Study of inheritance of fertility restoration in pigeonpea lines derived from *Cajanus cajanifolius*

S. L. SAWARGAONKAR^{1,2,3}, I. A. MADRAP² and K. B. SAXENA¹

¹International Crops Research Institute for the Semi-Arid Tropics, Patancheru 502 324, Andhra Pradesh, India; ²Marathwada Krishi Vidyapeeth, Parbhani 431 401, Maharashtra, India; ³Corresponding author, Email: sawargaonkar_sl@rediffmail.com *With 1 table*

Received August 23, 2011/Accepted December 7, 2011 Communicated by W. Link

Abstract

Jant Breeding

In this study, single crosses involving four CMS lines and three known fertility restorers were studied to determine the genetics of fertility restoration in pigeonpea (Cajanus cajan L. Millsp.). The F1 plants of all the four hybrids were selfed to produce F2 seed and simultaneously crossed to their corresponding A lines to produce BC1F1 seeds during 2007. The parents, F₁, F₂ and BC₁F₁ populations were planted in 2008 rainy season at the Marathwada Krishi Vidyapeeth, Parbhani, India. The interaction of dominant nuclear genes of ICP 2766 with ICPA 2092 produced 100% fertile F1 plant and showed complete dominance for fertility restoration. The restorer ICP 2766 when crossed with ICPA 2092 showed monogenic inheritance (3 : 1), while ICP 2766 crossed with ICPA 2043 revealed digenic inheritance (15:1) of fertility restoration. In a cross between ICP 2766 and ICPA 2043, dominant gene of fertility restoration at either of two loci masked the expression of male-sterile recessive alleles at two loci in such a way that it modified normal dihybrid ratio into 15: 1 ratio and produced duplicate gene interaction.

Key words: *Cajanus cajanifolius* — inheritance — male sterility — male fertility — genetics

Cytoplasmic nuclear male sterility (CMS) is maternally inherited and associated with specific (mitochondrial) genes without otherwise affecting the plant (Budar and Pelletier 2001). The fertility restorer (Rf or Fr) genes in the nucleus suppress the male-sterile phenotype and allow commercial exploitation of the CMS system for the production of hybrid seeds. In addition, the CMS-Rf system provides an excellent model for the study of nuclear-mitochondrial interaction in multicellular organisms. Pigeonpea is the only legume crop where commercial CMS-based hybrids are now available (Saxena and Nadarajan 2010). CMS-derived hybrids were coming from A2 (Cajanus scarbaeoides, Tikka et al. 1997) and A4 (Cajanus cajanifolius, Saxena et al. 2005) cytoplasm and that in the present paper only the A4 restoration genetics is studied. Saxena et al. (2005) crossed ICPW 29, an accession of C. cajanifolius, a wild relative of pigeonpea, as female parent with pigeonpea line ICPL 28. According to De (1974), C. cajanifolius resembles cultivated types in most morphological traits. The male-sterile lines derived from A4 cytoplasm are the best among those identified so far and found to be highly stable male-sterile lines across environments and years (Dalvi et al. 2008a, Saxena 2008, Sawargaonkar 2010) and never showed any morphological deformity. In the present study, the F_1 and F_2 generations were developed to determine the nature of gene action in the F_1 generation and the segregation pattern in F₂ generation in pigeonpea. In order to take advantage of this CMS hybrid technology, it is essential to breed higher yielding hybrids based on diverse genetic backgrounds. To achieve this, breeding of promising hybrid parents and knowledge of the inheritance of fertility restoration are essential.

