Characterization of Downy Mildew Resistance in Pearl Millet

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Abstract

Sixty-one pearl millet (Pennisetum glaucum) genotypes from diverse geographical origins identified as

downy mildew (Sclerospora graminicola) resistant in several field tests at IAC and other locations in India and Africa were evaluated against five host-specific pathotypes and a field population of S. graminicola, referred to as pathotype 6. Pot-grown seedlings of all the pearl millet genotypes, were spray-inoculated with a sporangial suspension (1×10^5 sporangia ml⁻¹) of each pathotype in a replicated experiment in a glasshouse. Observations were recorded for latent period and disease incidence. Host resistance index (HRI) is suggested as a measure of resistance stability, and was calculated as $[1+(ab^{-1})]^{-1}$, where a= disease incidence (%), and b= latent period (day). There were highly significant (P<0.001) effects of host genotype, pathotype, and their interaction on HRI. A number of genotypes were disease free to different pathotypes; IP 18292 was disease free from all six pathotypes. An additional 15 genotypes showed < 10% incidence to all six pathotypes. These genotypes with HRI values of 0.69 to 0.94 and are the best available sources of resistance. Eight genotypes (IP 5272-1, IP 18296, IP 18297, P 536-2, P 1564, P 2895-3, P 3281-1, and 700481-21-8) that

showed differential reactions to six pathotypes can be utilized for characterizing S. graminicola isolates.

विभिन्न भौगोलिक मूल से एकत्रित अन्तर्राष्ट्रीय एशिया केन्द्र एवं भारत के दूसरे स्थानों पर अनेक क्षेत्र परीक्षणों में तुलासिता (स्के. ग्रेमिनिकोला) से रोनरोधी वाजरे के 61 समित्रिकों को पाँछ पोषिता विशेष रोन प्राल्प व एक क्षेत्र जीव संख्या रोन प्राल्प 6 के दिपरीत मूल्यांकित किया गया। नमलों में उनाये गये वाजरे के तब समित्रिकों को काँच घर में एक अभ्यावृत्ति प्रयोग में प्रत्येक रोन प्रत्य के बीजाणुधानी घोल (1 x 10⁵ बीजाणुधानी मिली.-1) से फुझरन संक्रमित किये गये। रोन आपात एवं गुप्तावधि के वर्णन अभिलिखित किये गये। पोषिता रोधिता धातांक रोधिता त्यायित्व के मापन के लिये काम में लिया गया जिसका फलन [1+(ab-1)]-1, a = रोन आपात (%) व b = गुप्तावधि (दिन), द्वारा किया गया। पोषिता रोधिता धातांक पर पोषिता समिपित्रैक, रोन प्रत्य एवं उनकी निथ क्रिया के उच सार्थक (P<0.001) प्रभाव मिले। प्रत्येक समिपित्रैक विभिन्न रोन प्रत्यों से रोन मुक्त रहा। एक अतिरिक्त समिपित्रैक 15 वे सब रोन प्रत्य पर से आपात 10% से अधिक दिखलाया। ये समिपित्रैक 0.69 - 0.94 एच आर आई के साथ रोधिता के सबसे अच्छे लोत रहे। आठ तमिपित्रैक (आई.पी. 5272-1, आई पी 18296, आई पी 18297, पी 535-2, पी 1564, पी 2895-3 पी 3281-1 एवं 700481-21-8) ने सब रोन प्रत्यों से भिन्नक प्रतिक्रिया दर्शायी, को ग्रेमिनिकोला के अभिलक्षण के लिये काम में लिये जा सकते हैं।

Downy mildew of pearl millet [Pennisetum glaucum (L.) R. Br.], caused by Sclerospora

graminicola (Sacc.) Schröt, is a highly destructive and widespread disease in Africa and Asia (Rachie

and Majmudar, 1980; Williams, 1983). Genetic resistance is the most effective and economic means of managing this disease. Therefore, breeding for downy mildew resistance continues to be a major component of pearl millet improvement programs. Genetically homogeneous F1 hybrids, based on cytoplasmic male-sterile lines, are more susceptible to the disease than the heterogeneous open-pollinated varieties (Safeeulla, 1977). During the past 20 years, three commercial F1 hybrids HB 3, BJ 104 and MBH 110, have succumbed to downy mildew within 5-7 years of wide-scale cultivation (AICPMIP, 1992).

