

CP. 406

CP. 406

Incorporation of Rust Resistance from Wild *Arachis* Species into the Cultivated Groundnut

A.K. Singh¹, J.P. Moss² and B.G. Rao³

Abstract

*On the basis of genomic relationships in section Arachis in the genus Arachis several cytogenetic manipulations were adopted to aid gene transfer from the diploid wild species ($2n = 20$) into the cultivated tetraploid *A. hypogaea* ($2n = 40$). Triploid hybrids were produced between *A. hypogaea* and the eight diploid rust-resistant species of section Arachis. Chromosome numbers in these hybrids were doubled to produce hexaploids that were fertile and could be backcrossed with *A. hypogaea*. Some triploids did produce a few seeds and seedlings, these progenies had varying chromosome numbers ($2n = 20$ to 60) and produced a considerable range of recombinants. Synthetic autotetraploids and amphidiploids were produced from the diploid species. They were then crossed with *A. hypogaea*. This has bridged the ploidy gap between the diploid wild and the tetraploid cultivated species, and increased meiotic recombinations. Backcrossing the resultant hybrids with *A. hypogaea* with a few intervening selfing generations has produced a large number of *A. hypogaea*-like interspecific derivatives. Screening these derivatives identified segregants incorporating genes from the wild species *A. cardenasii*, *A. batizocoi*, *A. duranensis* and *A. species GKP 10038* that confer resistance to rust.*

Résumé

Incorporation de la résistance à la rouille dans les arachides cultivées à partir des espèces sauvages d'*Arachis* : L'existence de relations génomiques à l'intérieur de la section Arachis du genre Arachis a permis de faire des manipulations cytogénétiques en vue d'un transfert des gènes à partir des espèces sauvages diploïdes ($2n = 20$) en tétraploïde cultivé *A. hypogaea* ($2n = 40$). Des hybrides triploïdes sont obtenus des croisements d'*A. hypogaea* avec les huit espèces résistantes diploïdes de la section Arachis. Les nombres chromosomiques chez ces hybrides sont doublés afin d'obtenir des hexaploïdes féconds permettant le rétrocroisement avec *A. hypogaea*. Certains triploïdes ont en effet produit quelques semences et de plantules, ces descendances possèdent des nombres chromosomiques variables ($2n = 20$ à 60) donnant une large gamme de recombinants. A partir des espèces diploïdes obtenus des autotétraploïdes et des amphidiploïdes de synthèse. Leur croisement avec *A. hypogaea* permet de rapprocher, du point de vue génomique, les diploïdes sauvages et les tétraploïdes cultivés tout en augmentant les recombinaisons méiotiques. Le rétrocroisement des hybrides avec *A. hypogaea* en passant par quelques générations autofécondées produit un grand nombre de dérivés interspécifiques d'*A. hypogaea*. Parmi ces dérivés, on a repéré des ségréants qui comportent des gènes des espèces sauvages dont *A. cardenasii*, *A. batizocoi*, *A. duranensis* et l'espèce GKP 10038 d'*Arachis* responsables de la production de la résistance à la rouille.

Groundnut (*Arachis hypogaea* L.) rust caused by the fungus *Puccinia arachidis* Speg. often results in yield losses of over 50% (Subrahmanyam et al. 1979). The disease can be controlled with fungicides, but

resource-poor groundnut farmers in the semi-arid tropics (SAT) need a groundnut cultivar that has genetic resistance. A wide range of groundnut germplasm, cultivated as well as wild, has been screened

¹ Cytogeneticist ² Principal Cytogeneticist ³ Research Associate Groundnut Improvement Program International Crops Research Institute for the Semi-Arid Tropics Patancheru A.P. 502 324 India

ICRISAT (International Crops Research Institute for the Semi-Arid Tropics) 1987 Groundnut rust disease. Proceedings of a Discussion Group Meeting, 24-28 Sep 1984, ICRISAT Center, India Patancheru A.P. 502 324 India ICRISAT (CP 406)

for resistance to rust and several rust-resistant accessions have been identified (Subrahmanyam et al 1980, 1982, 1983, 1985)

The cultivated species, *A. hypogaea* (2n=40), has been grouped with a number of cross-compatible wild diploid species (2n=20) in the section *Arachis* (Gregory et al 1973). All these diploid species have a high degree of resistance to groundnut rust ranging from immunity (no visible symptoms) to hypersensitivity (a few small necrotic non-sporulating pustules on leaflets). These are good sources of rust resistance for use in genetic improvement of *A. hypogaea*.

