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Phosphate retention and extractability in soils of the humid zone in West Africa

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Abstract

Phosphate adsorption and desorption studies provide insight for developing P management strategies. Very few studies have concentrated on P desorption which provides information on the reversibility of adsorbed P. Phosphate adsorption and desorption studies were carried out on seven rice-growing soils from three countries in West Africa, with the objective of relating these processes to P management strategies for the soils. The standard P requirement (SPR) of soils, defined as the amount of P to be added to attain an equilibrium solution concentration of 0.2 mg P 1^{-1} , varied from 42 to 175 mg P kg⁻¹. Out of the seven soils, four required low amounts and the other three needed relatively higher P applications. The extractability of the adsorbed P in different extractants was: 0.01 M KCl 32–78%, Olsen P 21–112%, and Bray 1 P 15–40%. Differences in the amounts of P desorbed by the soils suggested that the critical P levels needed for P management must be different. Management options to increase P availability in the soils are suggested. © 2001 Elsevier Science B.V. All rights reserved.

Keywords: Phosphorus; Adsorption; Desorption; Extractability of P; Soils; Humid zone

1. Introduction

The rain-fed lowland and upland ecosystems in West Africa are very important for rice production. Approximately 40% of the lowland and about 70% of the upland rice is grown in the humid zone of West Africa. Phosphorus

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deficiency has been identified as one of the major limiting factors to crop production in highly weathered, low-activity, Ultisols and Oxisols (Sanchez and Salinas, 1981; Warren, 1992). Such soils are low in P and the applied P is made unavailable by reactions with iron oxides (Juo and Fox, 1977; Mokwunye et al., 1986).

It is important to understand the behaviour of phosphate, especially its adsorption and desorption by soils, to supplement the information already available on P response to direct and residual P (Sahrawat et al., 1995, 1997a), and calibration of soil and plant tests for developing P management strategies for rice-based production systems (Sahrawat et al., 1997b, 1998).While studies have been made on P adsorption of West African soils (Juo and Fox, 1977; Owusu-Bennoah and Acquaye, 1989; Owusu-Bennoah et al., 1997), little attention has been paid to desorption of the adsorbed P by acid soils used for crop cultivation in West Africa.

This study was undertaken to evaluate the phosphate adsorption-desorption characteristics of seven representative rice-growing soils in West Africa and to relate these with soil properties that can serve as indices for routine characterization of phosphate adsorption capacity of the soils.

2. Materials and methods

2.1. Soils

Surface (0-0.15 m) samples of seven highly weathered soils were selected (Narteh and Sahrawat, 1999) from typical rice-growing areas in Nigeria, Ghana and Côte d'Ivoire (Table 1). Soils 1, 2, 3, 4 and 5 were sampled from the lowlands. Soils 6 and 7 were from uplands and were included for comparison. The soils were air-dried and ground to pass through a 2-mm screen.

List of the soils used						
Soil no.	Location	Classification FAO (1988)	Vegetation			
1	Edozhighi, Nigeria (9°06' N, 5°59' E)	Dystric Gleysol	Derived savanna			
2	Ifaki–Ekiti, Nigeria (7°14′N, 5°08′E)	Eutric Gleysol	Secondary forest			
3	Itokin, Nigeria (6°36'N, 3°32'E)	Dystric Gleysol	Secondary forest			
4	Fumesua, Ghana (6°54'N, 1°35'W)	Gleyic Arenosol	Secondary forest			
5	Kikam, Ghana (4°53'N, 2°14'W)	Dystric Gleysol	Secondary forest			
6	Bouaké, Côte d'Ivoire (7°42'N, 5°00'W)	Eutric Leptosol	Forest–Savanna transition			
7	Man, Côte d'Ivoire (7°31'N, 7°37'W)	Ferric Acrisol	Secondary forest			

