpat, Kathmandu, Nepal; ³Plant Sciences, Weizmann Institute of Science, Rehovot 76100, Israel; ⁴CIMMYT, Mount Pleasant, P.O. Box 163, Harare, Zimbabwe; ⁵IFDC, P.O. Box 2040, Muscle Shoals, AL35662, USA; ⁶CIMMYT, Apdo Postal 6-641, Col. Juárez, Deleg. Cuauhtémoc, CP 06600, México, D.F., México (*requests for offprints; Fax: +972-8-934-4181; E-mail: jonathan.gressel@weizmann.ac.il)

Received October 2001, accepted in revised form November 2001

Key words: acetolactate synthase, Bt genes, herbicide resistance, legume intercropping, maize, transposons, witchweed

Introduction

The witchweeds *Striga hermonthica* and *S. asiatica* decimate maize, millet, sorghum, and upland rice throughout sub-Saharan Africa where, according to FAO studies, over 100 M people lose half their crop production to this flowering, root-attaching parasite (Berner et al., 1995). *Striga* spp. are a major reason that maize yield in the 1.2 million ha sown in sub-Saharan Africa has dropped from near the world average of 4.2 T/ha in the last few decades to the present 1.3 T/ha (FAO, 2001). Although crop rotation, especially with introduced intercrops or rotational crops (Carsky et al., 2000; Khan et al., 2000; Oswald and Ransom, 2001), organic and inorganic fertilizers (Combari et al., 1990; Mumera and Below, 1993; Gacheru and Rao, 2001) can partially allay the problem, no control measure has been developed that subsistence farmers find within their financial means, or that fit well into their traditional cropping systems. Moreover, these measures require several seasons of repeated use before they begin to produce yield benefits (Ransom, 2000). Thus, despite widespread extension efforts, they have not been and are unlikely to be widely adopted, as they are not what the farmers consider 'appropriate' for their needs of providing sufficient food for their families on small, intensively cultivated holdings.

Striga can be controlled by foliar applications of existing herbicides after the flower stalk has emerged,

requiring spray equipment and too much herbicide, and would be ineffective for the current season. In addition to draining photosynthate, minerals and water (Press and Graves, 1995), *Striga hermonthica* does most of its damage to maize in Africa through phytotoxins before the weed emerges from the soil (Gurney et al., 1995). Spray applications of most herbicides would kill intercropped legumes, which are planted by many subsistence farmers in an effort to reduce risk and increase the dietary intake of protein that would otherwise come from maize alone.

Subsistence farmers in Kenya and elsewhere cultivate maize with judiciously used, small inputs of fungicide and insecticide seed dressings, and weeks later, apply a few granules of insecticide into the whorl of maize leaves to control stem borers. We reasoned that small amounts of herbicide could control the parasitic *Striga* while it is still underground, before the weed damages the crop (Abayo et al., 1996, 1998; Kanampiu et al., 2001). African farmers adopt new maize varieties and technologies having perceived value, including hybrid maize, as borne out in economic studies of fertilizer adoption in African subsistence agriculture (McCown et al., 1992).

Maize is a very important part of the human caloric intake throughout sub-Saharan Africa, and the situation of production, consumption and parasitic weed constraints is similar to that throughout the region. Kenya has an average per capita maize consumption of 103 kilograms per year of which 91% is used for food

(Pingali and Pandey, 2001). Maize yield and production in Kenya are 1.7 tons/ha and 2.7 million tons, respectively, and are on the decline, making Kenya a net importer of 427 000 tons of maize. In Kenya alone, there are over 80 000 ha of land severely infested with *Striga hermonthica*. Doubling yields from 1 ton would produce enough maize to provide an additional 400 000 people with their current average annual maize consumption. The areas badly affected by *Striga* are also the areas where many of the poorest people live with the highest percentage of maize in their diet. Farmers who no longer lose their maize to *Striga* can be expected to put more input into weeding, and to invest in some fertilizer. They will certainly see the benefit of buying herbicide-coated seed each season, and can easily achieve 3.5 T/ha. Maize imports can be reduced, and the cost of distribution cut down.

Proof of concept that biotechnology can provide the answers to *Striga*

Some transgenic (Joel et al., 1995) and tissue culture derived (Abayo et al., 1996, 1998; Berner et al., 1997) herbicide-resistant crops enable the early control of parasitic weeds before or during attachment to the host. The specific genes confer herbicide resistance through a modification of the binding site such that the herbicide does not bind to its target in the crop (Newhouse et al., 1991). Such herbicides are exuded from crop roots and kill attached *Striga* as well as its seeds in the soil before they germinate (Kanampiu et al., 2002).