Major downy mildew resistance genes have been identified and incorporated in the commercial hybrid parents, and hybrids (ICMH 451 and Pusa 23) made from such parents are under cultivation in India (Singh et al., 1993). However, the resistance durability of these hybrids may be short-lived because of genetic uniformity of F1 hybrids and high level of genetic variability in the pathogen populations. A number of downy mildew resistant lines have been identified from germplasm and breeding materials at ICRISAT Asia Center (IAC) and other locations in India and Africa (Singh, 1990, 1995). Resistance in several of these lines has been variable at different locations indicating the existence of variable populations of the pathogen (ICRISAT, 1989).

In India, at least three distinct populations of the pathogen (referred to as pathotypes) specific to HB 3, MBH 110 and 852 B are known (R. P. Thakur, unpublished). Recently, at IAC Patancheru, we have established five pathotypes from a field population of S. graminicola by selection through asexual generations, on seedlings of five pearl millet hybrids and inbred lines (Thakur and Rao, 1993).

In the present study, 61 pearl millet genotypes that showed downy mildew resistance in multilocational tests in India and Africa (Singh, 1990), were evaluated against six pathotypes, including path 6 (a field isolate as a control) to determine the pathotype-specific/nonspecific resistance in these genotypes. A part of the results was presented earlier (Thakur et al., 1995).

Materials and Methods

Downy mildew resistant pearl millet lines. Sixtyone pearl millet genotypes, identified as resistant to
downy mildew (< 10% mean incidence) in several
screenings at IAC and a number of locations in
India and western Africa over several years, were
either germplasm accessions or selections thereof
from Botswana (1), Burkina Faso (4), Central
African Republic (1), Cameroon (7), Gambia (1),
India (4), Mali (4), Niger (11), Nigeria (11),
Senegal (5), South Africa (1), Sudan (6), Togo (2),
and Zimbabwe (1). A number of these lines also had
resistance to rust in India (Singh, 1990). Original
seed of these lines were obtained from the Genetic
Resources Division, IAC.

The pathotype isolates. The five pathotypes of S. graminicola: Path 1, Path 2, Path 3, Path 4, and Path 5, selected on specific host genotype for increased virulence from field populations of the pathogen (ICRISAT, 1993), were maintained on seedlings of their respective selection host genotypes, NHB, 3, BJ 104, MBH 110, 852B and 700651, respectively, in polyacralic isolation chambers in a glasshouse. Pot-grown seedlings of each host genotype were spray-inoculated with a sporangial suspension (1 x 105 sporangia ml-1) obtained from previously inoculated seedlings. The sixth pathotype, Path 6, an isolate from the IAC field population of the pathogen from a mix of downy mildew susceptible pearl millet genotypes 7042S and NHB 3, was included as a control.

Experiment 1. The experiment with 62 host genotypes, including 7042S a downy mildew susceptible control, and six pathotypes was conducted in a completely randomized design (CRD) with three replications in a glasshouse. Potting mixture of alfisol, sand, and farmyard manure (2:1:1 by vol) was autoclaved at 121.6° C for 60 min for two cycles and filled in 15 cm dia. plastic pots. Seeds were surface sterilized with 2% NaOCl (diluted from 5.25% clorox bleach, Oakland, Ca 94623, USA) for 5 min, washed thoroughly in distilled water and dried at the laboratory temperature (22-24° C) on blotting paper sheets. Twenty-five to 30 seedlings were raised in each pot and watered everyday. Seedlings at 2-leaf stage were spray-in-

8

x 105 sporangia ml⁻¹) using a hand sprayer. Sporangia for each pathotype were obtained from its respective host seedlings and suspended in sterilized ice-cold water (4-5° C) to prevent zoospore release prior to inoculation. Inoculation was done in an inoculation chamber and the seedlings were retained in this chamber at 20° C at high relative humidity (> 95%) for 24h (Singh and Gopinath, 1985). Seedlings were then moved to a glasshouse at 25± 2° C. The experiment was divided into six sets, each with one pathotype x 62 host genotypes x three replications x two pots per

oculated with an aqueous sporangial suspension (1

replication.

