

CARBON DIOXIDE CONCENTRATION IN SOIL EFFECTS ON NITRIFICATION, DENITRIFICATION AND ASSOCIATED NITROUS OXIDE PRODUCTION

D R KEENEY, K L SAHRAWAT* and SUSAN S ADAMS
Department of Soil Science, University of Wisconsin, Madison, WI 53706, U S A

(Accepted 30 December 1984)

Summary—Experiments were conducted to study the effects of a range of CO₂ concentrations (ambient to 100%) on nitrification, denitrification and associated nitrous oxide (N₂O) production in a silt loam soil. It was found that increase in CO₂ concentration from 0.3 to 100% CO₂ increasingly retarded the rate of nitrification. No nitrification occurred at 100% CO₂. Nitrous oxide production associated with nitrification increased as CO₂ increased from 0.3 to 2.6% and tended to be greater as CO₂ concentration increased to 73%. At 100% CO₂, no N₂O was produced during 7 days at 25°C.

Carbon dioxide did not affect N₂O production or reduction in a saturated NO₃⁻-amended soil or the rate of N₂O reduction in anaerobic environments.

INTRODUCTION

Carbon dioxide, a product of both aerobic and anaerobic metabolism, can have a direct influence on microbial growth. It is usually the sole C source for chemoautotrophs and photoautotrophs and apparently is also required, in addition to organic C, for most heterotrophs (Alexander, 1977). While CO₂ is rarely limiting in soil due to its continual evolution from plant roots and during organic matter decomposition, excess CO₂ may be toxic to some soil fungi (Alexander, 1977). High CO₂ in the soil atmosphere will also affect solution pH, nutrient availability, aeration status (Bohn *et al.*, 1979) and root growth (Buyanovsky and Wagner, 1983).

However, little attention has been paid to possible effects of CO₂ on soil microbial activities. Clark (1968) found more rapid nitrification in the range 0.6–2.9% CO₂ than at higher and lower CO₂ concentrations (see also Singh and Kanehiro, 1972). Yeomans and Beauchamp (1978) noted that the rate of reduction of N₂O in the absence or presence of C₂H₂ was delayed when the CO₂ produced during incubation was removed by sorption in an alkali trap.

Measurements of CO₂ in atmospheres of field soils and animal feedlots have given values ranging from ambient (0.3% CO₂) to as high as 100% (Enoch and Dasberg, 1971; Wood and Greenwood, 1971; Stevens and Cornforth, 1974; Focht *et al.*, 1979; Buyanovsky and Wagner, 1983; Norstadt and Porter, 1984), although values below 20% CO₂ are more commonly observed. However, these are bulk soil atmospheres and microsite CO₂ concentrations probably will be higher than those in large voids.

Our objective was to evaluate the effects of different CO₂ concentrations on nitrification, denitrification and N₂O production and reduction associated with a silt loam soil. This study was prompted by the previously cited findings of Yeomans and Beauchamp

(1978). A further effect of CO₂ was suggested by Sherlock and Goh (1983), who stated that CO₂ could be involved in creation of anaerobic microsites, leading to increased N₂O evolution from soils.

MATERIALS AND METHODS

Soil

A bulk sample of the Ap (0–20 cm) horizon of a Plano silt loam (Typic Argiudoll) was obtained from a site previously cropped to maize (*Zea mays* L.) and had a pH (soil water ratio of 1:2) of 7.0 and 14 mg organic C g⁻¹ (determined as described in Harris and Adams, 1979). Ammonium-N and NO₃⁻-N (determined by extraction with 2 M KCl and steam distillation, Keeney and Nelson, 1982) were 7 and 10 mg kg⁻¹, respectively. Before use in the incubations, the sample was air-dried and crushed (< 2 mm).

Gas analyses

Flask atmospheres were analyzed for N₂O with a Perkin-Elmer Sigma 3 GC equipped with a high temperature electron capture detector (Goodroad and Keeney, 1984). CO₂ in the soil atmosphere in the flasks was analyzed with a Tracor 150G gas chromatograph fitted with dual ultrasonic detectors (Blackmer and Bremner, 1977).