Ploidy differences between wild and cultivated species in section *Arachis* are barriers to genetic introgression. A basic understanding of genomic structure and interrelationships between the species has helped in the selection of procedures that can overcome these barriers. The present paper reports the progress of work at ICRISAT on the transfer of genes conferring rust resistance from a few diploid

wild species into the cultivated tetraploid species using different genomic and ploidy manipulations

Materials and Methods

The sources and identities of the eight diploid wild species (2n=20) and the cultivars belonging to two subspecies of *A. hypogaea* (2n=40), *A. hypogaea* subspecies *hypogaea* Krap et Rig and *A. hypogaea* subspecies *fastigiata* Waldron, all of section *Arachis*, are given in Table 1. Hybridization between the diploid species and cultivars of *A. hypogaea* was done in a greenhouse at ICRISAT Center. The techniques followed for hybridization, cytological analysis, polyploidy induction, and screening against rust in the field and under laboratory conditions have been described earlier (Subrahmanyam et al 1980, Singh et al 1983).

Table 1 Sources and taxonomic status of parents used in transfer of rust resistance from wild species.

Species/cultivar	Collector ¹	Coll No	ICG No ²	Origin
Wild				
<i>A. villosa</i> (Benth)	-	-	8144	Uruguay
<i>A. correntina</i> (Burk.) Krap et Greg	GKP	9530	8140	Argentina
<i>A. chacoense</i> Krap et Greg	GKP	10602	4983	Argentina
<i>A. cardenasii</i> Krap et Greg	GKP	10017	8216	Bolivia
<i>Arachis</i> species	HLK	410	8126	Brazil
<i>Arachis</i> species	GKP	10038	8139	Argentina
<i>A. duranensis</i> Krap et Greg	K	7988	8123	Argentina
<i>A. batizocoi</i> Krap et Greg	K	9484	8124	Argentina
Cultivated				
<i>A. hypogaea</i> L. ssp <i>fastigiata</i> Waldron var <i>fastigiata</i> (valencia)		-	2738	India
1 Gangapuri				
<i>A. hypogaea</i> L. ssp <i>fastigiata</i> Waldron var <i>vulgaris</i> (spanish)				
2 99-5		-	1472	Unknown
3 Chico		-	476	USA
4 Tifspan		-	3497	USA
5 91176		-	4117	India
<i>A. hypogaea</i> L. ssp <i>hypogaea</i> Krap et Rig var <i>hypogaea</i> (virginia)				
6 Robut 33-1		-	799	India
7 M 13		-	156	India
8 Makulu Red		-	6391	Zambia

¹ G = Gregory, H = Hammons, K = Krapovickas, L = Langford, P = Pietrarello

² ICG = ICRISAT Groundnut Accession

Results and Discussion

Transfer of rust resistance

Genome analysis in the section *Arachis* has revealed that *A. hypogaea* is a segmental allotetraploid with two genomes "A" and "B" each with base number 10. Among the diploid species there are several species with the "AA" genomic constitution although grouped as "A" genome species, these differ in karyotype and there are genetic differences within "A" genome species (Singh and Moss 1982, 1984a), the "BB" genome is represented by a single species, *A. batizocoi* (Husted, 1936, Smartt et al 1978, Stalker and Dalmacio 1981, Singh and Moss, 1982). Further studies have revealed that the two genomes "A" and "B" are closely related. *A. hypogaea* forms predominantly bivalents, suppressing A-B intergenomic pairing. However, such a suppression of A-B pairing does not seem to occur at different levels of ploidy in its experimental hybrids with wild species (Smartt and Stalker 1983, Singh and Moss 1984a). Therefore, genetic introgression from wild diploid species of section *Arachis* into *A. hypogaea* is possible provided suitable ploidy and genomic manipulations are adopted.