Experiment 2. Fourteen genotypes that showed either clear differential reactions (incidence), or uniformly low or high incidence to the six pathotypes in all three replicates of experiment 1, were re-evaluated to confirm the results. Experimental details were similar to those described above, except that all treatments and replications were conducted at one time.

Disease assessment. Disease assessment was done for latent period (time in days from inoculation to appearance of downy mildew symptom with sporulation) and disease incidence (percentage of seedlings showing symptoms). Seedlings were examined daily, from the 4th to 15th day after inoculation for latent period. Downy mildew incidence was recorded 15 days after inoculation and per cent disease incidence was determined.

Host resistance index (HRI). In a systemic disease, like downy mildew of pearl millet, where large variation exists for disease incidence and latent period, incidence alone may not provide a representative measure of resistance in a host genotype. We suggest that 'host resistance', in such pathosystems, could be better described by HRI, which as a function of incidence and latent period is

$$HRI = [1 \div ab^{-1})]^{-1}$$
 ...(1)

defined by the equation:

where a = incidence (%) and b = latent period (days). In equation (1), HRI = 1, if a = 0 which corresponds to a completely resistant line, and HRI <1, if a > 0, which corresponds to a less than

completely resistant line. The degree of resistance will be directly proportional to the value of HRI.

Statistical analysis. Data on downy mildew incidence, latent period and HRI were subjected to analysis of variance (ANOVA) using the GENSTAT ANOVA procedure (GENSTAT, 1986) for a two-factorial (host genotype and pathotype) CRD for experiment 1 and for three factorial (experiment, host genotype and pathotype) CRD for experiment 2. The error mean squares for the data sets from the two experiments for 14 genotypes were examined for homogeneity using F-test of significance before pooling them for analysis. A correlation analysis was run to determine the relationships among incidence, latent period, and HRI. Host genotypes were grouped, based on

centroid clustering analysis (SAS, 1985) of HRI

Results

values.

Downy mildew incidence. The mean and range of downy mildew incidence of 61 genotypes for individual pathotypes varied from 9 to 20% and from 0 to 86%, respectively (Table 1). The number of genotypes in each of seven incidence classes was quite variable for individual pathotypes. The number of genotypes with 0 incidence varied from 3 for Path 6 to 17 for Path 5, and there were 13 genotypes each for Path 1 and Path 4, 16 for Path 3, and only 5 for Path 2. However, with the > 50% incidence class, the number of genotypes varied from 1 to 5 for the six pathotypes.

Of the 61 genotypes, IP 18292 showed 0 incidence and 7042S between 72-97% to all six pathotypes (Table 2). There were 18 other genotypes that had between 0 and 10% mean incidence to any one of the six pathotypes. Highly significant (P < 0.001) effects of host genotypes, pathotypes, and their interaction on incidence were observed (Table 3).

Latent period. The latent period was quite variable for different host genotype x pathotype combinations (Table 4). The mean latent period across six pathotypes varied from 5.6 d for a highly susceptible genotype 7042S to 11.5 d for a highly resistant genotype IP 18293. Genotype P 24-1 with a mean

Table 1. Number of pearl millet genotypes in different downy mildew incidence (DMI) classes based on reaction to

	able 1. Number of Sclerospora graminicola DMI (%) class									
• - •		1-5	6-10	11-20	21-30	31–50	> 50	Mean	Range	
Pathotype					-		2	13	0-82	
Path I	13	14	15	4		6	2	. 13	0-77	
Path 2	5	23	7	14	3		÷	ji	0-60	
	16	18	5	11	5	I	3		0-86	
Path 3	12	24	7	11	0	3.	3	11		
Path 4	13	4-7		13	1	4	1	9	0-80	
Path 5	17	. 17	8			16	3	20	0-79	
Path 6	. 3	11	12	11	3					

Rounded to whole number.