Table 1

2.2. Soil analysis

Soil analysis was done as described in Narteh and Sahrawat (1999). Particle size distribution was determined by the pipette method. Soil pH was measured in water (1:2.5), 1 M KCl (1:2.5), 1 M NaF (pH 8.0) (1:40, Bolland et al., 1996) and 0.5 M NaHCO₃ (pH 8.5) (1:2.5). The rationale for determining soil pH in 1 M NaF and 0.5 M NaHCO₃ is to predict phosphate sorption capacity of soils from the measured pH. Total P of the soils was determined by digestion with perchloric acid. Available P was determined using 0.03 N NH₄F + 0.025 N HCl (Bray 1) at 1:7 soil/solution ratio (Olsen and Sommers, 1982). Dithionite–citrate–bicarbonate extractable Fe (Fe_d) and Al (Al_d) were determined as described by Mehra and Jackson (1960) and oxalate extractable Fe (Fe_o) and Al (Al_o) as described by McKeague (1978). Iron and Al in the extracts were determined by atomic absorption spectrophotometry.

2.3. Phosphorus adsorption

Phosphorus adsorption data were obtained following Fox and Kamprath (1970). Soil samples (2 g) were equilibrated in 50-ml centrifuge tubes with 20 ml of 0.01 M KCl containing 0–400 mg P kg⁻¹ soil as KH_2PO_4 . The weights for the tubes containing the 400 mg P kg⁻¹ were recorded for subsequent desorption studies. A few drops of toluene were added to suppress microbial activity during incubation and the soil samples were incubated at 25°C for 6 days. The suspensions were shaken for 30 min, twice daily on a mechanical shaker. After equilibration, the samples were centrifuged for 10 min and filtered through Whatman no. 1 filter paper. The P content in the clear solution was determined by the molybdenum blue method (Olsen and Sommers, 1982). Since there were no significant differences between the duplicates, the average value of the duplicate analysis was used to calculate the amount of P in solution. The difference in the amount of P added and that recovered in solution was considered as P retained by the soil.

A linear form of the Langmuir equation C/X = C/b + 1/kb was used to evaluate the adsorption data. In the equation, *C* is equilibrium P concentration (mg P 1⁻¹), *X* is the amount of P sorbed (mg P kg⁻¹), *b* is the Langmuir adsorption maximum, and *k* is a constant related to the binding energy. The *b* was determined from the slope of the plot of C/X against *C* and *k* was calculated from the intercept. Phosphate adsorption buffer capacity (PBC) was estimated from the P adsorption curves as the slope of the regression equation relating P adsorbed to log P concentration. The amount of P required by the soils to attain a concentration of 0.2 mg P 1⁻¹ after 6 days (standard P requirement (SPR)) was extrapolated from a graph of P adsorbed against equilibrium P concentration.

2.4. Phosphorus release

After separating the P in solution, samples to which 400 mg P kg⁻¹ soil was added were subjected to a desorption sequence to determine the P release. To correct for the entrained solution left in the tubes after filtration, the tubes were weighed and sufficient amount of 0.01 M KCl was added to bring the volume of solution back to 20 ml. The suspensions were then shaken for 2 h on a mechanical shaker (Tiessen et al., 1993), centrifuged and filtered through Whatman no. 1 filter paper. The P desorbed was determined as previously described. The desorption was conducted eight successive times and each time the released P was measured.

Another set of the soils held in centrifuge tubes was incubated for 1 week with P added at a rate of 40 mg P kg⁻¹. Phosphate desorption cycles were carried out successively for two times using Bray 1 and Olsen extractants. A blank without P but only the extractants added was included in the desorption experiment.