The cytogenetic manipulations used at ICRISAT to facilitate transfer of rust resistance from wild diploid species into *A. hypogaea* outlined in Figure 1 are discussed below.

Crosses between tetraploid *A. hypogaea* and diploid species

A. hypogaea is freely crossable with these diploid species and direct hybridization between them and *A. hypogaea* for gene transfer is the first logical proposal. Eight rust-resistant wild diploid species were crossed as male parents with cultivars belonging to two subspecies of *A. hypogaea*, and triploid hybrids were established. The hybrids were vigorous, with intermediate leaflet size and a trailing habit, and expressed the dominant morphological features of the wild species, they were also resistant to groundnut rust.

Cytological analysis of these hybrids revealed that the 10 chromosomes contributed by the wild species paired with 10 corresponding chromosomes of the homologous genome of *A. hypogaea* to form 10 bivalents. The 10 chromosomes of the non-homologous genome of *A. hypogaea* predominantly remained unpaired, as univalents. Homoeology of

wild species chromosomes with the non-homologous genome of *A. hypogaea* resulted in intergenomic pairing and the formation of more than 10 bivalents, or of multivalents in some pollen mother cells (PMCs) (Singh 1985). Such a pairing behavior indicates that meiotic recombination between wild and cultivated species does occur, but the gametes so formed abort as a result of irregular meiosis caused by high frequency of univalents, thus rendering the triploid hybrids sterile.

Use of amphiploids (hexaploids) of triploid hybrids.

Sterile triploids were treated with colchicine to double the chromosome number and restore fertility. This has been the most common method for genetic introgression from wild species and has been adopted by many workers (Smartt and Gregory 1967, Raman 1976, Moss et al 1981). At Reading University, UK and at ICRISAT Center, triploid hybrids between all 8 diploid species and *A. hypogaea* were raised to hexaploids. Cytologically, hexaploids formed mostly bivalents (range 10 to 30, mean 21 to 24) but a few multivalent associations (range 0 to 8, mean 1.1 to 2.7) have been observed (Singh 1985) involving the chromosomes of both wild and cultivated species (Spielman et al 1979). Consequently, recombinants with desirable traits of wild and cultivated species were formed, though at a very low frequency. They were screened for resistance to foliar diseases under field conditions during the rainy seasons of 1978 and 1979. Segregants resistant to rust and late leaf spot were selected and backcrossed with *A. hypogaea* to reduce their chromosome numbers and regain the agronomic traits of *A. hypogaea*.

Backcrossing of hexaploids with *A. hypogaea* resulted in the production of 32 *A. hypogaea*-like tetraploid derivatives incorporating genes from *A. chacoense*, *A. cardenasii* and 4 species HLK 410. These have been screened for resistance to rust under natural field conditions during several rainy seasons, and a large number of resistant segregants have been selected (Table 2).

Use of triploid progenies. Although triploids were reported sterile, they were found to produce some seeds and seedlings (Singh and Moss 1984b). Therefore, useful meiotic recombinations that occur in triploid hybrids are available for utilization. Eighty-two percent of plants in progenies from triploids were hexaploid, 10% aneuploid, and 8% tetraploid. The plants that have either 40, or less than 60, chromosomes are important, because their use redu-

