OVERCOMING INCOMPATIBILITY IN WIDE CROSSES

D. C. SASTRI and M. MALLIKARJUNA

CHISG

Wild species of crop plants attract much attention as a valuable gene pool. A few successes in improving crop plants have resulted from crossing wild taxs with cultivated ones. Several other taxs are not crossable with their cultivated relatives and are therefore unavailable for sexual gene transfers. Methods for breaking these barriers to interspecific hybridization and hybrid production have been developed.

The literature on exotic germplasm abounds with examples of genetic introgression from wild taxa conventionally crossed with their cultivated relatives. But there is a wealth of germplasm that cannot be crossed with cultivated taxa.

The barriers to hybridization were known even before the present range of germplasm became available. During the last five decades, interest in incompatibility has increased to encompass phylogenetic-taxonomic purposes and the genetic improvement of crop plants. While emerging somatic methods show promise, sexual manipulation is still the first choice, and sexual methods have contributed significantly to improving crops.

The germplasm amenable to sexual manipulation is difficult to quantify. It may be an insignificant proportion of existing germplasm. Even in the well-worked genus <u>Nicotiana</u>, of which about 65 species are known, only a little more than 300 hybrids have been realized. About 90% of the crosses in this genus have not. The situation is similar for most other crops. A number of review articles and books on incompatibility and methods to break it are available (9, 15, 16, 17, 18, 41, 49, 51).

CAUSES OF SEED FAILURE IN INCOMPATIBLE CROSSES

The characteristic prefertilization barriers and postfertilization breakdown of zygote or embryo in incompatible crosses have been extensively described. Inhibition of pollen germination on stigma and pollen tube growth through the style,

Cytogeneticist and research associate, International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru P. O. 502 324, A. P., India. failure to fertilize, and zygote growth cessation at some stage of development are common to many failing crosses (41, 49, 56). Unfortunately, their molecular causes are little understood. Even so, the fragmentary information available has led to development of methods for breaking crossability barriers in a few taxa.

METHODS

Among the several methods used are recognition pollen, hormones, immunosuppressants, and temperature. Early surgical methods are also important (28, 37, 41). Delayed and bud pollinations have overcome both sporophytic (Brassica spp.) and gametophytic (Petunia spp.) self-incompatibilities in a few taxa, but not any interspecific incompatibility. Their success depends on the stigma's receptiveness, i.e., buds of Arachis hypogaea even a day before anthesis are not receptive even to compatible pollen; so bud pollination does not work in A. hypogaea (43).

Growth hormones

The discovery of growth hormones in plants inspired investigations on their potential in overcoming incompatibility, which is not mentioned in books and conferences on plant growth hormones. The influence of exogenous and endogenous hormones on developing fruit has been investigated in normal compatible (self) crosses but rarely in incompatible crosses. The effects of different growth hormones on free pollen grains in in vitro cultures have been extensively studied (25, 26, 51). The effects of hormones on compatible and self-incompatible pollen tube growth in pistils of Lilium longiflorum (14) and a few other species (9, 41) have been reported but the studies are too few on self-incompatible taxa, let alone for conclusions Hormone applications have, however, been interspecific ones. found to enhance a few incompatible fruit and seed set crosses.

Crane and Marks (7) and Brock (5) first used hormones to delay floral abscission in incompatibly pollinated flowers with short life. Interest in use of hormones in incompatible crosses subsequently grew. The latest hormone-aided hybridizations are interspecific crosses in <u>Arachis</u>. In a few cases application of gibberellic acid to the bases of incompatibly pollinated flowers stimulates the initiation and geotropic elongation of the peg, the aerial phase of <u>Arachis</u> fruit (Table 1; 43, 44, 45). Subsequent application of IAA, NAA, or kinetin increases the number of pods (42) and cultured ovules and embryos (29).