3. Results and discussion

3.1. Soils

The texture and some chemical characteristics of the soils are summarized in Table 2. The data on particle size distribution, total P, Bray 1 P, organic C and DTPA extractable iron are from Narteh and Sahrawat (1999). Soil pH in various solutions and extractable Fe and Al fractions were determined in this study. Apart from soil 2, the soils are acid, low in organic C and available P. The clay content was low to moderate. The Fe_d values ranged from 1.8 to 28.6 g kg⁻¹ and the Fe_o/Fe_d ratio was ≥ 0.39 except in two soils, indicating that most of the soils have a high percentage of low-crystalline Fe oxides capable of immobilising P. In soil 2 situated at the bottom of a toposequence, the clay and silt content amounted to 962 g kg⁻¹. It is a redoximorphic soil with high pH due to reduction. In addition to its high pH value, the soil had high Fe_d (28.6 g kg⁻¹) and Fe_o (11.4 g kg⁻¹) values. As was expected from its high pH, the soil also had the highest Ca content (14.8 cmol (+) kg⁻¹) among the studied soils and a CEC of 30 cmol (+) kg⁻¹ (Narteh and Sahrawat, 1999).

3.2. Phosphorus adsorption

The P adsorption data for all the soils fitted well to the Langmuir equation $(r^2 > 0.972)$. The P adsorbed against equilibrium P concentration graphs for soils 2 and 3 failed to show a maximum (equivalent to a plateau) at the highest

Table 2	
Texture and chemical characteristics of the soils	

Soil no.	pН				Sand	Silt	Clay	Total P	Bray 1	Org. C	Fe-DTPA	Fed	Feo	$\mathrm{Fe_o}/\mathrm{Fe_d}$
	$\overline{H_2O}$	KCl	NaF	NaHCO ₃	$(g kg^{-1})$	$(g kg^{-1})$	$(g kg^{-1})$	$(mg kg^{-1})$	$(mg kg^{-1})$	$(g kg^{-1})$	$(g kg^{-1})$	$(g kg^{-1})$	$(g kg^{-1})$	
1	4.3	3.6	8.8	8.8	333	504	163	162	8	7.8	0.34	4.25	3.18	0.75
2	7.7	6.9	10.0	8.6	38	452	510	1125	3	46.0	0.18	28.7	11.4	0.40
3	4.9	3.9	9.8	8.7	419	366	215	163	8	9.8	0.18	8.80	3.40	0.39
4	5.4	4.2	8.8	8.8	813	142	45	89	12	8.8	0.11	3.88	3.56	0.92
5	5.1	4.0	9.0	8.7	160	490	350	448	5	35.2	0.26	1.85	1.10	0.59
6	6.3	5.0	8.7	8.8	684	228	88	211	10	7.4	0.02	9.63	0.93	0.10
7	5.0	4.1	9.3	8.6	408	294	298	234	6	15.6	0.17	9.53	1.53	0.16

Source: Partly from Narteh and Sahrawat (1999).



Fig. 1. Phosphorus sorption curves showing the relationship between P adsorbed and P in solution for seven West African soils (1 to 7).

equilibrium P concentration and therefore the calculated Langmuir b were greater than 400 mg P kg⁻¹ (Fig. 1 and Table 3). The Langmuir b was highest for soil 3 and lowest for soil 14 and was significantly related to Fe_d and Fe_o. The simple linear correlation coefficients between Langmuir P sorption maximum, percentage of P recovery after eight desorptions and soil characteristics are shown in Table 4. The Langmuir P sorption maximum was significantly related to Fe_d, Fe_o, Al_d and to pH determined in 1 M NaF. According to Owusu-Bennoah and Acquaye (1989) Al is important for P sorption in many acid soil of Ghana. There was no significant relationship between P sorption and pH in water or KCl, clay or organic C.

Soil no.	Langmuir		PBC ($l kg^{-1}$)	Amount of P adsorbed		
	$\frac{P \text{ maximum } (b)}{(\text{mg } P \text{ kg}^{-1})}$	Affinity index (k) $(l mg^{-1})$		(mg kg ^{-1}) at 0.2 mg P l ^{-1} of solution (SPR)		
1	296	0.97	18.5	50		
2	567	1.72	114	80		
3	449	4.86	85.4	175		
4	277	0.56	14.7	45		
5	210	0.63	7.36	44		
6	207	0.67	8.10	42		
7	369	1.96	54.5	115		