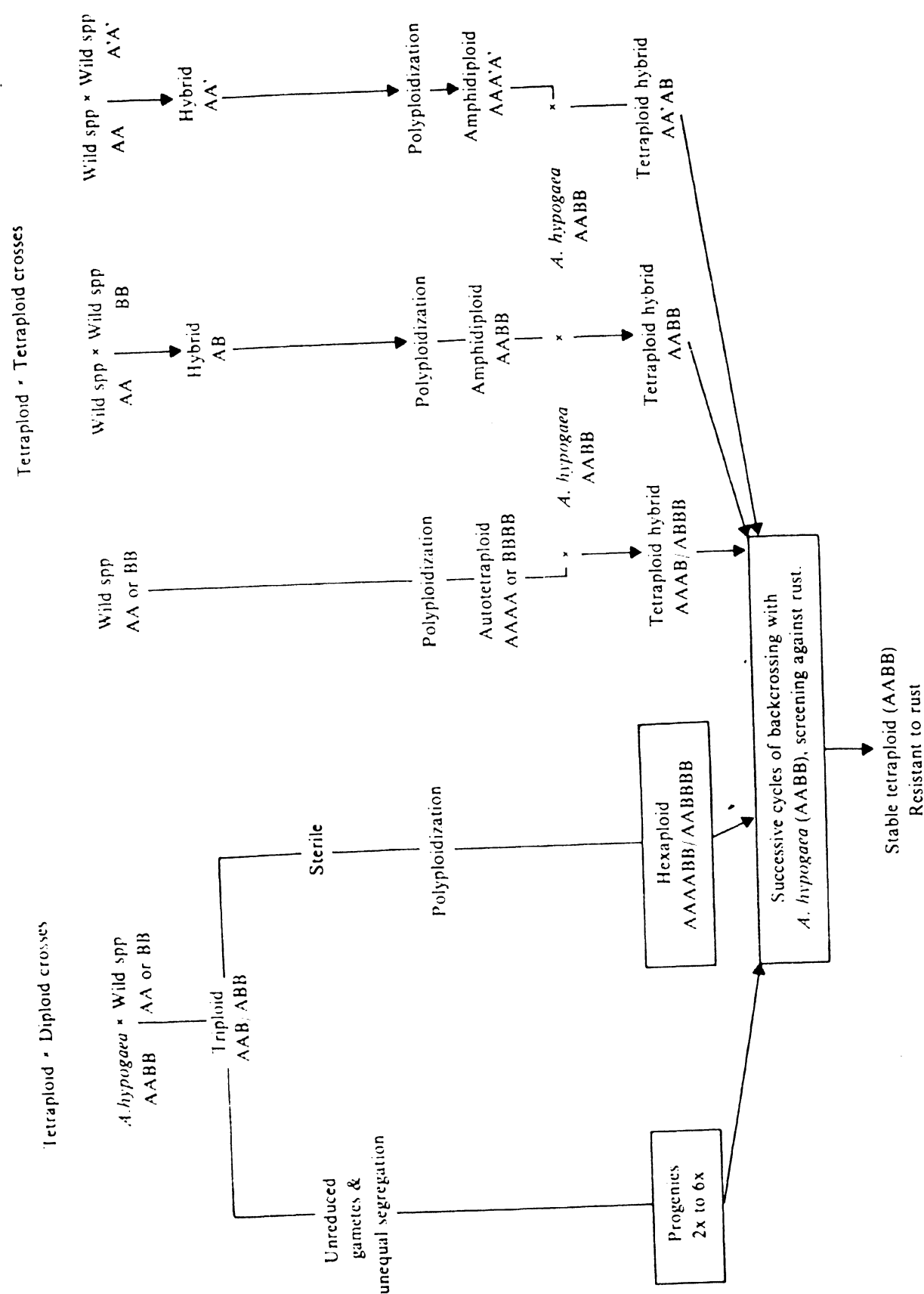


Figure 1. Manipulations for gene transfer in *Arachis*.

Table 2. Number of stable interspecific tetraploid derivatives produced and number of plants selected from their populations for resistance to rust, ICRISAT Center, 1982, 1984.