Over 20 interspecific or intergeneric hybrids have resulted from exogenous application of natural or synthetic plant growth hormones (41). We have yet to understand how hormones stimulate hybrid development from incompatible crosses. A comparison of hormonal balances in ovaries between compatibly and incompatibly pollinated flowers is called for and specific effects of exogenous hormones in successful cases need study. Nessling and Morris (32) found that endosperm from an interspecific <u>Phaseolus</u> cross does not develop normally and has much less cytokinin than

Cross	Pollinations (no.)	Pegs (%)	Pods/pegi (%)
Section Arachis/Section Triseminale			
A. duranensis (2n=20)/A. pusilla (2n=20)	33	79	39
A. hypogaea cv. Robut 33-1/A. pusilla (2n=20)	78	46	0
Section Arachis/Section Extranervosae			
A, hypogeee cv Robut 33-1/A, villosulicerpe (2n=20)	39	59	3
A. hypogaea cv MK 374/A. villosulicarpa (2n=20)	9	89	11
Section Extranervosae/Section Triseminale			
A. villosulicarpa (2n=20)/A. pusilla (2n=20)	24	54	46
Section Arachis × Section Rhizomatose			
A. hypogeee cv Robut 33-1/Arechis sp. Coll, No. 9649	82	44	6
A. hypogaea cv Robut 33-1/Arachis sp. Coll. No. 9797	46	67	2
A. hypogene cv Robut 33-1/Arechis sp. Coll. No. 9808	• 26	62	0
A. hypogaea cv TMV2/Arachis sp. PI 276233	408	76	20
A. hypogaea cv TMV2/Arachis sp. Coll, No. 9649	11	73	0
A. hypogaea cv MK 374/Arachis sp. PI 276233	648	68	32
A. hypogaea cv MK 374/Arachis sp. Coll, No. 9649	26	42	15
A. hypogaea cv M 13/Arachis sp. PI 276233	75	56	5
A. hypogaes cv Chico/Arachis sp. PI 276233	58	66	9
A. hypogees cv Chico/Arachis sp. Coll, No. 9649	26	73	19

Table 1. Peg and pod production after sibberellin treatment in some intersectional (

that from selfed pollinations. Such studies not only help elucidate hormonal regulation of fruit morphogenesis, but also assist in formulating strategy for further work with incompatible crosses.

Recognition pollen

Since Tsinger and Petrovskaya-Bananova (62) showed "pollen grain walls [to be] physiologically active substances," interest in postpollination changes in pollen grains and pistil has grown. It is now believed that extracellular (wall) pollen proteins extracellular (surface) stigma papillae communicate with proteins: pollen grains are thereby recognized as either compatible or incompatible (16, 17, 18). The existence of pollen grain walls (16) and stigma surfaces (19, 20) with a protein component has been demonstrated in a number of taxa, and in a few, even stigmas of immature flowers are lined with proteins (52, 53, 50).

Knox et al (27) exploited this information to produce hybrids from an interspecific cross in <u>Populus</u>. Earlier Stettler (57) had achieved interspecific hybridization in <u>Populus</u> by mixing live incompatible pollen grains with gamma-irradiated (killed) compatible pollen grains called recognition or mentor pollen (27). Mentor pollen stimulated incompatible pollen grains to produce fruit and seed.

Subsequently this method was tested in other taxa and in other systems of self-incompatibility and interspecific incompatibility. Sporophytic self-incompatibility was overcome in <u>Cosmos bipinnatus</u> (21), <u>Brassica oleracea</u> (40), and <u>Raphanus</u> <u>sativus</u> (48), but not in <u>Brassica campestris</u> (48). <u>Gametophytic</u> self-incompatibility was overcome in apple (8), <u>Petunia hybrida</u> (47, 48) and <u>Nicotiana</u> alata (55), but not in <u>Oenothera</u> organensis (55). Recognition pollen did not work in other interspecific crosses in <u>Sesamum</u> (46), <u>Arachis</u> (43), <u>Ipomoea</u> (13), <u>Trifolium</u> (58), and <u>Cucumis</u> (10).