Table 3Phosphorus sorption parameters of the soils

Table 4

Soil property	Langmuir P maximum	Recovery (%) of added
	C C	P in eight desorptions
pH, H ₂ O	0.473	0.415
pH, KCl	0.586	0.295
pH, NaF	0.935 * *	-0.286
pH, NaHCO ₃	0.694	0.349
Clay	0.640	-0.440
Organic C	0.454	-0.367
DTPA-Fe	0.075	-0.855*
Fe _d	0.819 *	0.166
Fe	0.832 *	-0.140
Ald	0.847 *	0.189
Al	0.726	-0.147

Correlation coefficients (r) between Langmuir P sorption maximum, desorption and selected soil characteristics of the seven soils

* Significant at 5% levels, respectively.

* * Significant at 1% levels, respectively.

The following multiple regression equations are the regressions for Fe_d and Fe_o separately although it should be noted that Fe_d may contain Fe_o :

$$b = 221 + 5.68 \text{Fe}_{d} + 18.03 \text{Fe}_{o}$$
 $r^{2} = 0.728$
 $b = -102 + 61.4 \text{pH} + 1.31 \text{clay} - 11.3 \text{OC}$ $r^{2} = 0.667$

and indicate that iron oxides accounted for 73% of the variability in P adsorption whereas pH, clay and organic C together accounted for only 67%. The amounts of P required to attain an equilibrium concentration of 0.2 mg 1^{-1} (SPR) varied between 42 and 175 mg kg $-^1$ (Table 3). The SPR of soils 2, 3 and 7 were higher than reported for some soils in West Africa (Adetunji, 1997; Mokwunye et al., 1986), but the second group of soils had SPR within the range for most West African soils. As expected, the soils with relatively high SPR also had high Langmuir k, consequently there was a close relationship between them (Fig. 2).

The P buffering capacity, PBC (Holford, 1979), was highest for soils 2, 3 and 7 (Table 3).

3.3. Phosphorus desorption

The soils retained between 206 (soil 6) and 375 mg P kg⁻¹ soil (soil 2) (Fig. 3). The P released during eight successive 2-h extractions with 0.01 M KCl varied from 68 mg kg⁻¹ (32%) in soil 5 to 160 mg kg⁻¹ (78%) in soil 6. This suggests that when P is applied to soil 6, it is more easily available to the rice



Fig. 2. Relationship between Standard Phosphorus Requirement (SPR) and the Langmuir affinity index (k) for seven West African soils.

crop than that of soil 5. During extraction, the amount of P remaining retained at a given equilibrium P concentration in solution was higher than during adsorption in soils 1, 4 and 5, indicating a low extractability of the bound P in these soils. Because these soils have low PBC or are poorly buffered, hysteresis was pronounced in them (Fig. 3 and Table 3). The opposite was true for soils 2 and 3, in which more P was in solution during desorption than during adsorption. This is consistent with the proposition that these soils have high PBC and can maintain solution concentration for a longer time than in the rest of the soils. The results of the desorption study, therefore, suggest that the critical limit for P in soil solution in the studied soils may be different and P management strategies for rice in these West African soils will depend on the soil type.

Except for soil 2, the greatest release of P from each sample into 0.01 M KCl was at the first step and decreased after the initial extractions, in conformity with the results of Dev et al. (1990). Despite low release, at the end of the eight successive 2-h extractions with KCl, the solution P maintained by all soils $(0.33-1.72 \text{ mg P } 1^{-1})$ was greater than the critical level of 0.2 mg P 1^{-1} . As judged from its extractability in KCl, these soils can well support rice production. For example, in a field study on P response of upland rice on soil 7, it was shown that soil solution P concentration of 0.05 mg 1^{-1} (equivalent to about 60 kg P ha⁻¹) was adequate for optimum yield (Sahrawat et al., 1995). According to this study, the soil factor that controlled release of the added P in the seven soils studied was DTPA-extractable Fe (Table 4), which was negatively and significantly correlated with the percent P recovery (r = -0.855).