Materials and Methods

In this study, single crosses involving four CMS lines (A4 cytoplasm) and three known fertility restorers (Table 1) were studied to determine the genetics of fertility restoration. The F1 plants of all the four hybrids were selfed to produce F2 seeds and simultaneously crossed to their corresponding A lines to produce BC1F1 seeds during 2007. The parents, F₁, F₂ and BC₁F₁ populations were planted in 2008 rainy season at Marathwada Krishi Vidyapeeth, Parbhani, India. For each F₂ population, 300-500 seeds and for BC₁F₁ population 125-175 seeds were sown. To determine the variation in pollen fertility in each generation, five fully developed floral buds were collected randomly from each plant and the anthers were squashed in 2% aceto-carmine solution. The pollen fertility of each plant was studied in three microscopic fields under light microscope. The fully stained pollen grains were identified as fertile, while the empty or partially stained pollen grains were considered as sterile. The goodness of fit to different expected ratios in F2 and test-cross generations was assessed using chi-square test.

Results

The results revealed that all the F_1 plants in the four crosses were male-fertile, indicating a dominant nature of the fertilityrestoring genes. As expected, the F_2 and BC₁F₁ populations of these crosses segregated for male sterility and fertility (Table 1). In cross ICPA 2092 × ICP 2766, of 458 F_2 plants grown, 350 were fertile and 108 were male-sterile. This segregation fit the expected ratio of 3 : 1 (fertile/sterile, P = 0.95). In BC₁F₁ generation, 64 plants were male-fertile and 59 plants were male-sterile, and it showed a good fit to 1 : 1 (fertile/sterile, P = 0.70) ratio. These observations suggested that the fertility restoration in cross ICPA 2092 × ICP 2766 was controlled by a single dominant gene. Similarly in cross ICPA 2048 × ICP 3477, the F_2 and BC₁F₁ data (Table 1) showed that the fertility restoration was controlled by a single dominant gene.

In cross ICPA 2047 × ICP 3513, the segregation in F_2 generation revealed that of 424 plants, 393 were male-fertile and only 31 were devoid of pollen grains. This segregation fit the expected dihybrid ratio of 15: 1 (fertile/sterile, P = 0.87).

| Crosses | Generation | Number of plants | | | | |
|----------------------|----------------------|------------------|---------|---------|-------|------------------------|
| | | Total | Fertile | Sterile | Ratio | Chi-square probability |
| ICPA 2092 × ICP 2766 | F_1 | 5 | 5 | 0 | _ | _ |
| | F_2 | 458 | 350 | 108 | 3:1 | 0.95 |
| | BC_1F_1 | 123 | 64 | 59 | 1:1 | 0.7 |
| ICPA 2043 × ICP 2766 | F_1 | 7 | 7 | 0 | _ | _ |
| | F_2 | 315 | 292 | 23 | 15:1 | 0.87 |
| | $\overline{BC_1F_1}$ | 111 | 84 | 27 | 3:1 | 0.57 |
| ICPA 2048 × ICP 3477 | F_1 | 9 | 9 | 0 | - | _ |
| | F_2 | 417 | 318 | 99 | 3:1 | 0.96 |
| | $\overline{BC_1F_1}$ | 114 | 53 | 61 | 1:1 | 0.51 |
| ICPA 2047 × ICP 3513 | F_1 | 17 | 17 | 0 | _ | _ |
| | F_2 | 424 | 393 | 31 | 15:1 | 0.87 |
| | BC_1F_1 | 153 | 110 | 43 | 3:1 | 0.63 |

Table 1: Segregation ratios for male-fertile and sterile plants in F_1 , F_2 and BC_1F_1 generations of crosses involving A4 CMS lines and fertility restorers in pigeonpea

In BC₁F₁ generation, 110 plants were male-fertile and 43 were male-sterile, which showed a good fit for 3 : 1 (fertile/sterile, P = 0.63) ratio. Similarly in cross ICPA 2043 × ICP 2766, of 315 plants studied, 292 were male-fertile and 23 male-sterile, exhibiting a good fit to a 15: 1 (fertile/sterile, P = 0.87) ratio. In BC₁F₁ generation, of 111 plants grown, 84 were male-fertile and 27 were male-sterile. This segregation fit the expected ratio of 3 : 1 (fertile/sterile, P = 0.57). The segregation patterns observed in these two crosses suggested the presence of two duplicate dominant genes in controlling the pollen fertility of CMS lines ICPA 2047 and ICPA 2043.