Route	Species	1982	1983	1984	Total
Self Triploids	<i>A. cardenasii</i>	5 (7) ¹	8 (14)	1	14 (21)
	<i>A. chacoense</i>	2		1	3
Hexaploids	<i>A. cardenasii</i>	11 (23)	1 (229)	(6)	12 (258)
	<i>A. chacoense</i>	5 (9)	2 (4)	(33)	7 (46)
	<i>A. sp</i> HLK 410	1 (6)	2 (1)	10	13 (7)
Autotetraploids	<i>A. batizocoi</i>	2	4 (1)	(2)	6 (3)
	<i>A. correntina</i>		2		2
	<i>A. sp</i> GKP 10038	1			1
	<i>A. sp</i> HLK 410	1			1
	<i>A. villosa</i>	1			1
Amphidiploids	<i>A. batizocoi</i> × <i>A. chacoense</i>		1		1
	<i>A. batizocoi</i> × <i>A. correntina</i>		2		2
	<i>A. batizocoi</i> × <i>A. duranensis</i>	1	2 (27)	(484)	3 (511)
	<i>A. correntina</i> × <i>A. batizocoi</i>	2	2 (1)	11	15 (1)
	<i>A. correntina</i> × <i>A. chacoense</i>		4		4
	<i>A. correntina</i> × (<i>A. chacoense</i> × <i>A. cardenasii</i>)	1			1
	<i>A. correntina</i> × <i>A. villosa</i>	2	2	1	5
	<i>A. duranensis</i> × <i>A. cardenasii</i>		1		1
	<i>A. duranensis</i> × <i>A. chacoense</i>	1	3 (1)		4 (1)
	<i>A. duranensis</i> × <i>A. sp</i> GKP 10038		1		1
	<i>A. villosa</i> × <i>A. batizocoi</i>	2	5 (18)	(1)	7 (19)
	<i>A. villosa</i> × <i>A. sp</i> HLK 410	2			2
	<i>A. villosa</i> × <i>A. duranensis</i>		1	3	4
	<i>A. sp</i> GKP 10038 × <i>A. sp</i> HLK 410	1	(1)	16	17 (1)
	<i>A. sp</i> HLK 410 × <i>A. chacoense</i>		4		4
	<i>A. sp</i> HLK 410 × <i>A. sp</i> GKP 10038		1		1
	Total	41 (45)	48 (297)	43 (526)	132 (868)

1. Figures in parentheses are number of plants selected.

ces the number of backcross cycles required for the production of stable *A. hypogaea*-like tetraploid derivatives compared to the number of backcross cycles required from hexaploids.

Backcrossing the progenies from triploids with *A. hypogaea* has resulted in the production of 17 stable *A. hypogaea*-like tetraploids involving *A. chacoense* and *A. cardenasii*. Rust-resistant segregants were selected by field screening of these tetraploid derivatives (Table 2). Certain selections were also found resistant to late leaf spot (*Phaeoisariopsis personata* (Berk. & Curt.) v. Arx.)

The gametic (pollen) fertility of these triploids also indicates that they can be backcrossed with *A. hypogaea* to produce *A. hypogaea*-like tetraploid derivatives, as has been done in wheat (Kerber and Dyck 1973).

Crosses between tetraploid *A. hypogaea* and synthetic tetraploids

The difference in ploidy levels between diploid wild *Arachis* species and tetraploid cultivated *A. hypogaea* restricts sexual genetic introgression, because of the low fertility of the triploid hybrids. Raising the ploidy level of the diploid species to that of *A. hypogaea* and then crossing with *A. hypogaea* at the tetraploid level is a useful option for gene transfer, as in cotton, potato and tobacco (Knight 1953, 1954, Wangenheim 1955, Stavely et al. 1973).

Use of autotetraploids of diploid species. The autotetraploids of diploid species not only facilitate crossing at the same ploidy level as the cultivated species, but also provide an additional dose of the

desired traits and may also permit a forced homoeologous intergenomic (A-B) pairing to effect genetic alteration in the non-homologous genome of *A. hypogaea*.