We (Abayo et al., 1996, 1998; Kanampiu et al., 2001) and others (Berner et al., 1997) have developed methods of coating small amounts of acetolactate synthase inhibiting herbicides on biotechnologically produced (non-transgenic) imidazolinone-resistant (IR) maize seed. These methods considerably lower the amount of herbicide required to control *Striga* on a per hectare basis, but the herbicide concentration is very high in the vicinity of the seed, necessitating the high level of resistance conferred by such target-site mutations in ALS. The case has also been made that herbicide-resistant crops would preclude the common practice of intercropping cereals with grain legumes, reducing dietary protein intake and availability. Our studies show that seed-applied herbicide has no effect on intercropped cowpeas (*Vigna unguiculata* (L.) Walp.), yellow gram (*Vigna* spp.) (Figure 1), or field beans (*Phaseolus vulgaris* L.) (Kanampiu et al., 2002)

Table 1. Increase in yield of maize using 'in-seed' treatment with herbicides

Lightly or moderately infested sites		Heavily infested sites	
control	treated	control	treated
Tons grain/ha	# sites	Tons grain/ha	# sites
1.0	3.5	2.2	3.7
	21		17

The infestations of *Striga hermonthica* and *Striga asiatica* on the 38 sites located in 4 different countries measured in 8 different seasons where farmers' sites were low/moderately (naturally) infested, and on-station were heavily (artificially) infested in each season (with each season's data counted as a site), and the average yields presented. It is not possible to have the equivalent of hand weeded controls in farmers' field experiments with *Striga*, as the damage is done before *Striga* emergence, and thus the use of infestation levels as a comparison point. The data for imazapyr and pyriithiobac seed treatments are bulked at all concentrations used.

if the legume is sown more than 12 cm from the treated maize seed. Since such legumes are typically sown equidistant between maize hills within the row (i.e. 30 cm from each hill), or between the maize rows (spaced at 75 cm), the technique poses negligible risk to the intercrop.

Imazapyr and pyriithiobac, which gave the best results, inhibit only acetolactate synthase (ALS), an enzyme not present in mammals. They have been exhaustively tested and no toxicological problems have been found. They are vastly less toxic than beta-cyfluthrin, the insecticide used for stem borer control which has an oral LD₅₀ (rats) of 500 mg kg⁻¹ (Tomlin, 1994) versus >5000 mg kg⁻¹, (i.e. immeasurable) for imazapyr (Gagne et al., 1991), and 1000–5000 mg kg⁻¹ for pyriithiobac (Tomlin, 1994). The herbicides dissipate from the soil well before the next planting season, without any ill effect to subsequent crops. Maize yield from treated seed is more than doubled when *Striga* infestations are moderate, the yield benefit can be almost infinite when the *Striga* infestations are severe as there is near total crop loss without seed treatment. The results of multi-field trials in four countries are summarized in Table 1.

Lightly or moderately infested farmers' rainfed fields have low inherent soil fertility while the heavily infested irrigated experiment-station fields were fertilized annually, explaining why the grain yield in untreated controls was 1.0 tons/ha in low/moderately-infested farmer sites as compared to 2.2 tons/ha in heavily infested field station sites.

The on-farm yields increased on average from 1.0 to 3.5 tons/ha, by only investing US\$ 4 in herbicides. There would have to be an additional expenditure of