		aminicola in a glasshouse experiment Downy mildew incidence (%) ²							
Pearl millet genotype	Origin	Path I	Path 2	Path 3	Path 4	Path 5	Path 6	Mean	HRI
D 332/1/2-2	Niger	0	1	1	I	1	3	1 -	0.91 0.68
DIC-P4-P2	_¢	1	1	2	0	39	41	7	
IP 70-L-1	India	0	0	0	2	1 -	1	1.	0.94
IP 94/1/2-1	Niger	10	14	. 2	0	0	15	7	0.71
IP 537B	Mali	16	24	. 24 .	1 7	14	31	18	0.40
IP 337B IP 1481–L–2	India	3	2	13	6	. 4	35	11	0.61
IP 1481-L-1 IP 2084-1	South Africa	1	4	4	0	8	7	· 4	0.74
	Niger	4	3	0	87	.1	56	25	0.60
IP 5272-1	•	23	32	54	19	80	79	4\$	0.16
IP 5749-1	Nigeria Cameroon	39	44	54	16	. 9	20	30	0.27
IP 6138-3	•	7	11	0	2	Ö	. 6	4	0.75
IP 6140-1	Cameroon Cameroon	48	37	60	9	9	33	33	0.28
IP 6147—4				16	4	0	4	7	0.63
IP 6240-2	Cameroon	7	- 11	6	14	15	16	10	0.53
IP 6249-4	Cameroon	5	3	4	17	32	34	24	0.28
IP 8695-1	Sudan	30	25	-	2	15	22	15	0.4
IP 8695—4	Sudan	10	15	28		13	27	13	0.63
IP 8703–1	Sudan	23	12	12	2	-			
IP 8710-1	Sudan	0	i	2	1.	0	10	2	0.9
IP 8714-1	Sudan	10	7	. 11	10	2	13	9	0.5
IP 8715÷	Sudan	32	37	4	3	10	43	22	0.4
IP 8830-2	Zimbabwe	0	i	0	16	0	17	6	0.8
IP 8876-2	Burkina Faso	82	58	16	17	4	25	34	0.3
	Burkina Faso	7	12	14	8	17	13	12	0.4
IP 8877-3	Nigeria	17	9	` 3	11	3	7	8	0.5
IP 8998–1	Nigeria Burkina Faso	26	28	13	19	35	39	27	0.2
IP 11541-1					0	r = 0	. 0	0	1.00
IP 18292	ICRISAT ^d	0	0	0			<u>`</u>		(Conti

Path 1

0

Table 2 Continued

Pearl millet

genotype

IP 18293	ICRISAT	U ·	U	•	•	•			
IP 18294	Mali	0	. 4	8	0	1 -	2 ·	3	0.89
		0	3	1	2 .	0	2	1	0.91
IP 18295	Cameroon	-	18	0	0	14	2	11	0.67
IP 18296	Nigeria	29		0	39	0	0	7	0.90
IP 18297	Mali	0	0	0	i	0	0	1	0.94
TP 18298	Burkina Faso	i	3	U			_		
P7-4	Nigeria	5	5	5	5	0 .	21	7 .	0.66
P 10-1	Nigeria	8	3	2	3	16	39	12	0.59
P 24-1	Cameroon	· 6	4	0	1	1	10	4	0.77
P 120-1	Cameroon	3	2	3	ı	3	. 7	3	0.78
P 181-2	CAR	0	3	16	7	4	15	. 8	0.74
P 536-2	Mali	13	19	ı	0	2	3 \$	12	0.59
		8	20	8 .	. 2	6	9	9-	0.60
P 1449-2	Senegal	31	11	0	\$5	0	35	27	0.52
P 1564	Senegal	71	77	16	85	34	41	54	0.15
P 1577	Senegal	19	16	21	19	12	34	20	0.30
P 1591	Senegal		3	21	8	6	35	13	0.47
P 1596	Senegal	7		21					. 0.07
P 2819-1	Niger	2	4	0	1.	0	4	. 2	0.87
P 2880	Niger	S	3	. 5	5	2	8	5	0.69
P 2895-3	Niger	0	i	12	50	0	15	13	0.65
P 2910-2	Niger	0	0	0	0	0	2	•	0.98
P 2914-3	Niger	5	7	I	0	13	10	6	0.65
P 2925-1	Niger	4	5	i	0	8	8	4	0.72
P 2947-2	Niger	10	S	6	5	2	21	9	0.63
P 2947-2 P 2950	Niger	10	9	54	31	16	64	31	0.26
	Togo	0	1	0	0	0	4	ı	0.93
P 3281-1		6	17	9	0	7	12	9	0.61
P 3346-1	Togo	2	4	3	10	0	9	5	0.73
P 8749-1	Botswana	4	6	1	2	3	2	3	0.80
RC-011-2	Gambia	4	U	•	-	-			