Discussion

The genetics of fertility restoration is important for the transfer of restorer genes from one genotype to another. In the present study, it was observed that restorer ICP 2766 when crossed with ICPA 2092 showed monogenic inheritance (3 : 1), while the cross with another ICPA 2043 revealed digenic inheritance of fertility restoration. Dalvi et al. (2008b) studied the inheritance of fertility restoration in three diverse early maturing CMS lines of pigeonpea and reported that fertility restoration in three pigeonpea crosses was controlled by one dominant gene; while in two crosses two duplicate dominant genes controlled fertility restoration. Nadarajan et al. (2008) reported variable restoration patterns among a common set of male parents within a single cytoplasmic source of pigeonpea. In the present study, fertility restorer line ICP 2766 when crossed with two different A lines produced different results as far as their fertility restoration was concerned. In cross ICPA $2092 \times ICP$ 2766, the fertility restoration was controlled by a single dominant gene, while in cross ICPA $2043 \times ICP$ 2766 a digenic duplicate dominance inheritance of fertility restoration was observed. The differential expression of fertility-restoring genes of ICP 2766 can be attributed to different genetic backgrounds of the F1 plants, arising from female parents of different genetic constitution. Such a variable restoration among cytoplasmic sources with the same set of male parents in pigeonpea had been reported (Saxena and Kumar 2003, Saxena et al. 2005). Saxena et al. (2010) revealed that the fertility restoration in A4 CMS lines of pigeonpea was controlled by either one or two fertility-restoring genes. Bai and Gai (2005) observed that in soybean fertility restoration was controlled by monogenic to digenic gene in CMS lines of NJCMS2A. Monofactorial inheritance associated with fertility restoration in red pepper (Capsicum annum L.) was reported by Gulvas et al. (2006). Schertz (1994) reported that fertility restoration in sorghum (Sorghum bicolor (L.) Moench) was dependent on interaction between cytoplasms and nuclear genes and was controlled by a single dominant gene in some combinations but by two or more dominant genes in other combinations. This happened when the same nuclear genes interacted with different cytoplasms. The differential reactions of CMS lines with the same cytoplasm in test crosses may be caused by the action of recessive nuclear genes of the recurrent male parents that were accumulated during backcrossing, which may induce changes in cytoplasmic genes controlling male sterility (Elkonin et al. 1998). Differences observed in segregation patterns also could be due to the presence of some modifier genes that influence the process of penetrance and expressivity of the fertility-restoring genes (Hossain et al. 2010).

In the present study, the inheritance of male-fertility restoration in CMS lines of pigeonpea was cross-specific and influenced by the nuclear background of male-sterile and fertility-restoring lines. In crosses ICPA $2048 \times ICP$ 3477 and ICPA $2092 \times ICP$ 2766, it was controlled by a single dominant gene; while in crosses ICPA $2043 \times ICP$ 2766 and ICPA $2047 \times ICP$ 3513, the fertility restoration was determined by two duplicate dominant genes. The differential behaviour of one fertility restorer line (ICP 2766) in two different crosses was attributed to the interactions of different nuclear genes of the two female parents.

Acknowledgement

The authors acknowledge Bill and Melinda Gates Foundation (through Tropicsl Legume II Project) for providing financial support to the first author as a PhD student.