Sastri and Shivanna (48) have suggested that the method of killing the compatible pollen grains determines success. The recognition pollen can be prepared by storage at normal temperature, repeated freezing and thawing, and gamma-ray irradiation until pollen grains lose their ability to germinate. In a few instances the recognition pollen function could be effected by leachates of compatible pollen grains (21, 48).

Pandey (33, 34, 35) found that in <u>Nicotiana</u> gene transfers could be effected with just gamma-irradiated compatible pollen grains. Jinks et al (22) have confirmed this. Recently Shuzikuda et al (54) combined this method with ovule culture to transfer an incomplete chromosome complement of <u>Nicotiana</u> <u>tabaccum</u> into <u>N. rustica</u>, two species which are not otherwise crossable.

Immunosuppressants

About 10 yr ago Bates and his coworkers began using in wide hybridization experiments animal immunosuppressants including E-aminocaproic acid (EACA), chloramphenicol, acriflavin, salycylic acid, and gentisic acid. The success rate varied, but EACA was the most effective: with EACA, hybrids were obtained in crosses between barley and rye, durum wheat and barley, and bread wheat and barley (3, 4). EACA also stimulated embryo development in crosses between Triticum turgidum and Secale cereale (60, 61).

In another interspecific cross, <u>Vigna radiata/V</u>. <u>umbellata</u>, a foliar spray of EACA (100 ppm) effected twice as many hybrids as those that resulted from the unsprayed controls (1). Baker et al (2) obtained optimum results in the same cross by injecting 250 ppm of EACA. Chen et al (6) observed in two cultivars that foliar EACA spray for 14 d starting no later than the premeiotic stage of flower development delayed but did not prevent embryo abortion. EACA application to florets on 4 consecutive d after pollination reduced embryo recovery from 30 to 19%, but increased the number of ovaries forming embryo and endosperm in <u>Triticum</u> timopheevi/Secale cereale (31). It was not effective in <u>Festuca</u> arundinacea/Dactylis glomerata (30).

OTHER METHODS

Embryo, ovule, and ovary cultures have yielded hybrids in more than 50 cross combinations with early abortion of embryo or endosperm, or both (see 36, 41). According to Johnston et al (24), when there are ploidy differences between the parents, an abnormal endosperm is caused by a deviation of the maternal and paternal genome ratio from 2 to 1 in the endosperm itself. By assigning a specific number (endosperm balance number) to the endosperm of each parent irrespective of its ploidy, and by manipulating the numbers, Johnston and Hanneman (23) has succeeded in crossing some diploid Solanum species.

Ploidy manipulations are not new and a species with higher ploidy is generally better as a female parent. (Even when the parents have the same ploidy, one of them is raised to a higher level through colchicine treatment.) In other situations, a third species, itself crossable with each incompatible parent, is used as a bridge. Cultivar crossability differences indicate the need for a range of cultivars. Genes for crossability, such as Krl and Kr2 in wheat, should be investigated in other taxa. Most of these aspects have been reviewed by Rees et al (38), Thomas (59), Driscoll (11), and Riley et al (39).

CONCLUSIONS

Several means for hybridization between incompatible species have been found. Until the somatic methods (e.g., 12) can find wider applications, these methods, with or without modifications, must be exploited. In some instances combining methods with ovule or embryo culture is needed. For example, in a few interspecific Arachis, applying gibberellin crosses of to incompatibly pollinated flowers stimulated normal development of the pegs, and a few pods (Table 1) up to certain stages. Only ovules in the pods were found to be immature, with poorly differentiated embryos (29, 43, 44, 45). Ovules and embryos had to be cultured Similarly, in in vitro to yield plantlets (29, 43, 44). interspecific crosses in <u>Sesamum</u> (46) and <u>Cucumis</u>, (10) mentor pollen effected only part of the desired response. For further development, another method must be adopted.

Incompatibility must be better understood before it can be overcome and the wild germplasm exploited for further improvement of crops.