After 1 week of incubation of the soils with 40 mg P kg⁻¹, the percentage of Olsen-P was consistently greater than Bray 1-P in all soils (Table 5) and was positively related to soil pH ($r = 0.766^*$). This was evident in soil 2 (pH 7.7), in which P released was > 40 mg P kg⁻¹ and thus some native P was also



Fig. 3. Phosphorus adsorption and desorption isotherms for the seven soils. The sorption curves represent a standard sorption isotherm. The seven points along each desorption curve represent a successive step in the repeated extraction.



extracted. The relationship between pH and percentage P extracted by the Olsen reagent may be explained by the high HCO_3^- concentration. The high pH of Olsen's reagent increases the negative charge of the soil, thereby competing with phosphate ions at ligand exchange sites, resulting in release of P from soils (Hingston et al., 1972).

Table 5

Soil no.	Extraction 1 (mg kg ^{-1})	Extraction 2 $(mg kg^{-1})$	$\frac{\text{Sum}}{(\text{mg kg}^{-1})}$	P recovery in two extractions (%)
Bray 1				
1	9.47	5.27	15.2	38
2	4.94	4.20	9.14	23
3	5.10	3.85	8.95	22
4	8.47	2.87	11.8	28
5	5.86	1.88	7.74	19
6	14.70	1.19	15.9	40
7	3.41	2.74	6.15	15
Olsen				
1	21.00	6.10	27.1	68
2	28.80	15.8	44.6	112
3	6.43	1.79	8.22	21
4	16.80	4.98	21.8	55
5	12.60	3.54	16.1	41
6	24.30	4.60	28.9	72
7	5.86	3.56	9.42	24

Bray 1 and Olsen extractable P in the soils after 1 week of incubation with 40 mg P kg⁻¹ soil. Values reported are net amounts of P extracted from soils

In contrast, there was no relationship between percentage P extracted by Bray 1 and pH (r = 0.009), indicating that pH was not the main factor controlling the extraction by the acid reagent. Also, the Olsen reagent extracted more P (21–112%) than the Bray 1 (15–40%) at the end of two successive extractions.

3.4. Relationship between pH in various salts and phosphorus retention by soils

The pH in 1 M NaF was between 8.7 and 10.0, showing that F^- replaces OH⁻ from soil minerals (Perrot et al., 1976a; Bolland et al., 1996). In 0.5 M NaHCO₃, soil pH was fairly constant at 8.6–8.8 (Table 2). The pH in NaF correlated positively with Langmuir b ($r = 0.935^{**}$) and Fe_d ($r = 0.819^{*}$) (Table 4) indicating that both F⁻ and phosphate adsorption related to surface OH⁻ groups mainly of Fe oxides. Rapid laboratory and field tests of soil pH in NaF have been used to detect allophanic and aluminium-rich soils (Perrot et al., 1976b). These results are in agreement with those reported by Bolland et al. (1996) for Australian soils and the test merits further evaluation in soils of the humid forest zone of West Africa.

4. Conclusions

Based on the study, two categories of soils are presented: soils 1, 4, 5 and 6 are low P-adsorbing soils and require low rates of P additions whereas soils 2, 3

and 7 will need relatively large amounts of P. In the second group of soils, the key process in their P management is the amount of P released and made available for plant uptake.

Soil 2 with the highest P adsorption capacity had all the adsorbed P released within two extractions by Olsen reagent which suggests that the critical P level of such a soil will be low despite its high P adsorption capacity. The residual P effects may be stronger on this soil. On the other hand, our experience with soil 7 shows that residual P effects of applied P in the field decreased tremendously after the first year of application. Hence, frequent, small rates of P may be beneficial for rice farmers on soil 7. Addition of organic matter or leaving crop residues in the field to decompose may reduce P adsorption and increase P availability (Easterwood and Sartain, 1990). Breeding for rice varieties that are adapted to low soil P conditions is currently being pursued by the West Africa Rice Development Association (WARDA). The use of such cultivars along with P management practices is seen as promising for the sustainable increase in rice productivity in the humid zone of West Africa (Sahrawat et al., 2000).

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