# Research on the aflatoxin problem in groundnut at ICRISAT\*

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Received 10 July 1983 Revised December 1983

Key words Aflatoxin Aspergillus flavus Groundnut Seed colonization Seed invasion

Summary Aflatoxin contamination of groundnut is a serious problem in most groundnut producing countries and as such is given high research priority by the Groundnut Improvement Program of ICRISAT Since 1979 we have concentrated on selecting cultivars resistant to seed invasion and colonization by toxigenic Aspergillus flavus, and/or to aflatoxin production following invasion by the fungus. Resistance to invasion and colonization by A. flavus of rehydrated, mature seed has been found, and confirmed, in some cultivars. We have also screened several groundnut cultivars for seed resistance in the field, both under natural conditions and with the inoculum of the fungus added to the soil in the pod zone. Some cultivars with resistance to seed colonization also showed resistance to seed invasion by A. flavus. None of the cultivars tested has shown complete resistance to aflatoxin production but significant cultivar differences occurred in the amounts of aflatoxin produced in seeds inoculated with a toxigenic strain of A. flavus.

## Introduction

Invasion of seed by toxigenic strains of A. flavus and consequent aflatoxin contamination is a continuing serious problem in most countries where groundnuts are grown. Farmers of the semi-arid tropics have apparently failed to adopt agronomic measures to minimize aflatoxin contamination. We have therefore concentrated on the utilization of genetic resistance of seed to invasion by the toxigenic A. flavus and production of aflatoxin. Since 1979, a research program has been pursued at ICRISAT to select cultivars resistant to seed invasion and colonization by toxigenic A. flavus and to aflatoxin production. This paper reports on screening of germplasm for (a) resistance to in vitro seed colonization by A. flavus (b) resistance to invasion of seeds by A. flavus in the field, both under natural conditions and with inoculum of the fungus added to the soil in the pod zone, and (c) resistance to aflatoxin production following invasion and colonization by a toxigenic strain of the fungus

<sup>\*</sup> ICRISAT Journal Article No. JA-316

#### Materials and methods

#### Source of wed

Seeds of all cultivars/lines tested were obtained from rains and subsequently post rains seasons crops grown in alfisols at ICRISAT Center farm. Cultivars lines were harvested at maturity and plants arranged in inverted windrows in the field. After windrow drying for 2 days in the post rains and 3 days in the rains seasons the post were hand picked and sun-dised on mats until the seed moisture content was below 8 per cent. Dried posts were then stored in cloth bags at room temperature until required for testing.

#### Seed colonitation test

For all trials, undamaged, mature sends from the stored pods were tested for resistance to colonization by A. flavus using a modification of the method described by Mixon and Rogers<sup>13</sup>. For each trial, one lot of 20 g of seed was tested from each plot of each cultivar/line. Sends were surface-sterilized by soaking them in a 0.1% solution of mercuric chloride for 2 min followed by 2 risses in sterile distilled water and then their moisture contents were raised to 20%. The rehydrated sends were placed in sterile Petri plates and surface-inoculated with 1 ml of a conditial suspension (4 x 10° condita/ml) from 8-day-old cultures of the toxi genk A. flavus strain AF 8-3-2A. The percentage of sends colonized was recorded after 8 days of incubation at 25°C.

# Effect of soil inoculation on seed infection by A. flavus

Replicated field trials were carried out in the 1979/80 and 1980/81 post-rainy and 1980 and 1981 rainy seasons in which some cultivars/lines were examined for the effects on seed infection of applying inoculum of the toxigenic A. flavus strain AF 8-3-2A to the soil around pods at 30 days before harvest. The trials were laid out in a 3-replicated split-plot arrangement, the main phots assigned to isoculation versus control and the subplots to test cultivar reaction. Ten randomly selected plants in each replicated plot of each cultivar/line received 500 ml isoculum (15 × 10<sup>4</sup> condia/ml) of A. flavus. After completion of postharvest drying, replicate samples of 100 undamaged, mature seed were surface sterilized and plated out on Cxapek Dox Rose Bengal Streptomycin agar for isolation of A. flavus. The plates were incubated at 25°C and colonies of A. flavus growing from infected seeds were recorded after 5-7 days.

# Aflatox in production test

Aflatoxin production tests were carried out on seeds of each cultivar/line by the method described by Mehan and McDonald'. The rehydrated seeds in replicate plates had their testas damaged by acraping them with a sterile needle. The seeds were then surface-inoculated with the toxigenic strain of A. flavux as described above for the seed colonization test. After 10 days of incubation aflatoxins were extracted using the method of Pons' et al. 11 and quantitative determination made by the method of Nabney and Nesbitt 14.

### Results and discussion

Of 850 cultivars/lines screened, eight showed resistance to seed colonization by A. flavus. Three breeding lines (PI337394F, PI337409 and UF71513), reported resistant in the U.S.A.<sup>12,3</sup> were also resistant in the ICRISAT tests, and five new sources of resistance were identified in 1981. These cultivars/lines were further tested in the 1981/82 postrainy and 1982 rainy seasons and the seed resistance was confirmed (Table 1). Although percentages of seeds colonized by A. flavus were

Table 1: Groundhort seed resistance to colonization by Asporallian things in tests at ICRISAL Center, 1981-82.

