

## HOST RANGE OF SUGARCANE MOSAIC VIRUS (SCMV-B) FROM RAYALASEEMA

K. GOPAL\*\*, K. SEETHARAMA REDDY, H. D. UPADHYAYA\*\* AND D. RAJARAMA REDDY

Department of Plant Pathology

S. V. Agricultural College, Tirupati-517 502

\*\*Regional Research Station, P. B. No. 24, Raichur 584 101

### SUMMARY

Sugarcane mosaic virus (SCMV) is important seed piece transmissible disease. Most of the cultivated varieties are susceptible to this virus in Rayalaseema region of Andhra Pradesh. Though lot of work has been done on this disease in U. P. (Rishi, 1969; Bhargava *et al.*, 1971 a, b; Srivastava *et al.*, 1977), Tamil Nadu (Singh, 1976) and Maharashtra (Ghorpade and Joshi, 1982), no systematic work has been done in Rayalaseema region of Andhra Pradesh. Therefore preliminary study of host range of SCMV was made.

The virus inoculum was collected from COT-8201 (ruling Sugarcane variety in A. P.) from Agricultural Research Station, Perumalapalli, and it was maintained on sweet sorghum cv. Rio. Twenty plants of each species were inoculated with freshly prepared inoculum of the virus. The inoculated plants were observed for 4-6 weeks in the glass house. Healthy sweet sorghum cv Rio was used as control. The plants which did not show the symptoms of disease were further tested for susceptibility by back inoculation to sweet sorghum cv Rio by the sap extracted from the inoculated plants. The virus was identified as SCMV-B based on morphological and serological tests (Ouchterlony double gel diffusion and immunosorbent electron microscopy) against antisera of SCMV-A, B, D, H and I.

Different plant species used for host range studies of SCMV-B are presented in table. Perusal of the table indicated that majority of plants in family Gramineae were susceptible to SCMV-B. However, nine plants of Leguminosae, four of Solanaceae, two each of Chenopodiaceae, Violaceae, Euphorbiaceae and Caryophyllaceae, three each of Malvaceae and Compositae did not show SCMV-B symptoms on inoculation. Of the 25 species tested in Gramineae, only four were not infected by SCMV-B. These included *Cyperus rotundus*, *S. bulbosus*, *Hordeum vulgare* and *Oryza sativa*. However, rice and barley were reported to be susceptible to SCMV by Abbott and Tippett (1964) and Anzalone and Lemey (1968). Infection in Gramineae, ranged from 10 to 100 per cent (Table). *Avena sativa*, *Brachiaria mutica*, *Cynodon dactylon*, *Eleusine coracana* cv. Kalyani and AKP-2, *Leptochloa panica*, *Panicum miliare*, *Paspalum conjugatum*, *Polypogon monspeliensis* are the new hosts found susceptible to SCMV-B. Because of their wide prevalence around the vicinity of sugarcane field the possibility of posing problems as potential hosts of the virus cannot be ruled out.

### ACKNOWLEDGEMENT

Authors are grateful to Dr. A. G. Gillaspei Jr., Applied Plant Pathology lab. Beltsville Agricultural Research Centre, Beltsville, Maryland 20705, USA, for providing antisera of SCMV (A, B, D, H and I) and seeds of sweet sorghum cv Rio for this study.

TABLE 1

Host range of Sugarcane mosaic virus (SCMV-B)

Plant species	Infection (%)
FAMILY : GRAMINEA	
<i>Avena sativa</i> L.	20
<i>Bracharia mutica</i> (Forsk) Stapf.	30
<i>Cenchrus ciliaris</i> L.	40
<i>Cynodon dactylon</i> (L.) Pers.	20
<i>Cyperus rotundus</i> L.	0
<i>C. bulbosus</i> Vahl.	0
<i>Dactyloctenium aegyptium</i> (L) Beauv.	50
<i>Digitaria adscenens</i> Henn.	40
<i>Echinochloa crusgalli</i> Beauv.	20
<i>Eleusine coracana</i> Gaertn. c. v. Kalyani	30
<i>E. Coracana</i> Gaertn. cv. APK-2	20
<i>E. Indica</i> Gaertn.	20
<i>Hordeum vulgare</i> L.	0
<i>Leptochloa panica</i> Pers.	10
<i>Oryza sativa</i> L. cv. IR-50	0
<i>Panicum miliaceum</i> L.	30
<i>P. miliare</i> Lam.	40
<i>Paspalum conjugatum</i> Berg.	40
<i>Pennisetum typhoides</i> (Burn.) Stapf and C. E. Hubb.	40
cv. Balaji	40
cv. BJ-104	50
cv. TBS-11	40
<i>Polypogon monspeliensis</i> L.	30
<i>Setaria italica</i> L. cv. Arjuna	30
cv. SiA-326	30
<i>Setaria glauca</i> L.	10
<i>Sorghum bicolor</i> (L) Moench cv. N-13	100
cv. CSH-5	90
cv. CSH-9	95
cv. Co-24	95
<i>Triticum aestivum</i> L.	20
<i>Urochloa mosambicensis</i> (Hack) Dandy	20
<i>Zea mays</i> L. cv. DHM-130	60
cv. Popcorn	60