REFERENCES CITED

- AVRDC (Asian Vegetable Research and Development Center).
 1976. Progress report for 1975 (Mungbean). Shanhua, Taiwan, Republic of China. p. 35-40.
- Baker, L.R., N.C. Chen, and H.G. Park. 1975. Effect of an immunosuppressant on an interspecific cross of the genus Vigna. Hortic. Sci. 70:325-333.
- Bates, L.S. 1974. Wide crosses. In Proceedings: worldwide maize improvement in the 70s and the role for CIMMYT 5, IB-7B.
- 4. Bates, L.S., and C.W. Deyoe. 1973. Wide hybridization and cereal improvement. Econ. Bot. 27:401-412.
- 5. Brock, R.D. 1954. Hormone-induced pear-apple hybrids. Heredity 8:421-429.

- 6. Chen, N.C., J.F. Parrot, T. Jacobs, L.R. Baker, and P.S. Carlson. 1978. Interspecific hybridization of food grain legumes by unconventional methods of breeding. Pages 247-252 in International mungbean symposium. Asian Vegetable Research and Development Center, Taiwan.
- Crane, M.B., and E. Marks. 1952. Pear-apple hybrids. Nature (London) 170:1017.
- Dayton, D.F. 1974. Overcoming self-incompatibility in apple with killed compatible pollen. J. Am. Soc. Hortic. Sci. 99:190-192.
- 9. de Nettancourt, D. 1977. Incompatibility in Angiosperms. Springer-Verlag, Berlin.
- 10. Den Nijs., A.P.M., and E.H. Oost. 1980. Effect of mentor pollen on pollen pistil incongruities among species of <u>Cucumis</u> L. Euphytica 29:267-272.
- 11. Driscoll, C.J. 1981. Perspectives in chromosome manipulation. Philos. Trans. R. Soc. London 292:535-546.
- 12. Evans, D.A., C.E. Flick, S.A. Kut, and S.M. Reed. 1982. Comparison of <u>Nicotiana</u> tabacum and <u>N. nesophila</u> hybrids produced by ovule culture and protoplast fusion. Theor. Appl. Genet. 62:193-198.
- 13. Guries, R.P. 1978. A test of the mentor pollen technique in the genus Ipomoea. Euphytica 27:825-830.
- 14. Henny, R.J., and P.D. Ascher. 1973. Effect of auxin (3-indole acetic acid) on in vivo compatible and incompatible pollen tube growth in detached styles of <u>Lilium</u> longiflorum Thunb. Incompatibility Newsl. 3:14-17.
- Heslop-Harrison, J. ed. 1971. Pollen: development and physiology. Butterworths, London.
- Heslop-Harrison, J. 1975. The physiology of pollen grain surface. Proc. R. Soc. London 1908:275-299.
- 17. Heslop-Harrison, J. 1978. Genetics and physiology of angiosperm incompatibility systems. Proc. R. Soc. London 202B:73-92.
- 18. Heslop-Harrison, J. 1978. Recognition and response in the pollen-stigma interaction. Pages 121-138 in Cell-cell recognition. Vol. 32. A.S.G. Curtis, ed. Society for Experimental Biology, Cambridge, England.
- 19. Heslop-Harrison, Y. 1981. Stigma characteristics and angiosperm taxonomy. Nord. J. Bot. 1:402-420.
- 20. Heslop-Harrison, Y., and K.R. Shivanna. 1977. The receptive surface of the angiosperm stigma. Ann. Bot. 41:1233-1258.
- 21. Howlett, B.J., R.B. Knox, J.B. Paxton, and J. Heslop-Harrison. 1975. Pollen-wall proteins: physico-chemical characterization and role in self-incompatibility in <u>Cosmos bipinnatus</u>. Proc. R. Soc. London 188B:167-182.
- 22. Jinks, J.L., P.D.S. Caligari, and N.R. Ingram. 1981. Gene transfer in <u>Nicotiana</u> rustica using irradiated pollen. Nature (London) 291:586-588.

