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## Genetics, breeding, and genomics of Indian barnyard millet (Echinochloa frumentacea): status and perspectives

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#### 1. Introduction

Despite the substantial rise in agricultural production in the second half of the twentieth century, just over 12 crops account for 75% of global food resources, and three primary crops, rice, wheat, and maize, accounting for 50% of global food choices (Joshi et al., 2018). The narrow agricultural portfolio raises serious questions on how effectively major crops alone can contribute toward food security and poverty alleviation. Although the fact that these main staple cereal grains are an integral part of many people's diets, they have an unbalanced nutritional profile mainly in terms of essential minerals and protein quality. Besides this, existing cropping systems are also becoming vulnerable to climate change due to their overdependence on few crops (Joshi et al., 2019). To address this issue, efforts must be expanded to incorporate diverse crop species into modern cropping systems. Supplementing major cereals with small millets possessing higher nutritive value and nutraceutical qualities may be a potential strategy for increasing food diversity and building the climate resilience of the modern agroecosystems (Kumar et al., 2021).

Millets are ancient supergrains belonging to the family Poaceae. Among the small millets, barnyard millet (*Echinochloa* spp.) is one of the oldest cultivated crop species. Barnyard millet is a versatile crop that can be used for both food and fodder. In recent times, it has become important due to its noble nutritional composition, recognized health profits, versatile environmental adaptability, viability to marginal agriculture, and organic cultivation (Sood et al., 2020). Barnyard millet in India is predominantly grown under two

varied agro-ecological conditions, one in the Northern part of the country under the lower and mid-hills of the Himalayan region and the other in the southern part mainly in the Deccan plateau region (Sood et al., 2016).

The genus *Echinochloa* comprised two cultivated species *viz. Echinochloa frumentacea* and *E. esculenta. E. frumentacea* is known as Indian Barnyard millet, and *E. esculenta* is known as Japanese barnyard millet (Sood et al., 2015a). Both cultivated species, as well as their progenitors, are hexaploid with 2n = 6x = 54, where x = 9. The Indian barnyard millet is cultivated in India, Nepal, China, Malaysia, and Africa and consumed as a substitute for rice (Anuradha et al., 2014). It has the fastest growing character among all the millets and is well adapted to fragile and hilly mountain ecosystems (Gururani et al. 2021). At present, India is the largest producer of barnyard millet, both in terms of area (0.146 m ha) and production (0.147 mt) (IIMR, 2018).

## 2. Phylogeny, origin, and history

The genus *Echinochloa* belongs to the family Poaceae and subfamily *Panicoideae* and includes 250 described annual and perennial species, of which *E. frumentacea* (Indian barnyard millet) and *E. esculenta* (Japanese barnyard millet) are the most important and widely cultivated (Clayton and Renvoize, 2006). All the species of genus *Echinochloa* have wide adaptation and scattered throughout the warmer and temperate regions of the globe (Bajwa et al., 2015).

*Echinochloa frumentacea* (Roxb.) Link; syn. *E. colona* var. *frumentacea* is generally acknowledged as Indian barnyard millet and thought to have originated from its wild hexaploid (2n = 6x = 54) progenitor *E. Colona* (Hilu, 1994). The wild barnyard millet (*Echinochloa colona*) is a common weed of rice fields, and in several parts of India, it is used as a food source during extreme drought spells (Padulosi et al., 2009).

Indian barnyard millet is one of the oldest domesticated crops and archaeological records of its cultivation date back to 5000 B.C. in India and 3000 B.C in China (Doggett, 1989). Based on the morphology of inflorescence, *E. frumentacea* has been classified into four races, namely, *stolonifera*, *intermedia*, *robusta*, and *laxa*, which are widely distributed in India, Nepal, China, Central Africa, Malawi, and Tanzania (Upadhyaya et al., 2014).

## 3. Botanical description and floral biology

The *E. frumentacea* plant is an annual herb growing 1.0-2.2 m tall characterized by slender to strong culm (Padulosi et al., 2009). In general, plants are green in color, but there are violet tinges in the somatic as well as sexual parts. The plants are generally characterized by smooth, glabrous, and liguleless leaves with slight hairiness and an arc of 15-40 cm long and 1-2.5 cm wide (Sood et al., 2020) (Fig. 5.1).



**Figure 5.1** Barnyard millet plants (A), inflorescence, (B) and grains (C).

The inflorescence is a typically erect and terminal panicle (10-28 cm long) of green to violet color, with numerous and 1-3 cm long racemes, seldom drooping, and mostly awnless (Fig. 5.1). The spikelet present on the panicle is small, unbranched, densely packed in three to five rows, 2-4 mm long, acute, and awnless on the rachis (Renganathan et al., 2020). The spikelet is occupied with stiff bristles and congested. The spikelets are subsided by two glumes within which there are two florets. The upper floret is bisexual, while the lower floret is sterile. The color of the glume varies from white to red. There are three stamens present, and two distinct types of styles each with plumose stigma are found on the superior ovary (Sundararaj and Thulasidas, 1976).