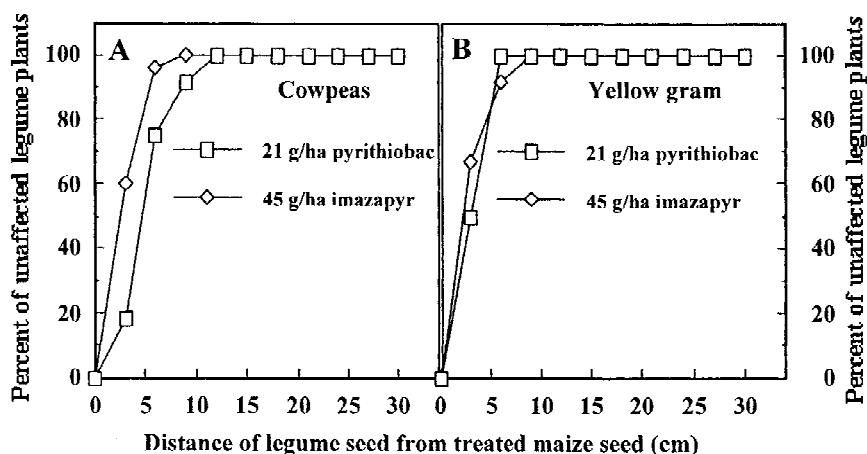


Figure 1. Lack of effect to normally-spaced intercropped legumes from herbicide treatment to seeds of herbicide resistant maize. The data are presented as the average number of completely unaffected (healthy) yellow gram and cowpeas. The maize variety used is a synthetic, open-pollinated imidazolinone-resistant (IR) variety bred by backcrossing Pioneer hybrid 3245IR into local varieties (Kanampiu et al., 2002). The Pioneer hybrid 3245IR, originally derived from a tissue culture selection (Newhouse et al., 1991), bears a tryptophan 552 to leucine mutation in the acetolactate synthase gene (Bernasconi et al., 1995). Imazapyr was applied at 0.84 mg per seed and pyriithiobac at 0.4 mg per seed, with a commercial fungicide insecticide seed dust. These rates are equivalent to 45 and 21 g per hectare of pure herbicide, respectively, and are more than double the dose that we recommend for use for seed treatments. The rates were chosen to accentuate any potential risk to the intercropped legumes from over-application of herbicide. The field experiment shown (one of three in successive seasons, all with similar results) was conducted during March–August 1999 at the Kenya Agricultural Research Institute (KARI) National Sugar Research Center at Kibos, in western Kenya. The field site was heavily infested with *Striga hermonthica*. The experimental design was a randomized complete block with three replications. The normal spacing between legume and maize is 30 cm.

\$29 for open pollinated or hybrid ALS resistant seed and \$77 for fertilizer per hectare, and the herbicide would be applied by the supplier together with the insecticide+fungicide seed dressing. Currently the local price of maize is \$142/ton. Therefore, farmers can realize a net of more than \$300 per hectare from the yield increase, an approximately tenfold increase on investment in a single 5 month season. The benefits from this technology are high and once adopted, it can ease undernourishment within the grower families, with some extra maize for sale.

Appropriateness of this technology

We therefore propose that this biotechnology is not only appropriate for African agriculture, but as opposed to the developed world, it is more appropriate for African agriculture where hunger is common (Pearce, 2000) and yield losses impinge directly on human caloric intake. In Kenya, 91% of maize produced is directly consumed by humans (CIMMYT, 1999) while maize is mainly livestock feed in the developed countries. The single dominant gene can be easily backcrossed into local varieties by conventional

breeding methods, as we have done from the initial USA corn-belt material.

Other appropriate technologies for *Striga* and for maize cultivation

Bt genes

More than 50% of the maize area in developing countries faces serious problems of insect infestation. In Kenya alone, farmers estimate crop losses due to stem borers at 15% of their ultimate harvest, in a country where many people live on less than US \$1 a day. Infestations of these pests can decimate individual maize fields – depriving a rural family of vital income and a year's supply of their main food source.

Various strains of commonly found *Bacillus thuringiensis* (Bt) produce protein crystals containing one or more δ -endotoxins during sporulation. The δ -endotoxins bind to specific receptors in the insect mid-gut after ingestion, leading to insect death. Each Bt protein is active in only one or a few insect species, making them far more selective than most chemical insecticides. Bt sprays have been used for many years to control several insect pests.

The genes coding for various Bt proteins were among the first to be introduced into agriculturally important cotton and maize. North American farmers planted nearly 8 million hectares of transgenic Bt crops in 1999. The area is expected to increase globally, as other countries begin to approve these crops. Controversial concerns over Bt crops range from a general dislike of genetic engineering, to issues related to insect resistance management and increased profits to the companies that commercialize the crops. The concerns surrounding the release of Bt crops have not been realized, and farmers have seen increased yields, with lower insecticide use, which is needed more in many developing countries than in the developed ones.

One of the Bt genes that have been introduced into maize and commercially released in the USA, *cryIab*, is effective against some of the African insect pests, as are some proteins produced by other *cry* genes (CIMMYT, unpublished data). These natural insecticide genes could be introduced into local open-pollinated as well as in hybrid maize varieties. Farmers prefer their inputs being in the form they are most used to – the seed, and not insecticides. Many farmers are currently not using insecticides due to costs, and thus would immediately see their yields increase. Many others who now can afford limited amounts of chemicals would see increased yields and incomes, with perhaps most important, decreased health risks due to the decreased pesticide use. These benefits must clearly outweigh the slight risks that any new technology brings.