20

. 4

0

4

14

3

72

11

2

25

12

11

12

97

34

0

59

13

0

26

76

LSD (P < 0.05) for DM incidence for pathotype x host genotype means=13.00 and for HRI for pathotype x host genotype means=0.26

25

6

49

13

20

29

82

HRI based on the formula [1+(axb-1)]-1 where a=downy mildew incidence %; b=latent period (days).

31

3

39

7

28

27

83

Mean of 150-180 seedlings in each of three replications.

Derived from crosses involving lines from Sudan, India and USA.

Not known. CAR = Central African Republic.

Downy mildew incidence (%)2

Path 4

ı

Path 3

0

Path 2

0

Path 5

0

HRI

0.98

Path 6

. 1

Mean

26

4

36

11

15

23

83

34

7

43

18

18

42

90

0.25

0.74

0.30

0.49

0.44

0.38

0.06

Origin

ICRISAT⁴

Gambia

Nigeria

Nigeria

Nigeria

Nigeria

Nigeria

Nigeria

India

RC-011-2

SDN 503

700481-21-8

700481-22-8

700481-23-2

700512-3-2

7042S control

700251

Table 3. Analysis of variance for downy mildew incidence (DMI), latent period (LP), and host-resistance index

		M			
Source of variation	- df	DMI	df	LP	HRI
Replication	2	874.26	2	63.81	0.20
Pathotype (P)	5	2671.72	5	208.32***	1.79***
Host genotype (G)	61	4004.45***	60/61 ²	21.07	1.02
PxG	305	470.61	239/305*	5.67***	0.11
Error ·	742	66.02	454/742*	2.45	0.03
*** Significant at 0.1%	a. For HRI				· —————

Table 4. Latent period^a (in days) on 61 pearl millet genotypes induced by six pathotypes of *Sclerospora graminicola* in a glasshouse experiment

Pearl millet genotype	Path I	Path2	Path3	Path4	Path5	Path6
D 332/1/2-2	_b	6.5	12.0	14.0	11.0	6.5
DIC-P4-P2	11.0	6.0	\$.0		7.2	7.0
IP 70-L-1	_	-	- ,	11.5	\$.0	7.0
IP 94/1/2-1	8.3	7.5	10.0	-	-	7.5
IP 537B	7.7	6.5	8.3	14.0	7.3	6.2
IP 1481-L-2	6.0	9.5	9.0	11.0	7.8	6.0
IP 2084-1	9.0	11.7	9.0	~	.S.S	7.3
IP 5272-1	7.7	10.8	_	7.3	11.0	6.0
IP 5749-1	7.3	5.7	7.0	7.3	5.7	5.7
IP 6138-3	7.7	6.0	7.3	9.2	7.8	6.2
IP 6140-1	7.7	6.7	-	12.0	-	6.0
IP 6147-4	7.0	6.3	8.0	10.0	9.7	6.2
IP 6240-2	10.3	7.0	8.3	9.0	-	7.3
IP 6249-4	7.8	6.5	8.0	8.8	6.8	6.3
IP \$695–1	5.8	6.5	7.0	7.2	5.8	5.7
IP 8695-4	7.3	7.2	7.2	12.5	6.8	7.2
IP 8703-1	9.0	6.5	- 9.3	12.0	8.0	6.5
P 8710-1		8.5	14.5	14.0	-	6.5
P 8714-1	7.2	6.5	6.8	` 9.8	7.5	6.3
P 8715-4	7.5	6.2	11.3	9.5	8.3	6.3
P 8830-2	_	9.0	-	9.0	. –	7.0
P 8876-2	5.8	5.7	8.8	9.5	13.0	6.8
P 8877-3	10.2	6.7	9.3	9.8 ·	8.0	7.3
P 8998-1	7.8	6.0	9.0	8.5	7.8	7.3
P 11541-2	6.5	6.8	7.7	8.5	6.7	6.2
P 18292	_	-	` -	-	-	-
P 18293	. -	<u>-</u>	-	14.0	_	9.0
P 18294	_	9.3	12.0	·- <u>-</u>	11.0	11.0
P 18295	. <u>-</u>	7.5	8.0	11.5	- •	6.5
P 18296	7.5	9.0 •	_	_	8.7	7.0