The blooming begins at the inflorescence apex and progresses down and usually takes 10-15 days to complete. Anthesis starts from 5 to 10 a.m., while most of the flowers open between 6 and 7 a.m. (Sundararaj and Thulasidas, 1976; Jayaraman et al., 1997; Gupta et al., 2012). Flowering begins first at the marginal ends of each raceme and progresses to the individual raceme's center. The stigmatic branches extent and the hermaphrodite flower opens afore the anthers dehisce (Seetharam et al., 2003). E. frumentacea has smaller, awnless spikelets, with membranous glumes, whereas *Echinochloa esculenta* has large generally awned spikelets with cretaceous upper glumes and lower lemma (Yabuno, 1987). Contact method is the traditional hybridization method followed in barnyard millet. However, growing all seeds harvested from female panicle for identification of few hybrid plants in contact method requires more resources time, space, and labor (Sood et al., 2020). Hot water treatment of inflorescence at  $48^{\circ}$ C for 4-5 min was also found effective for the emasculation under hill conditions  $(79^{\circ}39' \text{ E and } 29^{\circ}35' \text{ altitude } 1250 \text{ m amsl})$  in barnyard millet (Parihar et al., 2021). In contrast to the contact method, only few seeds set in the female panicle in hot water treatment, most of which give rise to hybrid plants. Grain is around 2-3 mm in length and 1-2 mm in width and bounded by hardened lemma and palea with white shining. The grain color is generally yellow or white.

## 4. Growth, development, and agronomic attributes

Barnyard millet is an important crop for famine areas owed to its rapid growth, drought tolerance, and capability to thrive well under marginal growing conditions (Majid et al., 1989). Although, it can be cultivated in a wide range of growing conditions but requires a warm and moderately humid climate for abundant growth and yield. In general, the crop is sown from the second fortnight of June to the first fortnight of July with the onset of monsoon rains. Planting late lessens the growth period and adversely affects the crop yield. Generally, *E. frumentacea* takes about 70–90 days to complete its life cycle, but under unfavorable circumstances, it may take more time to complete its life span. Owing to its quick growth and short duration, it is an ideal catch crop. Barnyard millet grows under natural precipitation and does not require irrigation facilities.

It favors sandy soil with good drainage, having 4.6–7.4 pH values but has also been adapted to flooded marshland, marginal grounds, and acidic soils with a low pH value of 4.5 and electric conductivity of  $3.12-4.68 \text{ dS m}^{-1}$  (Farrell, 2011). It can be used as retrieve crops in a field that is else saline for rice cultivation. The ideal temperature range for optimal growth of *E. frumentacea* is  $27-33^{\circ}$ C during the day and  $15-22^{\circ}$ C at night (Muldoon et al., 1982). While low-temperature conditions ( $15/10^{\circ}$ C day/night) stop the growth and have an impact on plant production, largely on leaf area and dry weight traits.

Under favorable growing conditions, the crop seeds germinate within 24 h of water imbibition and plants' adventitious roots emerge after 5 days of germination (Rahn et al., 1968). The first tillers appear about 10 days after emergence (Holm et al., 1991). The plants exhibit abundant vegetative growth from 20 to 25 days after emergence, and subsequently, the switch from vegetative to reproductive phase takes place quickly in less than 40 days after emergence (Maun and Barrett, 1986). After emergence, the seed filling period starts in the 55-60 days. After the inception of the reproductive process, seeds mature within 20-40 days. Barnyard millets should be harvested within 10-12 weeks after sowing when 70-75% of the seeds have attained physiological maturity. For drying purpose, the ideal temperature and moisture content are about 45  $^{\circ}$ C and 16%, respectively. Indian barnyard millet has been reported as a quantitative short-day plant (Muldoon, 1985). Plants are small and flower quickly but abundantly under short-day conditions, whereas, under long-day conditions, the plants are more vigorous with a high seed yield (Maun and Barrett, 1986; Manidool, 1992). However, most Echinochloa spp. can grow and reproduce in a wide range of photoperiods, including short (8-13 h) and long (16 h) days (Maun and Barrett, 1986; Mitich, 1990).

The nutrient requirement of barnyard millet is fairly low as compared with major cereals. Farmyard manure or compost at the rate of 5-10 tons per hectare is sufficient for enhancing productivity. Along with organic manures, the crop with 40 kg N<sub>2</sub>, 20 kg P<sub>2</sub>O<sub>5</sub> and 20 kg K<sub>2</sub>O per hectare or as per soil testing recommendations. All the fertilizer should

be added to the soil during sowing time. But, if irrigation facilities are available, half of the nitrogen dose should be top-dressed in a standing crop after 25-30 days of sowing.