Incubation procedures

In all experiments, 10 g dry wt of soil were added to a 125 ml Erlenmeyer flask fitted with a rubber septum and the flasks were incubated at 25°C. Water content was adjusted to either 2.5 ml 10 g⁻¹ (–33 kPa water tension) or 10 ml 10 g⁻¹. In experiments involving varying concentrations, the appropriate volume of flask air was removed by syringe and replaced with high purity CO₂ or N₂O. Experiments involving the addition of inorganic N used (NH₄)₂SO₄ or KNO₃ as amendments. All experiments were made in triplicate.

Unless otherwise stated, flask atmospheres were sampled using a syringe and analyzed after 4 and 7

*Present address ICRISAT, Patancheru P O, Andhra Pradesh 502 324, India

Table 1. Nitrogen transformations in an aerobic soil under varying concentrations

Treatment			CO ₂ 7 days (%)	N ₂ O		Inorganic N		Sum
CO ₂ (%)	NH ₄ ⁺ (μg N g ⁻¹)	NO ₃ ⁻		4 days (ng N g ⁻¹)	7 days	NH ₄ ⁺ (μg N g ⁻¹)	NO ₃ ⁻	
Air	0	0	3.3	96 (1)	117 (2)	2 (0)	44 (2)	46 (1)
22	0	0	25	65 (12)	83 (14)	16 (0)	29 (1)	45 (1)
50	0	0	49	132 (5)	154 (11)	26 (1)	17 (7)	43 (1)
73	0	0	70	329 (38)	544 (55)	30 (1)	13 (0)	43 (0)
100	0	0	100	0	0	22 (6)	0	22 (6)
Air	0	95	3.1	94 (8)	123 (10)	0	140 (2)	140 (1)
23	0	95	25	112 (15)	168 (23)	14 (1)	127 (2)	141 (1)
50	0	95	47	133 (6)	250 (28)	24 (0)	115 (0)	139 (0)
73	0	95	71	516 (102)	716 (303)	28 (1)	113 (0)	141 (0)
100	0	95	100	62,000 (2740)	46,400 (1350)	20 (0)	26 (1)	46 (1)
Air	90	0	3.6	205 (21)	396 (126)	48 (1)	78 (2)	126 (1)
2.6	90	0	6.2	474 (141)	1120 (208)	65 (2)	69 (3)	134 (2)
5.2	90	0	8.6	389 (90)	972 (88)	66 (1)	70 (2)	136 (1)
11	90	0	14	538 (202)	1616 (637)	67 (2)	61 (0)	128 (1)
23	90	0	26	399 (16)	1111 (104)	80 (4)	50 (1)	130 (1)
50	90	0	49	298 (45)	1034 (152)	104 (3)	24 (1)	128 (2)
73	90	0	71	609 (260)	1270 (562)	121 (4)	13 (5)	134 (4)
100	90	0	100	0	0	113 (1)	0	113 (1)

^aPlano silt loam (10 g) was incubated at 25°C with 2.5 ml of water or with 2.5 ml of a solution containing either 900 μg NO₃⁻-N (as KNO₃) or 950 μg NH₄⁺-N [as (NH₄)₂SO₄]. The soil initially contained 7 and 11 μg NH₄⁺-N and NO₃⁻-N g⁻¹ soil, respectively. Figures in parentheses are standard deviations.

days of incubation, and the flasks were not flushed between measurements. Soil was analyzed for NH₄⁺-N, and (NO₂⁻ + NO₃⁻)-N at 7 days. Nitrite-N was never >0.5 mg kg⁻¹ and usually was below detection limits.

RESULTS

The CO₂ levels in the flasks did not increase markedly during incubation (Table 1). With the exception of the 100% CO₂ treatment, O₂ was still present in all flasks at the end of 7 days. The presence of CO₂ affected both net N₂O and NO₃⁻ production. In the nonamended system, 22% CO₂ lowered the amount of NO₃⁻ produced by *ca.* 45%. This effect continued so that by 73% CO₂ only 2 μg NO₃⁻-N g⁻¹ soil was formed (control, 33 μg NO₃⁻-N g⁻¹; 22%, 18 μg). However, with the exception of the 100% CO₂ treatment, net N mineralization was not greatly affected as evidence by the nearly constant value for

(NH₄⁺ + NO₃⁻)-N. This was true for the control and for the N-amended soils where NH₄⁺-N produced increased and NO₃⁻-N produced decreased as CO₂ concentrations increased.