Transposon transmitted suicide genes for Striga

It has been proposed that it may be possible using genetic engineering to debilitate weeds leading to self control, for *Striga* (Gressel and Levy, 2000) and for other cross-pollinated weeds where there are no closely-related crop, nor wild or weedy species (Gressel, 2002). This concept is an adaptation of a proposal for controlling insects with the TAC-TIC model: “Transposons with Armed Cassettes for Targeted Insect Control”. The insects are transformed with assisted-suicide (inducible) genes (*'kev'* (Kevorkian) genes), which if activated, debilitate the host (Pfeifer and Grigliatti, 1996; Grigliatti et al., 2001).

Adapting this would entail developing transgenic *Striga* with high copy number transposons carrying *kev* genes, i.e. deleterious transposons (DTs), which will quickly spread the gene to field populations because *S. hermonthica* is an obligate outcrosser

requiring exogenous pollination (Aigbokhan et al., 1998). The antisense constructs for *Striga* could be introduced within the Ac transposon. The recently mapped gene for haustorial recognition and attachment (Stranger et al., 1999) could be included in a *kev* construct (in the antisense direction or for suppressive overexpression); the ideal inducer would be one emanating from maize roots (Gressel and Levy, 2000).

Many targets can be considered where the genes are known, e.g. in primary biosynthesis pathways that are targets for herbicides (Gressel and Levy, 2000). Plants expressing such an antisense gene will react as if treated by a herbicide when the *kev* inducer is activated. Tandem constructs of more than one gene could add to the versatility of the approach; e.g. the addition of genes abolishing secondary dormancy might assist transposon spread as well as facilitate a more rapid depletion of the seed bank (Gressel and Levy, 2000).

Biocontrol

Various mycoherbicidal organisms (Abbasher et al., 1995; Marley et al., 1999; Ciotola et al., 2000), bacteria (Berner et al., 1999; Miche et al., 2000) and arthropods (Smith et al., 1993; Pronier et al., 1998) that attack *Striga* might serve as biocontrol agents, protecting crops from damage. None have yet fully proven themselves in the field, and there are good scientific/ecological reasons to doubt that a single organism can be cost-effectively applied to provide the level of control needed in field crop situations. The organisms might have the needed effectiveness if they are transgenically-enhanced with virulence genes, as has been generally described in Vurro et al. (2001), although failsafe mechanisms must be considered to prevent off-target movement or gene introgression (Gressel, 2001). Transgenes encoding hormone overproduction conferring hypervirulence have been successfully introduced into fungi controlling a related parasite (Cohen et al., 2002).

Concluding remarks

We advise that extreme caution be used in accepting designations of what is appropriate to African cropping systems from those with a non-scientific agenda, and from those who have little awareness of the needs or the agricultural ecosystems in question. Conversely, crop protection solutions rarely last forever in agriculture; pest organisms typically evolve mechanisms

to circumvent all control tactics, whether chemical or otherwise. One should expect that *Striga* will evolve resistance to herbicides as well as to rotational, trap, or catch crops, or to biocontrol agents. The best one can do is to devise strategies to delay the inevitable. In the case of seed treatment with herbicides, it has been proposed that farmers should rogue the five resistant plants per hectare that models suggest will appear each year, to extend the life of this technology (Gressel et al., 1996). If the seed treatment technology is integrated with hand rouging of any *Striga* that appears, and/or crop rotation, the duration of utility of all the techniques should be enhanced.

Acknowledgements

The authors acknowledge the participation of maize breeder Dr Kevin Pixley; CIMMYT-Zimbabwe for his contribution in developing the maize material used in this study, the staff at Chitedze Research Station, Malawi, Kenya Sugar Research Foundation and Alupe sub-center for efficient management of the fieldwork. This research is supported in part by the Kenya Agricultural Research Institute, International Maize and Wheat Improvement Center (CIMMYT) and the Rockefeller Foundation *Striga* program. J. Gressel is the Gilbert de Botton Professor of Plant Sciences.