- 23. Johnston, S.A. and R.E. Hannenman, Jr. 1982. Manipulations of endosperm balance number overcome crossing barriers between <u>Solanum</u> species. Science 217:446-448.
- 24. Johnston, S.A., T.P.M. den Nijs, S.J. Peloquin, and R.E. Hannenman, Jr. 1980. The significance of genic balance to endosperm development in interspecific crosses. Theor. Appl. Genet. 57:5-9.
- 25. Johri, B.M., D.C. Sastri, and K.R. Shivanna. 1977. Pollen viability, storage and germination. Pages 120-139 in Advances in pollen spore research. P.K.K. Nair, ed. Vol. 2. Today and Tomorrow Printers and Publishers, New Delhi.
- 26. Johri, B.M., and I.K. Vasil. 1961. Physiology of pollen. Bot. Rev. 27:325-381.
- 27. Knox, R.B., R.R. Willing, and L.B. Pryor. 1972. Interspecific hybridization in poplars using recognition pollen. Silvae Genet. 21:65-69.
- 28. Maheshwari, P. 1950. An introduction to the embryology of angiosperms. McGrawhill Book Co. Inc., New York.
- 29. Mallikarjuna, N., and D.C. Sastri. 1983. In vitro culture of ovule and embryos from some interspecific crosses in the genus <u>Arachis</u> L. <u>In</u> Proceedings international workshop on cytogenetics of <u>Arachis</u>. (in press)
- 30. Matzk, F. 1981. Successful crosses between Festuca arundinacea Schreb. and Dactylis glomerata L. Theor. Appl. Genet. 60:119-122.
- 31. Mujeeb-Kazi, A. 1981. <u>Triticum timopheevii x Secale</u> cereale crossability. J. Hered. 72:227-228.
- Nessling, F.A.V., and D.A. Morris. 1979. Cytokinin levels and embryo abortion in interspecific <u>Phaseolus</u> crosses.
 Z. Pflanzenphysiol. 91:345-358.
- 33. Pandey, K.K. 1975. Sexual transfer of specific genes without gametic fusion. Nature (London) 256:311-312.
- 34. Pandev, K.K. 1978. Gametic gene transfer by means of irradiated pollen. Genetica 49:53-69.
- 35. Pandey, K.K. 1980. Further evidence for egg transformation in <u>Nicotiana</u>. Heredity 45:15-29.
- 36. Raghavan, V. 1977. Applied aspects of embryo culture. Pages 375-397 in Applied and fundamental aspects of plant cell, tissue and organ culture. J. Reinert and Y.P.S. Bajaj, eds. Springer-Verlag, Berlin.
- 37. Rangaswamy, N.S. 1963. Control of fertilization and embryo development. Pages 327-353 in Recent advances in the embryology of angiosperms. P. Maheshwari, ed. International Society of Plant Morphologists, University of Delhi, Delhi.
- 38. Rees, H., R. Riley, E.L. Breese, and C.N. Law, eds. 1981. The manipulation of genetic systems in plant breeding. Philos. Trans. R. Soc. London 292B:401-609.
- Riley, R., C.N. Law, and V. Chapman. 1981. The control of recombination. Philos. Trans. R. Soc. London 292:529-534.
- 40. Roggen, H.P. 1975. Stigma application of an extract from

(Brassica rape pollen L.) effects napus self-incompatibility in Brussels sprout (Brassica oleracea var. "gemmifera"). Incompatibility Newsl. 6:80-84.