References

- Bai, Y. N., and J. Y. Gai, 2005: Development of a new cytoplasmicnuclear male-sterility line of soybean and inheritance of its malefertility restorability. Plant Breed. 125, 85–88.
- Budar, F., and G. Pelletier, 2001: Male sterility in plants; occurrence, determinism, significance and use. Life Sci. 324, 543—550.
- Dalvi, V. A., K. B. Saxena, I. A. Madrap, and V. K. Ravikoti, 2008a: Cytogenetic studies in A₄ cytoplasmic–nuclear male-sterility system of pigeonpea. J. Hered. **99**, 667–670.
- Dalvi, V. A., K. B. Saxena, and I. A. Madrap, 2008b: Fertility restoration in cytoplasmic-nuclear male-sterile lines derived from three wild relatives of pigeonpea. J. Hered. 99, 671–673.

- De, D. N., 1974: Pigeonpea. In: J. Hutchinson, (ed.), Evolutionary Studies in World Crops, Diversity and Change in the Indian Subcontinent, 79–87. Cambridge University Press, London.
- Elkonin, I. A., V. V. Kozhemyakin, and A. G. Ishin, 1998: Nuclearcytoplasmic interactions in restoration of male fertility in the 9E and A₄ CMS-inducing cytoplasm of sorghum. Theor. Appl. Genet. 97, 626–632.
- Gulyas, G., K. Pakozdi, J. Lee, and Y. Hirata, 2006: Analysis of fertility restoration by using cytoplasmic male-sterile red pepper [*Capsicum annuum* L.] lines. Breed. Sci. 56, 331–334.
- Hossain, M. D., A. K. Singh, and F. Zamanb, 2010: Genetics of fertility restoration of 'WA'-based cytoplasmic male-sterility system in rice (*Oryza sativa*) using indica/japonica derivative restorers. Sci. Asia **36**, 94–99.
- Nadarajan, N., S. Ganesh, and K. I. Petchiammal, 2008: Fertility restoration studies in short duration redgram [*Cajanus cajan* (L.) Millsp.] hybrids involving CGMS system. Madras Agric. J. 95, 320—327.
- Sawargaonkar, S. L., 2010: Study of Heterosis, Combining Ability, Stability, and Quality Parameters in CGMS-Based Pigeonpea Hybrids. PhD Agri. Thesis, Marathwada Krishi Vidyapeeth, Parabhani, Maharashtra, 151.
- Saxena, K. B., 2008: Genetic improvement of pigeonpea a review. Trop. Plant Biol. 1, 159–178.

- Saxena, K. B., and R. V. Kumar, 2003: Development of a cytoplasmic-nuclear male-sterility system in pigeonpea using [*C. scarabae*oides (L.) Thours.]. Indian J. Genet. 63, 225–229.
- Saxena, K. B., and N. Nadarajan, 2010: Prospects of pigeonpea hybrids in Indian agriculture. J. Plant Breed. 1, 107–1117.
- Saxena, K. B., R. V. Kumar, N. Srivastava, and B. Shiying, 2005: A cytoplasmic–genic male-sterility system derived from a cross between *Cajanus cajanifolius* and *Cajanus cajan*. Euphytica **145**, 291–296.
- Saxena, K. B., R. Sultana, R. K. Saxena, R. V. Kumar, J. S. Sandhu, A. Rathore, and R. K. Varshney, 2010: Genetics of fertility restoration in A₄ based diverse maturing hybrids in pigeonpea [*Cajanus cajan* (L.) Millsp.]. Crop Sci. **51**, 1–5.
- Schertz, K. F., 1994: Male sterility in sorghum: its characteristics and importance. In: Use of molecular markers in sorghum and pearlmillet breeding for developing countries (J. R. Witcombe, and R. R. Duncan, eds). Proceedings of the International Conference on Genetic Improvement, Overseas Development Administration (ODA), 35–37. Plant Sci. Res. Conf., 29 March–1 April 1993, Norwich, UK.
- Tikka, S. B. S., L. D. Parmer, and R. M. Chauhan, 1997: First record of cytoplasmic genetic male-sterility in pigeonpea [*Cajanus cajan* (L.) Millsp.] through wide hybridization. Gujarat Agric. Univ. Res. J. 22, 160—162.