References

- Abayo GO, Ransom JK, Gressel J & Odhiambo GD (1996) *Striga hermonthica* control with acetolactate synthase inhibiting herbicides seed-dressed to corn with target site resistance. In: Moreno MT, Cubero JI, Berner DK, Joel DM, Musselman LJ & Parker C (eds) *Advances in Parasitic Weed Research* (pp 762–768). Junta de Andalucia, Cordoba, Spain
- Abayo GO, English T, Eplee RE, Kanampiu FK, Ransom JK & Gressel J (1998) Control of parasitic witchweeds (*Striga* spp.) on corn (*Zea mays*) resistant to acetolactate synthase inhibitors. *Weed Sci.* 46: 459–466
- Abbasher AA, Kroschel J & Sauerborn J (1995) Microorganisms of *Striga hermonthica* in northern Ghana with potential as biocontrol agents. *Biocon. Sci. Tech.* 5: 157–161
- Aigbokhan EI, Berner DK & Musselman LJ (1998) Reproductive ability of hybrids of *Striga aspera* and *Striga hermonthica*. *Phytopathology* 88: 563–567
- Bernasconi P, Woodworth AR, Rosen BA, Subramanian MV & Siehl DL (1995) A naturally occurring point mutation confers broad range tolerance to herbicides that target acetolactate synthase. *J. Biol. Chem.* 270: 17381–17385
- Berner DK, Kling JG & Singh BB (1995) *Striga* research and control: A perspective from Africa. *Plant Dis.* 79: 652–660
- Berner DK, Ikie FO & Green JM (1997) ALS-inhibiting herbicide seed treatments control *Striga hermonthica* in ALS-modified corn (*Zea mays*). *Weed Technol.* 11: 704–707
- Berner DK, Schaad NW & Volksch B (1999) Use of ethylene-producing bacteria for stimulation of *Striga* spp. seed germination. *Biol. Control* 15: 274–282
- Carsky RJ, Berner DK, Oyewole BD, Dashiell K & Schulz S (2000) Reduction of *Striga hermonthica* parasitism on maize using soybean rotation. *Intl. J. Pest Manage.* 46: 115–120
- CIMMYT (1999) 1997/1998 World Maize Facts and Trends; Maize Production in Drought-Stressed Environments: Technical Options and Research Resource Allocation. CIMMYT, Mexico, DF
- Ciotola M, DiTommaso A & Watson AK (2000) Chlamydospore production, inoculation methods and pathogenicity of *Fusarium oxysporum* M12-4A, a biocontrol for *Striga hermonthica*. *Biocont. Sci. Tech.* 10: 129–145
- Cohen BA, Amsellem Z, Maor R, Sharon A & Gressel J (2002) Transgenically-enhanced expression of indole-3-acetic acid (IAA) confers hypervirulence to plant pathogens. *Phytopathology* (in press).
- Combari A, Pineau R & Schiavon M (1990) Influence du degre de decomposition de produits organique sur la germination de graines de *Striga hermonthica*. *Weed Res.* 30: 29–34
- FAO (2001) <http://apps.fao.org/page/collections/subset=agriculture>
- Gacheru E & Rao MR (2001) Managing *Striga* infestation on maize using organic and inorganic nutrient sources in western Kenya. *Intl. J. Pest Manage.* 47: 233–239
- Gagne JA, Fischer JE, Sharma RK, Traul KA, Diehl SJ, Hess FG & Harris JE (1991) Toxicology of the imidazolinone herbicides. In: Shaner DL & O'Connor SL (eds) *The Imidazolinone Herbicides* (pp 179–182). CRC Press, Boca Raton
- Gressel J (2001) Potential failsafe mechanisms against the spread and introgression of transgenic hypervirulent biocontrol fungi. *Trends Biotech.* 19: 149–154
- Gressel J (2002) *Molecular Biology of Weed Control*. Taylor and Francis, London
- Gressel J & Levy A (2000) Giving *Striga hermonthica* the DT's. In: Haussmann BIG, Hess DE, Koyama ML, Grivet L, Rattunde HFW & Geiger HH (eds) *Breeding for Striga Resistance in Cereals* (pp 207–224). Margraf Verlag, Weikersheim
- Gressel J, Segel LE & Ransom JK (1996) Managing the delay of evolution of herbicide resistance in parasitic weed. *Intl. J. Pest Manage.* 42: 113–129
- Grigliatti TA, Pfeifer TA & Meister GA (2001) TAC-TICS: transposon-based insect control systems. In: Vurro M, Gressel J, Butts T, Harman G, Pilgeram A, St.-Leger R & Nuss D (eds) *Enhancing Biocontrol Agents and Handling Risks* IOS Press (pp 201–216), Amsterdam
- Gurney AL, Press MC & Ransom JK (1995) The parasitic angiosperm *Striga hermonthica* can reduce photosynthesis of its sorghum and maize hosts in the field. *J. Exp. Bot.* 46: 1817–1823
- Joel DM, Kleifeld Y, Losner-Goshen D, Herzlinger G & Gressel J (1995) Transgenic crops against parasites. *Nature* 374: 220–221
- Kanampiu FK, Ransom JK & Gressel J (2001) Imazapyr seed dressings for *Striga* control on acetolactate synthase target-site resistant maize. *Crop Protect.* 20: 885–895
- Kanampiu FK, Ransom JK, Friesen D & Gressel J (2002) Imazapyr and pyriithiobac movement in soil and from maize seed coats controls *Striga* while allowing legume intercropping. *Crop Protect.* (accepted)
- Khan ZR, Pickett JA, van den Berg J, Wadhams LJ & Woodcock CM (2000) Exploiting chemical ecology and species diversity: Stem borer and *Striga* control for maize and sorghum in Africa. *Pest Manage. Sci.* 56: 957–962