Autotetraploids of 6 wild diploid species have been crossed with *A. hypogaea*. The F_1 plants were vigorous, and resembled *A. hypogaea*. These hybrids can be either AAAB or ABBB depending on whether an AA or BB species autotetraploid was crossed with *A. hypogaea*. Homology of a genome of *A. hypogaea* with a diploid species can result in the formation of bivalents due to intragenomic (A-A or B-B) pairing. Homoeology with the other genome of *A. hypogaea* results in the formation of more than 10 bivalents (11.2 to 14.1) due to intergenomic (A-B) pairing or multivalents (1.8 to 2.5) due to intra- and intergenomic (A-A-B; A-B-B; A-A-A-B; A-B-B-B) pairing (Singh 1985). The hybrids between *A. hypogaea* and the autotetraploids of section *Arachis* species were resistant to rust (Singh et al. 1984c) and were backcrossed with *A. hypogaea*. Eleven stable *A. hypogaea*-like derivatives have been produced. Of these, six were derived from the hybrids between *A. hypogaea* and autotetraploid *A. batizocoi*. These have been screened during rainy seasons. Several rust-resistant segregants have been selected (Table 2), and are being advanced.

Use of amphidiploids of diploid species. The presence of two homoeologous genomes, "A" and "B", among diploid wild *Arachis* species, and the occurrence of both genomes in *A. hypogaea*, suggest that hybridization at the same ploidy level between tetraploid *A. hypogaea* and synthetic amphidiploids of diploid wild species can be a promising approach to provide a high degree of recombination and highly fertile hybrids.

Amphidiploids were produced from sterile or partially sterile interspecific hybrids, representing 34 combinations of the 8 diploid wild species of section *Arachis*. Of these, 22 (AABB and AAAA amphidiploids) have been crossed with *A. hypogaea*. All the F_1 hybrids between *A. hypogaea* and amphidiploids were resistant to rust. The hybrids between *A. hypogaea* and AABB amphidiploids had higher bivalent associations (14.4 to 16.4) and pollen and pod fertility, than the hybrids between *A. hypogaea* and AAAA amphidiploids (10.8 to 15.0). In *A. hypogaea* × AAAA amphidiploid hybrids (AAAB), homoeology between A and B genome results in the formation of more than 10 bivalents and a few multivalents as a result of intra and intergenomic pairing. Subsequent backcrossing with *A. hypogaea*, sometimes

with intervening selfing generations, have resulted in the production of 72 stable *A. hypogaea*-like tetraploid progenies. These tetraploid progenies were screened for resistance to rust and late leaf spot during rainy seasons. In derivatives *A. hypogaea* × (*A. batizocoi* × *A. duranensis* amphidiploid) and *A. hypogaea* × (*A. villosa* × *A. batizocoi* amphidiploid) hybrid fertility has enabled the advancement of the progenies into subsequent generations even without backcrossing. Resistant segregants have been selected from these progenies (Table 2).

A large number of *A. hypogaea*-like interspecific derivatives incorporating genes from diploid wild *Arachis* species have been produced that confer a high degree of resistance against rust. The most advanced lines involve a perennial species such as *A. cardenasii*, (resistant to both rust and late leaf spot) and three annual species, *A. batizocoi*, *A. species* GKP 10038, and *A. duranensis* (resistant to rust and some groundnut pests). A number of been evaluated in replicated trials characters and for rust resistance (I frequently several lines resistant to rust for agronomic traits, e.g., ICG(C) 8, and ICG(C) 12, are being many locations in the All India Research Project for Oil Seeds (AI). In addition, a large number of derivatives four other species that are resistant to other pathogens and pests are being

Genetics of rust resistance

Preliminary investigations on the rust resistance derived from diploid wild species shown that the F_1 hybrids between diploid species, their autotetraploids, are resistant to rust, suggest resistance is governed by a partially (Singh et al. 1984c). Identification loci is in progress. The interspecific *A. hypogaea*-like derivatives with wild species have been crossed susceptible and rust-resistant *A. hypogaea* rust-resistant germplasm lines have also been crossed with susceptible cultivars. These studies should reveal the inheritance pattern of the two resistances and their relationships. These results have generated a great interest in the utilization of wild species as sources of rust resistance and in combining resistance of wild species with that of *A. hypogaea*.