- Sastri, D.C. 1984. Incompatibility in angiosperms 41. significance in crop improvement. Adv. Appl. Biol. 10:71-111.
- Sastri, D.C., and N. Mallikarjuna, 1983. Methods for 42. utilization of incompatible species: current options and their limitations. In Proceedings international workshop on cytogenetics of Arachis. (in press)
- Sastri, D.C., and J.P. Moss. 1982. Effects of growth 43. regulators on incompatible crosses in the genus Arachis L. J. Exp. Bot. 33:1293-1301.
- 44. Sastri, D.C. J.P. Moss, and M.S. Nalini. 1982. The use of in vitro methods in groundnut improvement. Pages 365-370 in Proceedings international symposium on plant cell culture in crop improvement. S.K. Sen and K.L. Giles, eds. Plenum Press, New York.
- 45. Sastri, D.C., M.S. Nalini, and J.P. Moss. 1981. Tissue culture and prospects of crop improvement in Arachis hypogaea and other oilseed crops. Pages 42-57 in Proceedings symposium on tissue culture of economically important plants in developing countries, A.N. Rao, ed. National University of Singapore, Singapore.
- Sastri, D.C., and K.R. Shivanna. 1976. Attempts to 46. overcome interspecific incompatibility in Sesamum using recognition pollen. Ann. Bot. 41:891-893.
- 47. Sastri, D.C., and K.R. Shivanna. 1976. Recognition pollen alters incompatibility in Petunia. Incompatibility News1. 7:22-24.
- Sastri, D.C., and K.R. Shivanna. 1980. Efficacy of mentor 48. pollen in overcoming intraspecific incompatibility in Petunia, Raphanus and Brassica. J. Cytol. Genet. 15:107-112.
- Shivanna, K.R. 1982. Pollen-pistil interaction and control 49. Pages 131-174 in Experimental of fertilization. embryology of vascular plants. B.M. Johri, ed. Narosa Publ House, New Delhi.
- 50. Shivanna, K.R., Y. Heslop-Harrison, and J. Heslop-Harrison. 1978. The pollen stigma interaction: bud pollination in the Cruciferae. Acta Bot. Neerl. 27:107-119.
- 51. Shivanna, K.R., B.M. Johri, and D.C. Sastri. 1979. Development and physiology of angiosperm pollen. Today and Tomorrow Printers and Publishers, New Delhi. 117 p.
- Shivanna, K.R., and D.C. Sastri. 1976. Stigma surface 52. bud prot**eins** and pollinations in Petunia: coarelative study. Incompatibility News1. 7:16-21.
- 53. Shivanna, K.R., and D.C. Sastri. 1981. Stigma surface esterase activity and stigma receptivity in some taxa characterized by wet stigma. Ann. Bot. 47:53-64. Shuzikuda, N., K. Yamamoto, and T. Nakajima. 1983. Sexual
- 54. transfer of an incomplete chromosome complement from

Nicotiana bacum L. to N. rustica. Jpn. J. Breed. 33:15-22.

- 55. Sri Ramulu, K., G.M.M. Bredemeijer, P. Dijkhuis, D. de Nettancourt, and H. Schibilla. 1979. Mentor pollen effects on gametophytic incompatibility in <u>Nicotiana</u>, <u>Oenothera</u> and <u>Lycopersicum</u>. Theor. Appl. Genet. 54:215-218.
- 56. Stebbins, G.L. 1958. The inviability, weakness, and sterility of interspecific hybrids. Adv. Genet. 9:147-215.
- 57. Stettler, R.F. 1968. Irradiated mentor pollen: its use in remote hybridization of black cottonwood. Nature (London) 219:746-747.
- 58. Taylor, N.L., R.F. Quarles, and M.K. Anderson. 1980. Methods of overcoming interspecific barriers in Trifolium. Euphytica 29:441-550.
- Thomas, M. 1981. Interspecific manipulation of chromosomes. Philos. Trans. R. Soc. London 292:519-528.
- 60. Tiara, T., and E.N. Larter. 1977. Effects of δ-amino-n Caproic acid and L-Lysine on the development of hybrid embryo of triticale (x-triticale-Secale). Can. J. Bot. 55:2330-2334.
- 61. Tiara, T., and E.N. Larter. 1977. The effects of variation in ambient temperature alone and in combination with δ -amino-n-caproic acid on development of embryos from wheat rye crosses (Triticum turgidum var. durum cv. gora x Secale cereale. Can. J. Bot. 55:2335-2337.
- 62. Tsinger, N.V., and T.P. Petrovskaya-Baranova. 1961. The pollen grain wall: a living physiologically active structure. Dokl. Akad. Nauk SSSR 138:466-496.