	Percentage of seeds colonized					
Cultivat Tine	Postrains seas in	Rams seas it				
	1981-82	1982				
PI 3373941	180	10 1				
PI 337409	18 1	9.1				
UF 71513	15.3	8 ×				
J11	1 7 7	10.4				
Ah 7223	15.8	8.6				
Vat 27	19 4	12.4				
<b>Fa</b> ириг	18.1	12.1				
C55-437	•	17.3				
TMV.	46 9	34 4				
OG 43-4-1 <sup>b</sup>	99 2	94 7				
SE 1	1.20	1 11				
CV (%)	7 19	8 84				

Susceptible check

low in both seasons for resistant cultivars/lines, levels were generally lower in seeds from the rainy season crop than from the postrainy season crop. This finding agrees with those of previously unreported trials carried out in the 1980/81 and 1981 seasons which showed that seed colonization of cultivars/lines by A. flavus could be significantly influenced by season, crop location (field) and rate of postharvest drying<sup>10</sup>. High temperatures during postharvest drying of the irrigated postrainy season crop could damage the seed testa and contribute to increased levels of seed colonization. In all reported cases of mature dried groundnut seed resisting colonization by A. flavus the protective role of the seed testa has been emphasized4,13. The maximum advantage that can be derived from groundnut cultivars resistant to A. flavus will only occur if the seed testa is not damaged during cultivations. harvesting, curing, decortication or storage. Resistance in these cultivars/lines to seed invasion and colonization by A. flavus is likely to be of value in the event of stored groundnuts absorbing sufficient moisture to permit fungal growth. Also, it would perhaps be useful in seasons when field drying conditions were unfavourable for example. because of late rains. There is some experimental evidence<sup>17</sup> that the cultivars with seed resistance to colonization may have some preharvest field resistance to seed invasion by the fungus.

Experiments were carried out to investigate possible varietal differences with regard to pre-harvest resistance to seed infection with A. flavus in the field. The results of effects on seed infection of

b Highly susceptible check

<sup>.</sup> Not tested

Table 2. Infection with Aspergillus flavus of seeds from field dried pods of cultivars/lines following inoculation with the fungus of soil around developing pods 30 days before harvest

	Percentages of seeds infected with A flavus								
Cultivar/line	Seed from Postrainy season of 1979/80		ion cro	торs 1980/81		Rainy season crops 1980 1981			
	lnoc.	No.Inc	c. I	noc	No.Inoc	Inoc	No.Inoc.	Inoc.	No.Ino
PI 337394 F	1.6	0.3	2	.6	1.0	1.0	0.3	2.3	0.6
PI 337409	3.0	0.6	4	.0	1.6	1.6	0.3	2.6	0.6
UF 71513	2.0	0.6	2	.6	1.3	1.0	0.0	2.0	0.3
J 11	1.3	0.3	2	á.	0.3	1.3	0.6	1.6	0.3
TMV 2	6.6	1.3	7	.3	2.6	5.6	1.6	4.6	1.3
Krapovicka Strain									
#16	4.0	1.6	6	.0	2.3	4.6	2.0	•	•
EC 76446 (292)	6.0	1.3	6	.6	3.3	6.3	2.0	7.3	1.6
OG 43-4-1	9.0	1.6	7	.3	2.3	5.0	1.3	•	•
Robut 33-1	8.0	1.3	11	.3	2.3	7.0	1.3	•	•
M 13	6.3	2.0		.6	3.0	7.3	2.0	•	•
SE :									
For Inoculation Treatments		ts (	0.82	0.97		0.64	0.56		
For cultivars		(	0.68	0.	90	0.59	0.55		
C.V. (%)									
For Inoculation	Treatmen	ts 39	9.27	31.	84	27.05	26.02		
For cultivars		21	3.28	27.	88	31.41	25 72		

<sup>\*</sup> Not Tested: Inoc. = Soil Inoculation: No. Inoc. = No Soil Inoculation

applying inoculum of a toxigenic strain of A. flavus to the soil around developing pods of cultivars resistant and cultivars susceptible to seed colonization by the fungus are given in Table 2. The inoculation of soil around developing pods significantly increased seed infection with A. flavus. Cultivars/lines differed significantly for levels of A flavus infection and the cultivar times inoculation treatment interaction was significant. Natural A. flavus seed infection was also lower in cultivars/lines whose rehydrated stored seeds were resistant to colonization by the fungus than in susceptible cultivars. Further experiments on field inoculation techniques to evaluate cultivars for reaction to seed infection with A flavus and to formation of aflatoxins are in progress.