Pearl millet genotype	PathI	Path2	Path3	Path4	PathS	Path6
P 18297	-	-	-	9.0	-	<u>-</u>
IP 18298	9.0	9.3	-	11.0	-	<u>-</u>
P7-4	7.8	9.0	8.5	10.2	· -	6.3
P 1—4 P 10−1	9.5	10.0	14.0	11.5	6.8	6.0
	7.8	6.7	-	6.0	12.5	- 7.8
P24-1	9.5	9.0	8.5	9.0	9.7	7.5
P 120-1).J	10.0	11.3	3.3	13.0	7.0
P 181-2		6.5	9.0	. <u></u>	8.0	6.2
P 536–2	7.8	7.3	9.0	7.5	7.5	8.3
P 1449-2	10.0	10.7	-	7.2	-	· 6.8
P 1564	8.8	7.2	8.2	6.7	6.0	6.5
P 1577	7.3	6.8	7.8	8.0	6.5	5.8
P 1591 -	7.3			9.0	7.7	6.2
P 1595	8.5	6.7	8.0	13.0		9.2
P 2819-1	9.5	7.5	-	10.0	8.3	8.3
P 2880	8.7	9.7	11.8	7.2	-	7.2
P 2895-3	-	11.0	9.7	1.2		8.0
P 2910-2		-	_		7.2	7.3
P 2914-3	8.3	6.8	13.0	- ,	•	6.5
P 2925-1	9.0	7.2	11.0	-	6.3	6.8
P 2947-2	8.3	7.7	9.0	11.0	10.0	5.7
P 2950	8.2	6.8	7.0	7.3	6.8	8.2
P 3281-1	_	7.0	-	-	_	8. <u>2</u> 7.5
P 3346-1	9.8	6.5	10.0	-	8.3	7.5 7.0
P 8749-1	8.0	9.0	9.0	10.3	-	
RC-011-2	9.0 -	9.0	6.0	12.0	10.3	6.5
SDN 503	6.7	6.7	7.5	8.7	\$.0	5.8
		6.3	_	11.3	\$.0	6.3
700251	10.0	6.3	\$.2	_	7.5	6.3
700481-21-8	7.8	7.0	10.5	7.5	8.3	7.0
700481-22-8	9.3	7.0 6.7	-	9.0	8.0	6.2
700481-23-2	8.2	7.5	7.5	13.5	9.0	6.8
700512-3-2	11.0			5.7	5.7	5.6
7042S Centrel	5.3	5.3	6.3	J.1		

Differential reactions. A number of host genotypes

exhibited clear and strong differential incidence to specific pathotypes. For example, IP 18297 from Mali had 39% incidence with path 4 and 0 with other pathotypes (Table 2). The differential reactions of 8 of the 12 host genotypes (IP 5272-1, IP 18296, IP 18297, P 536-2, P 1564, P 2895-3, P 3281-1 and 700481-21-8) were confirmed in ex-

periment 2 (Table 5). Genotype P 1564 showed 0%

incidence with Path 3, 2% with Path 5, 11% with

LSD (P < 0.05) for pathotype x host genotype means = 2.51 b. Data not available because of no infection. Mean of 3 replications. disease incidence of 4% had a latent period of 6 d with Path 4 and 12.5 d with Path 5; and IP 8710-1 a highly resistant genotype (2% mean incidence) had a latent period of 6.5 d with Path 6 and 14.5 d with Path 3. There were significant effects of host genotypes, pathotypes and their interaction on latent period (Table 3). A significant negative, correlation (r = -0.45 at P < 0.01, df=756) was found between latent period and downy mildew

incidence across host genotypes and pathotypes.