- Marley PS, Ahmed SM, Shebayan JAY & Lagoke STO (1999) Isolation of *Fusarium oxysporum* with potential for biocontrol of the witchweed (*Striga hermonthica*) in the Nigerian savanna. *Biocont. Sci. Tech.* 9: 159–163
- McCown RL, Keating BA, Probert ME & Jones RK (1992) Strategies for sustainable crop production in semi-arid Africa. *Outlook Agric.* 21: 21–31
- Miche L, Bouillant ML, Rohr R, Salle G & Bally R (2000) Physiological and cytological studies on the inhibition of *Striga* seed germination by the plant growth-promoting bacterium *Azospirillum brasilense*. *Eur. J. Plant Path.* 106: 347–351
- Mumera LM & Below FE (1993) Crop ecology, production and management. *Crop Sci.* 33: 758–763
- Newhouse KE, Singh B, Shaner D & Stidham m (1991) Mutations in corn (*Zea mays* L.) conferring resistance to imidazolinone herbicides. *Theor. Appl. Genet.* 83: 65–70
- Oswald A & Ransom JK (2001) *Striga* control and improved farm productivity using crop rotation. *Crop Protect* 20: 113–120
- Pearce F (2000) Feeding Africa – interview with Florence Wambugu. *New Sci.* 166: 40–49
- Pfeifer TA & Grigliatti TA (1996) Future perspectives on insect pest management: Engineering the pest. *J. Invert. Pathol.* 67: 109–119
- Pingali PL & Pandey S (2001) Meeting world maize needs: technological opportunities and priorities for the public sector. In: Pingali PL (ed) CIMMYT 1999–2000 World Maize Facts and Trends. Meeting World Maize Needs: Technological Opportunities and Priorities for the Public Sector (pp. 1–20). Mexico, D.F.: CIMMYT.
- Press MC & Graves JD (1995) *Parasitic Plants*. Chapman & Hall, London (p 292)
- Pronier I, Pare J, Vincent C & Salle G (1998) Impact of *Smicronyx* spp. (Coleoptera: Curculionidae) on fruit development of the parasitic weed *Striga hermonthica* (Scrophulariaceae): Histological study and prospects for biological control. *Acta Biol. Cracov. Ser. Bot.* 40: 9–13
- Ransom JK (2000) Long term approaches for the control of *Striga* in cereals: Field management. *Crop Prot.* 19: 759–763
- Smith MC, Holt JS & Webb M (1993) Population model of the parasitic weed *Striga hermonthica* (Scrophulariaceae) to investigate the potential of *Smicronyx umbrinus* (Coleoptera: Curculionidae) for biological control in Mali. *Crop Prot.* 12: 473–476
- Stranger A, Corbett JM, Dunn MJ, Totty NF, Sterling A & Bolwell GP (1999) Identification of developmentally-specific markers in germinating and haustorial stages of *Striga hermonthica* (Del.) Benth. seedlings. *J. Exp. Bot.* 50: 269–274
- Tomlin C (1994) *The Pesticide Manual*, 10th edn. British Crop Protection Council, Farnham, UK
- Vurro M, Gressel J, Butts T, Harman G, Pilgeram A, St.-Leger R & Nuss D (eds.) (2001) *Enhancing Biocontrol Agents and Handling Risks*. IOS Press, Amsterdam (p 295)