Table 3. Rust resistance (1-9 scale) and other agronomic characters of some of the advanced interspecific derivatives¹ (8 × 8 triple lattice, plot size 10.8 m²), ICRISAT Center, 1984 rainy season.

Code	Pedigree	Rust score	Yield (kg ha ⁻¹)	Haulm weight (kg ha ⁻¹)	Shelling %
CS 36	<i>A. hypogaea</i> × <i>A. cardenasii</i>	1.4	4470	8690	71
2403	<i>A. hypogaea</i> × <i>A. cardenasii</i>	1.6	3600	6150	71
2404/4	<i>A. hypogaea</i> × <i>A. cardenasii</i>	1.6	4770	7190	70
CS 7	<i>A. hypogaea</i> × <i>A. cardenasii</i>	1.7	4280	8580	75
CS 33	<i>A. hypogaea</i> × <i>A. cardenasii</i>	1.7	4200	9880	74
2245	<i>A. hypogaea</i> × <i>A. cardenasii</i>	1.9	4050	7410	69
CS 2	(<i>A. batizocoi</i> × <i>Arachis</i> sp GKP 10038) × <i>A. hypogaea</i>	1.7	3510	5590	61
CS 19	<i>A. hypogaea</i> × (<i>A. batizocoi</i> × <i>Arachis</i> sp GKP 10038)	1.7	2730	4840	66
Checks					
Robut 3301		8.9	2880	3600	54
TMV 2		8.9	2170	4940	40
EC 76446 (292)		2.8	3620	5840	54
NC Ac 17133-RF		7.2	4410	5000	61
	CV (%)	27	15	18	16
	SE	±0.7	±314.4	±764.7	±2.9

¹ All the derivatives are virginia-bunch type

Acknowledgement: We thank Dr. D.C. Sastri and Mr. T.P.S. Rau for the help extended in preparation of the manuscript and Mrs. Sashikala for typing assistance.

References

- Gregory, W.C., Gregory, M.P., Krapovickas, A., Smith, B.W., and Yarbrough, J.A. 1973. Structures and genetic resources of peanuts. Pages 47-133 in *Peanuts—culture and uses*. Stillwater, Oklahoma, USA: American Peanut Research and Education Association.
- Husted, L. 1936. Cytological studies of the peanut. II. Chromosome number, morphology and behaviour and their application to the origin of cultivated forms. *Cytologia* 7:396-423.
- Kerber, E.R., and Dyck, P.L. 1973. Inheritance of stem rust resistance transferred from diploid wheat (*Triticum monococcum*) to tetraploid and hexaploid wheat and chromosome location of the gene involved. *Canadian Journal of Genetics and Cytology* 15:397-409.
- Knight, R.L. 1953. The genetics of blackarm resistance IX. The gene B6m from *Gossypium arboreum*. *Journal of Genetics* 51:270-75.
- Knight, R.L. 1954. Cotton breeding in the Sudan. Parts I and II. Egyptian cotton. *Empire Journal of Experimental Agriculture* 22:68-92.
- Moss, J.P., Spielman, I.V., Burge, A.P., Singh, A.K., and Gibbons, R.W. 1981. Utilization of wild *Arachis* species as a source of *Cercospora* leaf spot resistance in groundnut breeding. Pages 673-677 in *Perspectives in cytology and genetics* (Manna, G.K., and Sinha, U., eds.). New Delhi, India: Hindasia Publications.
- Raman, V.S. 1976. Cytogenetics and breeding in *Arachis*. New Delhi, India: Today and Tomorrow Printers and Publishers.
- Singh, A.K. 1985. Genetic introgression from compatible *Arachis* species into groundnut. Pages 107-117 in *Proceedings of the International Workshop on Cytogenetics of Arachis* 31 Oct-2 Nov 1983, ICRISAT Center, India. Patancheru, A.P. 502 324, India: International Crops Research Institute for the Semi-Arid Tropics.
- Singh, A.K., and Moss, J.P. 1982. Utilization of wild relatives in genetic improvement of *Arachis hypogaea* L. Part 2. Chromosome complements of species of section *Arachis*. *Theoretical and Applied Genetics* 61:305-314.
- Singh, A.K., and Moss, J.P. 1984a. Utilization of wild relatives in genetic improvement of *Arachis hypogaea* L. Part 5. Genome analysis in section *Arachis* and its implications in gene transfer. *Theoretical and Applied Genetics* 68:355-364.
- Singh, A.K., and Moss, J.P. 1984b. Utilization of wild relatives in genetic improvement of *Arachis hypogaea* L. Part VI. Fertility in triploids: cytological basis and breeding implications. *Peanut Science* 11:17-21.