Path 2, 26% with Path 1, 34% with Path 6 and 77% with Path 4; IP 18297 showed 0% incidence with each Path 1, Path 2, Path 3, and Path 5, < 1% with Path 6, and 33% with Path 4. These genotypes can serve as good host differentials for pathogenic variability studies. ANOVA indicated strong and significant (P < 0.001) effects of host genotype, pathotype and their interaction on downy mildew incidence, latent period and HRI (Table 6).

Table 5. Mean per cent downy mildew incidence (DMI) and host resistance index (HRI)a in 14 pearl millet pes caused by six pathotypes of Sclerospora graminicola

genotypes car				th2		:h3	Paih4		Pa	Path5		th6
Pearl millet genotype DN		thl .				HRI	DMI	HRI	DMI	HRI	DMI	HRI
	DNU.	HRI	DMI	HRI	DMI				1	0.93	42	0.14
IP 5272-1	4	0.68	4	0.69	0	1.00	78	0.03	-	0.08	72	0.08
IP 5749-1	21	0.26 -	25	0.21	50	0.12	17	0.30	72		3	0.15
IP 8715-4	29	0.21	34	0.17	4	0.71	4	0.72	19	0.29	_	0.13
IP 8876-2	66	0.09	- 53	0.11	17	0.31	17	0.33	. 7	0.57	26	
	28	0.21	. 19	0.35	0	1.00	1	0.83	13	0.41	2	0.84
IP 18296	0	. 1.00	Ò	1,00	0	1.00	- 33 .	0.21	0	1.00	< 1	0.96
IP 1\$297			4			0.63	0	1.00	7	0.51	34	0.17
P 536-2	9	0.48	21	0.25	4		77	0.08	2	0.79	34	0.17
P 1564	26	0.22	11	0.47	0	1.00	"				4	0.16
	67	0.11	67	0.10	15	0.35	79	0.0	5	0.11		0.16
P 1577	0.	1.00	<1	0.97	30	0.21	45	0.13	< 1	0.97	24	0.24
P 2895-3	0	1.00	2	0.81	0	1.00	0	1.60	< 1	0.97	- 7	0.55
P 3281-1		0.17	44	0.14	52	0.13	0	1.00	34	0.18	48	0.12
700481-21-8	35	0.17	-	U.1 .				•				
Controls							^	1.00	0	1.00	0	1.00
IP 18292	0	1.00	.0	1.00	0	1.00	0		89	0.06	93	0.06
7042S	81	0.07	78	0.07	84	0.07	71	0.68				
LSD (P < 0.05) for		DMI for	P x G means	=	11.37							
Lib (1 < 0.03	,			x G means	=	0.10				•		

Mean of 2 experimental runs.

Source of	MS										
variation	df	DMI	df	LP	HRI						
· · · · · · · · · · · · · · · · · · ·	2	247.0	2	31.82	0.042						
Replication	1	334.1	1	. 13.73	0.094						
Experiment (E)	. 2	1388.8	2	19.39	0.075						
Error	_	18300.4	. 12	40.48	1.267						
Host genotype(G)	13		12	8.46	0.053						
ExG	13-	159.0	1. 	28.84	0.666**						
Pathotype (P)	, 5	2476.0	3	27.88	0.093						
ExP	5	383.2	5 -	. 21.00	0.218						
PxG	65	2135.7	47	9.18							
ExGxP	65	` 155.3	41	2.43	0.015						
24041	332	100.9	207	1.57	0.00\$						

^{*} and *** Significant at 5, 1 and 0.1%, respectively

rusa :