Nitrous oxide production tended to increase with CO₂ concentration in the unamended and NO₃⁻-amended soils to 50% CO₂, increased dramatically at 73% CO₂ and declined to nil at 100% CO₂ with the nonamended soil. It would appear that at least with the air and 22% CO₂ treatments, the source of N₂O was largely from nitrification (Bremner and Blackmer, 1978) since the ratio of N₂O-N produced to NO₃⁻-N produced (Table 2) was in the range of 3 × 10⁻³ to 7 × 10⁻³. Goodroad and Keeney (1984) found with the Plano at 0.2 m³ H₂O m⁻³ (the same water ratio as used here) the N₂O-N to NO₃⁻-N ratio was 1 × 10⁻³ at 20° to 30°C. The ratio increased at higher water contents and at lower soil pH's.

The N₂O-N to NO₃⁻-N ratio increased at 50 and 73% CO₂, indicating either (a) that more of the N₂O was being released during nitrification; (b) that some denitrification was occurring at microsites due to lower *p*O₂ in the high *p*CO₂ flasks, or (c) chemical reactions with NO₂⁻ (Nelson, 1982) due to the low pH in the CO₂-saturated systems. It is not possible with the data presented here to ascertain which of these mechanisms may be dominating. Soil pH could not be measured in the closed systems, but was generally about 6.4 after incubation. The pH of a pure water-H₂CO₃ system would be 4.3 at 20% and 3.4 at 100% CO₂ (Stumm and Morgan, 1970).

The pattern of N₂O evolution from the NH₄⁺-amended soils was more erratic than it was in the control and NO₃⁻-amended treatments. However, there is little doubt that CO₂, even at concentrations commonly occurring in soil profiles, yet sufficiently low that soil pH or aeration would be little affected (up to 11% CO₂), markedly increases N₂O production during nitrification.

At 100% CO₂, lack of O₂ would prevent nitrification and denitrification would be rapid. N₂O

Table 2. Effect of CO₂ on the ratio of N₂O produced to NO₃⁻ produced

Treatment			N ₂ O-N to NO ₃ ⁻ -N (× 10 ³)
CO ₂ (%)	NH ₄ ⁺ (μg g ⁻¹)	NO ₃ ⁻	
Air	0	0	3.4
23	0	0	4.4
50	0	0	22
73	0	0	181
Air	0	95	3.6
23	0	95	7
50	0	95	25
73	0	95	90
Air	90	0	6
2.6	90	0	19
5.2	90	0	16
11	90	0	32
23	90	0	28
50	90	0	74
73	90	0	423

Table 3. Effect of CO₂ on reduction of N₂O^a

Treatment	N ₂ O		
	Initial	4 days ($\mu\text{g N}_2\text{O-N g}^{-1}$ soil)	7 days
Control	0	0.1	0.2
CO ₂	0	0	0
N ₂ O-air	189 (9)	149 (6)	137 (5)
N ₂ O-air-C ₂ H ₂	190 (3)	190 (3)	156 (15)
N ₂ O-He	190 (2)	0	0
N ₂ O-CO ₂	191 (2)	18 (16)	0
N ₂ O-CO ₂ -C ₂ H ₂	190 (5)	144 (2)	85 (49)
N ₂ O-He-C ₂ H ₂	191 (4)	152 (2)	132 (0)

^aPlano silt loam (10 g) was treated with 10 ml H₂O and with CO₂ (100%) or He (100%) with or without N₂O (ca 1%). Acetone-free C₂H₂ was added at 0.1%. Figures in parentheses are standard deviations.

formed with the nonamended and NH₄⁺-amended treatments was rapidly reduced to N₂. The NO₃⁻ treatment at 100% CO₂ resulted in a large evolution of N₂O with subsequent reduction of a portion of this to N₂.

Under aerobic conditions, N₂O disappeared from the aerobic system although C₂H₂ inhibited this loss for at least 4 days (Table 3). Reduction of N₂O was rapid under CO₂ or He. In the presence of C₂H₂, N₂O reduction was slower but still significant. Loss of effectiveness of C₂H₂ in blocking N₂O reduction has been reported by others (Yeomans and Beauchamp, 1978). There is some indication that high CO₂ may accelerate the loss of N₂O in the presence of C₂H₂, but the effect was not significant. CO₂ also had no effect on N₂O production from a saturated soil (data not reported). An additional experiment (data not reported) to evaluate the effect of prior treatment of soil with 100% CO₂ for 4 days showed that there was no significant effect of this treatment on nitrification, denitrification, or N₂O production rates.