## REFERENCES

- Abbott, I. V. and Tippet, R. L. 1964. Additional hosts of Sugarcane mosaic virus *Plant Dis. Repr.* 48 : 442.
- Anzures, E. and Lemey H. A. 1968. Possible differential reaction of certain rice varieties to Sugarcane mosaic virus. *Plant. Dis. Repr.* 52 : 775-77.
- Bhatnagar, K. S., Joshi, R. D. and Rishi, Narayan. 1971a. Occurrence of strains A and F of sugarcane mosaic virus in Uttar Pradesh (India). *Proc. XIV Cong. Int. Soc. Sugarcane Technologists, Louisiana, U. S. A.* pp. 949-54.
- Bhatnagar, K. S., Joshi, R. D. and Rizvi, S. M. A. 1971b. Some observations on the insect transmission of Sugarcane mosaic virus in India *Sugarcane Pathol. Newsl.* 6 : 20-22.
- Chatterpade, L. N. and Joshi, R. D. 1981. Effect of Sugarcane mosaic virus on photosynthetic enzymes in in Sugarcane clone Co-740. *Sugarcane Pathol. Newsl.* 28 : 42.
- Rishi, Narayan. 1969. Studies on the mosaic disease of sugarcane in Uttar Pradesh. Ph. D. Thesis, University of Gorakhpur, Gorakhpur, pp. 142.
- Singh, S. 1976. *Dactyloctenium aegyptium* ; an additional host of Sugarcane mosaic virus. *Sugarcane Pathol. Newsl.* 15/16 : 1-2.
- Srivastava, V. K., Tripathi, A. M., Shukla, K. and Agarwal, N. 1977. In vitro effect of heteropolyanions on some plant viruses. *Sugarcane Pathol. Newsl.* 19 : 10-11.

Wider spacing both row to row and plant to plant significantly increased the disease incidence. Almost double disease incidence was recorded in  $60 \times 15$  cm spacing irrespective of season as compared with  $30 \times 10$  cm spacing. Similar results were reported by Heathcote for beet mild yellows virus and by Reddy *et al.* (1983) for tomato spotted wilt virus. Greater disease incidence in wider spacing could be due to greater activity of whiteflies in full sunlight. In narrow spacing, the dense plant population cut the light considerably going to less activity of the whiteflies. The second possible reason may be that after monsoon rainfall, a different micro-climate develops in dense plant population which may be fatal for the vector. It has been observed that in narrow spacing, on the surface of leaves of densely populated plants, more water drops exist for a longer time and when the whiteflies visit such leaf surface their wings are stuck in these drops and ultimately the white-

#### REFERENCES

- P. S. 1990. Studies on crinkle stunt disease of French bean (*Phaseolus vulgaris* L.). Ph. D. thesis, G. B. Pant Univ. of Agri. and Tech., Pantnagar, India.
- Patil, G. D. 1974. The effect of plant spacing, nitrogen fertilizer and irrigation on the appearance of symptoms and spread of virus yellows in sugarbeet crops. *J. Agric. Sci. Coimbatore*, 82: 53-60.
- P. S. and Azmi, O. R. 1988. *Bemisia tabaci* a vector of Frenchbean leaf crinkle disease. *Indian Phytopath.* 41: 268.
- Prasad, V. R., Amin, P. W., Donald, D. Mc and Ghanekar, A. M. 1983. Epidemiology and control of groundnut bud necrosis and other diseases of legumes caused by TSWV. In: *Plant Virus Epidemiology*, C. Plumb and J. M. Thresh (eds.), pp. 93-102, Blackwell scientific Publications.