• Singh, A.K., Moss, J.P., and Sastri, D.C. 1983. Utilization of wild relatives in genetic improvement of *Arachis hypogaea* L. I Technique. Pages 151-157 in Current approaches in cytogenetics (Sinha R.P., and Sinha, U., eds.). New Delhi, India: Hindasia Publications.

Singh, A.K., Subrahmanyam, P., and Moss, J.P. 1984c. The dominant nature of resistance to *Puccinia arachidis* in certain wild *Arachis* species (Summaries in Es, Fr.) Oleagineux 39:535-537.

Smartt, J., and Gregory, W.C. 1967. Interspecific cross-compatibility between the cultivated peanut *Arachis hypogaea* L. and other members of the genus *Arachis*. Oleagineux 22 455-459.

Smartt, J., Gregory, W.C., and Gregory, M.P. 1978. The genomes of *Arachis hypogaea* I Cytogenetic studies of putative genome donors. Euphytica 27:665-675.

Smartt, J., and Stalker, H.T. 1983. Speciation and cytogenetics in *Arachis*. Pages 21-49 in Peanut science and technology (Pattee, H E , and Young, C T , eds) Yoakum, Texas, USA: American Peanut Research and Education Society.

Spielman, I.V., Moss, J.P., and Burge, A.P. 1979. Chromosome loss and meiotic behaviour in interspecific hybrids in the genus *Arachis* and their implication in breeding for disease resistance. Zeitschrift für Pflanzenzuchtung 83:236-250.

Stalker, H.T., and Dalmacio, R.D. 1981. Chromosomes of *Arachis* species, section *Arachis*, Leguminosae. Journal of Heredity 72:403-408.

Stavely, J.R., Pittarelli, G.W., and Burk, L.G. 1973. *Nicotiana repanda* as potential source of disease resistance in *N. tabacum*. Journal of Heredity 64:265-271.

Subrahmanyam, P., Ghanekar, A.M., Nolt B.L., Reddy, D.V.R., and McDonald, D. 1985. Resistance to groundnut diseases in wild *Arachis* species. Pages 49-55 in Proceedings of the International Workshop on Cytogenetics of *Arachis*, 31 Oct-2 Nov 1983, ICRISAT Center, India. Patancheru, A.P. 502324, India International Crops Research Institute for the Semi-Arid Tropics.

Subrahmanyam, P., Gibbons, R.W., Nigam, S.N., and Rao, V.R. 1980. Screening methods and further sources of resistance to peanut rust. Peanut Science 7:10-12.

Subrahmanyam, P., Reddy, D.V.R., Gibbons, R.W., Rao, V.R., and Garren K.H. 1979. Current distribution of groundnut rust in India. PANS 25:25-29.

Subrahmanyam, P., McDonald D., Gibbons, R.W., Nigam, S.N., and Nevill D.J. 1982. Resistance to rust and late leaf spot disease in some genotypes of *Arachis hypogaea*. Peanut Science 9:6-10.

Subrahmanyam, P., Moss, J.P., and Rao, V.R. 1983. Resistance to peanut rust in wild *Arachis* species Plant Disease

Von Wangenheim, K.H.F., 1955. Zur Ursache der kreuzungsschwierigkeiten Zwischen *Solanum tuberosum* L. and *S. acaule* Bittl. bzw. *S. stoloniferum* Schlecht. et. Bouche. (In De.). Zeitschrift für Pflanzenzuchtung 34 7-48.