DISCUSSION

These results indicate that CO₂ increase in soils, particularly arable soils containing high amounts of nitrifiable N, could result in increased N₂O production during nitrification. The effect is not large, however, until CO₂ comprises >50% of the soil atmosphere (Tables 1 and 2), a situation not normally encountered in the field. There appears to be little effect of CO₂ concentrations on N₂O reduction or denitrification (Table 3).

Acknowledgements—Research contribution from the Department of Soil Science, University of Wisconsin-Madison and by the National Science Foundation.

REFERENCES

- Alexander M. (1977) *Introduction to Soil Microbiology*, 2nd edn. Wiley, New York.
- Blackmer A. M. and Bremner J. M. (1977) Gas chromatographic analysis of soil atmospheres. *Soil Science Society America Journal* **41**, 908–912.
- Bremner J. M. and Blackmer A. M. (1978) Nitrous oxide: emission from soils during nitrification of fertilizer nitrogen. *Science* **199**, 295–296.
- Bohn H., McNeal B. and O'Conner G. (1979) *Soil Chemistry*. Wiley, New York.
- Buyanovsky G. A. and Wagner G. H. (1983) Annual cycles of carbon dioxide level in soil air. *Soil Science Society America Journal* **47**, 1139–1145.
- Clark F. E. (1968) The growth of bacteria in soil. In *The Ecology of Soil Bacteria* (T. R. G. Gray and D. Parkinson, Eds), pp. 441–457. University of Toronto Press.
- Enoch H. and Dasberg S. C. (1971) The occurrence of high CO₂ concentrations in soil air. *Geoderma* **6**, 17–21.
- Focht D. D., Stolzy L. H. and Meek B. D. (1979) Sequential reduction of nitrate and nitrous oxide under field conditions as brought about by organic amendments and irrigation management. *Soil Biology & Biochemistry* **11**, 37–46.
- Goodroad L. L. and Keeney D. R. (1984) Nitrous oxide production in aerobic soils under varying pH, temperature and water content. *Soil Biology & Biochemistry* **16**, 39–43.
- Harris R. F. and Adams S. S. (1979) Determination of the carbon-bound electron composition of microbial cells and metabolites by dichromate oxidation. *Applied and Environmental Microbiology* **37**, 237–243.
- Keeney D. R. and Nelson D. W. (1982) Inorganic forms of nitrogen. In *Methods of Soil Analysis*, Part 2 (2nd edn). Agronomy **10**, 643–698.
- Nelson D. W. (1982) Gaseous losses of nitrogen other than through denitrification. In *Nitrogen in Agricultural Soils*. Agronomy **22**, 327–363.
- Norstadt F. A. and Porter L. K. (1984) Soil gases and temperature: a beef cattle feedlot compared to alfalfa. *Soil Science Society of America Journal* **48**, 783–789.
- Sherlock R. R. and Goh K. M. (1983) Initial emission of nitrous oxide from sheep urine applied to pasture soil. *Soil Biology & Biochemistry* **15**, 615–617.
- Singh B. R. and Kanehiro Y. (1972) Interaction of CO₂ tension and salt concentration on mineral nitrogen release in an allophanic soil. *Soil Science* **114**, 401–403.
- Stevens R. J. and Cornforth I. S. (1974) The effect of pig slurry applied to a soil surface on the composition of the soil atmosphere. *Journal of Science of Food & Agriculture* **25**, 1263–1272.
- Stumm W. and Morgan J. J. (1970) *Aquatic Chemistry. An Introduction Emphasizing Chemical Equilibria in Natural Waters*. Wiley, New York.
- Wood J. T. and Greenwood D. J. (1971) Distribution of carbon dioxide and oxygen in the gas phase of aerobic soils. *Journal of Soil Science* **22**, 281–288.
- Yeomans J. C. and Beauchamp E. G. (1978) Limited inhibition of nitrous oxide reduction in soil in the presence of acetylene. *Soil Biology & Biochemistry* **10**, 517–519.