Previous research indicated varietal differences in aflatoxin production when autoclaved seeds were colonized by toxigenic strains of A flavus and A. parasiticus<sup>6, 16</sup>. Although claims of resistance to aflatoxin production were not confirmed by later research<sup>2,4</sup>, quantitative varietal differences in aflatoxin production were indicated. Of 195 cultivars tested at ICRISAT none has proved to be totally resistant to aflatoxin production. Some test data are presented in

Table 3 Affatoxin B, production in groundhut cultivars/lines following inoculation with A, flavus strain AF83-2A = 1981 rainy season

Cultivar/line	Aflatovin B <sub>1</sub> (µg/g seed				
PI 337394 F	115 1				
PI 337409	104 5				
UF 71513	110.7				
JII	122 6				
Ah 7223	117 4				
Var. 27	97.8				
Faizpur	118 8				
Monir 240-30	103.8				
TMV,	241.5				
FESR-11-P11-B2-B1	52.6				
OG 43-4-1	70.4				
SE ±	2.63				
CV (%)	4.29				

Table 3. Aflatoxin was produced in all cultivars tested, but cultivar differences in amounts of the toxin produced were found. Some of the cultivars with good resistance to colonization by the fungus proved good substrates for aflatoxin production, while some others that were highly susceptible to fungal colonization were not as good substrates for the production of the toxin. There was no correlation between resistance to seed colonization by A. flavus and the ability of seed to support aflatoxin production. The cultivar FESR-11-P11-B2-B1 which was highly susceptible to seed colonization by A. flavus, had the lowest level of toxin produced. We have tested only a small proportion of the world groundnut germplasm collection and it is hoped that we may find cultivars with even higher levels of resistance to aflatoxin production with good resistance to seed invasion by the fungus.

#### References

- Anonymous 1977 Report of the joint FAO/WHO/UNEP Conference on Mycotoxins, Nairobi, 19 27 September, 1977.
- 2 Aujia S S, Chohan J S and Mehan V K 1978 The accomming of peanut varieties for the accumulation of aflatoxin and their relative reaction to the toxigenic isolate of Aspergillus flavus Link ex Fries Punjab Agricultural University J. Res. 15, 400-403
- 3 Bartz Z A, Norden A J, LaPrade J C and Demuynk T J 1976 Tolerance to colonization by Aspergillus flavus found in certain peanut (Arachis hypogees L) breeding lines and cultivars. J Am. Peanut Res. Educ. Assn. 6, 94.
- 4 Dieckert M C and Dieckert J W 1977 Genetically determined structural parameters of the wed cost affecting the colonization of peanut seed by aflatoxin producing Aspergilli. Oleanneux 33, 78

- 5 Doupnik B 1969 Affatoxin produced on peanut varieties previously reported to inhibition production. Phytopathology 59, 1554.
- 6 Kulkarni L. G., Sharief Y. and Sarma V. S. 1967. "Asiriya Mwstunde" groundnut gives good results in Hyderabad. Indian Farming 17, 11-12.
- 7 Mehan V K and McDonald D 1980 Screening for resistance to Aspergillus flasus invasion and aflatoxin production in groundnuts. ICRISAT Groundnut Improvement Program. Occasional Paper-2, 1-15.
- 8 Mehan V K and McDonald D 1981 Affatoxin production in groundnut cultivars resistant and susceptible to seed invasion by Aspergillus flavus. In Proceedings, International Symposium On Mycotoxins, 6-8 September, 1981, Cairo, Egypt.
- 9 Mehan V K, McDonald D and Gibbons R W 1982 Seed colonization and aflatoxin production in groundnut genotypes inoculated with different strains of Aspergillus flavus. Oleagineux 37, 185-191.
- Mehan V K, McDonald D and Lalitha B 1983 Effect of season, location and field-drying treatment on in vitro seed colonization of groundnut genotypes by Aspergillus flavus Oleagineux (In press).
- 11 Mixon A C 1979 Developing groundnut lines with resistance to seed colonization by toxin-producing strains of Aspergillus species. Pest Artic. News Summ. 25, 394 -400
- 12 Mixon A C and Rogers K M 1973 Peanut accessions resistant to seed infection by As pergillus flavus. Agron. J. 65, 560-562.
- 13 Mixon A C and Rogers K M 1975 Factors affecting Aspergillus flavus Lk. ex Fr. colonization of resistant and musceptible genotypes of Arachis hypogene L. Peanut Sci. 2, 18-22.
- 14 Nabney J and Nesbitt B F 1965 A spectrophotometric method for determining the aflatoxins. Analyst 90, 155-160.
- 15 Pons W A, Robertson J A, Goldblatt L A 1966 J. Assn. Off. Chemists' Soc. 43, 665 --669.
- 16 Rao K S and Tulpule P G 1967 Varietal differences of groundnut in the production of aflatoxin. Nature London 214, 738-739.
- 17 Zambittakis Ch, Waliyar F, Bockele-Morvan A and Pins O de 1981 Results of four years of research on resistance of groundnut varieties to Aspergillus flavus. Oleagineux 36, 377-385.