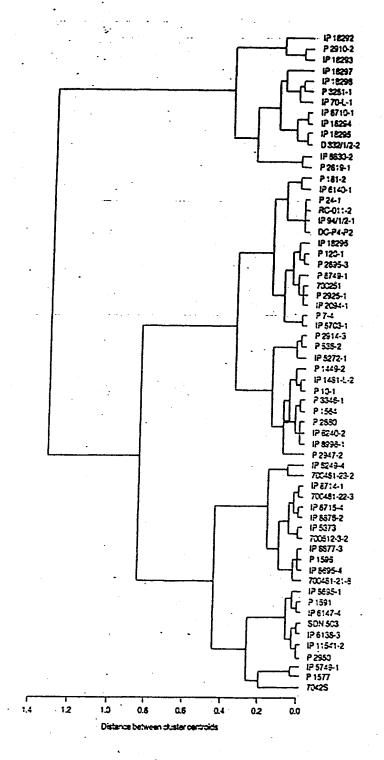


Figure 1. Classification of 62 pearl millet genotypes into distinct groups based on cluster analysis of HRI values across six pathotypes of Sclerospora graminicola. The cluster indicates five groups from highly resistant to highly susceptible genotypes.

Host resistance index (HRI). Large and significant variations were observed for HRI among host genotype x pathotype combinations. We considered the mean HRI value across all pathotypes to indicate the likely resistance stability of a line. A highly resistant genotype IP 18292 with no infection had HRI value 1.0 and a highly susceptible 7042S with 83% mean incidence had HRI of 0.06 (Table 1). Highly significant (P < 0.001) effects of host genotype, pathotype and their interaction were observed on HRI (Table 3). The cluster analysis provided useful groupings of pearl millet genotypes from a highly resistant to highly susceptible level and indicated the possible evolution of susceptibility/resistance in the gene pool (Fig. 1).

We have introduced a new concept of 'HRI' for

Discussion

measuring resistance in a systemic disease, such as downy mildew of pearl millet where both the host and the pathogen are highly variable. The two components of HRI -- disease incidence and latent period, measured in this study, are considered important for characterizing resistance of a host genotype resistance to different populations of a pathogen. Of the 61 known resistant genotypes, only 19 were uniformly resistant (< 10% incidence) to all the six pathotypes, and the remaining 42 had variable reactions. We consider that host genotypes that show higher HRI values across a number of pathotypes or locations (environments) over time would probably be more stable than those with lower HRI values. Sixteen of the 61 host genotypes that had < 10% downy mildew incidence with all six pathotypes, also had HRI values > 0.69 and these are likely to be more stable than others. It is interesting to note that host genotypes that have shown high HRI values have their origin in western Africa - the primary center of origin of pearl millet, conforming the hypothesis of existence of greatest diversity in the center of origin. The results provide ample evidence that resistance in the other 45 genotypes may not be useful in different parts of country and might be easily overcome by virulence factors in the pathogen populations, if utilized as

resistance source. It is pertinent to mention here that pathogen isolates tested in this study represent only a fraction of the total likely variability in S. graminicola populations in India, and therefore a wider testing involving more isolates will be required to account for the entire range of variability. Nevertheless, this study provided a vital indication of the likely variability in the pathogen population to serve as a guideline for further investigations.

Highly significant negative correlations between incidence and latent period provide a reasonable basis of understanding resistance stability in the source lines. However, the lower disease incidence alone cannot be taken as a true measure of resistance in a highly variable pearl millet-downy mildew pathosystem. A cultivar having fewer infected plants with shorter latent period could contribute more towards disease spread in a crop than those having a greater number of infected plants with longer latent period. Our earlier studies (Thakur et al., 1992) indicated a negative relationship between latent period and sporulation intensity. The results also provide useful information on differential reactions of some genotypes to the six pathotypes. However, these genotypes need to be made genetically homozygous for downy mildew reaction before they can be used as true differentials. These results and evidence (Gill et al., 1978; Jones et al., 1995) that resistance to downy mildew in pearl millet is governed by major dominant genes, form a good basis of future research in developing genotypes with different resistance genes and their strategic utilization in breeding programs to develop cultivars with stable resistance to downy mildew. Pearl millet genotypes, such as IP 18292 and IP 18293 are the best source of resistance available which could provide stable resistance to downy mildew in India.

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