### 5. Nutrition composition and nutraceutical potential of barnyard millet

Barnyard millet is a decent source of high-quality digestible protein with the least caloric compactness in comparison to major cereals. Barnyard millet is highly nutritious, comprising about 55% starch, 11% protein, 3.9% fat, and 13.6% crude fiber content, as well as with noteworthy content of calcium and iron (Saleh et al., 2013). The average nutritional composition of barnyard millet in comparison to other major cereals based on per 100 g edible portion is furnished in Table 5.1.

Nutrients	Barnyard millet	Rice	Wheat	Maize	
Proximate composition (per 100 g grain)					
Carbohydrate (g)	65.5	78.2	69.4	66.2	
Protein (g)	11.6	6.8	12.1	11.1	
Fat (g)	5.8	0.5	1.7	3.6	
Fiber (g)	13.5	0.2	1.9	2.7	
Mineral (g)	4.4	0.6	2.7	1.5	
Mineral elements (mg per 100 g grain)					
Iron (mg)	5	0.7	4.9	2.3	
Calcium (mg)	20	10	48	10	
Zinc (mg)	3	1.4	2.2	2.8	
Phosphorus (mg)	280	160	355	348	
Magnesium (mg)	82	90	132	139	
Essential amino acids composition (mg per g protein)					
Isoleusine	288	181	148	154	
Leusine	725	345	269	480	
Lysine	106	166	130	126	
Methionine	133	103	56	85	
Cysteine	175	101	101	97	
Phenylalanine	362	222	179	199	
Tyrosine	150	199	125	166	
Threonine	35	144	125	148	
Tryptophan	63	55	43	16	
Valine	388	269	199	211	

Table 5.1: Nutritional composition of barnyard millet (per 100 g dry weight basis) in comparison to major cereals.

Based on Sources: FAO, 1995. Sorghum and Millets in Human Nutrition, FAO Food and Nutrition Series No. 27. Food and Agricultural Organization, Rome, Italy; Gopalan, C., Ramshashtri, B.V., Balasubramanian, S.C., 1999. Nutritive Value of Indian Foods, second ed.; Narasinga Rao, B.S., Deosthale, Y.G., Pant, K.C. (ed.) National Institute of Nutrition, Indian Council of Medical Research, New Delhi; Gopalan, C., Rama Shastri, B.V., Balasubramanian, S.C., 2009. Nutritive Value of Indian Foods, National Institute of Nutrition, Indian Council of Medical Research, Hyderabad, p 99.

The carbohydrate content of barnyard millet is slowly digestible, making it a natural gift for people suffering from diabetes (Veena et al., 2005). It has also more fiber and iron content than barley, wheat, and other millets. The high fiber content benefits in inhibiting digestion problems such as constipation, excess gas, bloating, and cramping (Rao and Bhaskarachary, 2017). These features make it a worthy contestant for the preparation of industrially processed food products like baby foods, snacks, and dietary food products (Ugare, 2008; Vijayakumar et al., 2009; Anju and Sarita, 2010; Surekha et al., 2013).

Barnyard millet grains are dehusked and baked in the same way as rice grains. Barnyard millet in the Himalayan region is consumed as *paleu* or *chencha*, a savoury cooked porridge made with buttermilk. Whereas, in some parts of South India, the grain is processed very similar to rice parboiling (Bhat et al., 2019) and used in *Idli, dosa, chakli*, etc. Besides these, the biscuits, cakes, sweets, pasta, rusk, ready mix, popped products, and other specialty food products have all been made with barnyard millet (Arora and Srivastava, 2002; Poongodi et al., 2003).

Barnyard millet grains are rich reservoir of flavonoids such as luteolin and tricin, which are known to have anticarcinogenic, anti-inflammatory, antioxidant, and antileukemic properties (Han et al., 2007; Shahidi and Chandrasekara, 2013; Rao et al., 2017). Immature seeds are known to possess diuretic properties and improve virility strength (Chauhan and Johnson, 2011). It is an ideal food for those patients who have gluten intolerance and celiac disease. The low glycemic index of barnyard millet makes it an excellent diet for type II diabetic patients (Sharma et al., 2013). Other functional constituents of barnyard millet grains include  $\gamma$ -aminobutyric acid and  $\beta$ -glucan, which act as antioxidants and help to lower blood lipid levels (Sharma et al., 2016). There is an inevitability to advance millet-based foodstuffs in the formulation of ready-to-use foodstuffs and functional nourishments to fulfill the requirements of the prevailing-day users. Value addition does not merely deal to provide diversity, handiness, and value diet to consumers; however, it is also vital for the resurgence of barnyard millet production (Sood et al., 2015a).

## 6. Adaptation to climate change and local environments

By virtue of its adaptation to low input agriculture and harsh weather patterns, barnyard millet is well adapted to fragile mountain agro-ecosystem. The short life cycle (80–90 days), dense tap root system, thickened cell walls, C4 photosynthesis system, and large leaf area index with high radiation use efficiency barnyard millet makes barnyard millet an ideal crop for future agricultural systems.

## 7. Intercropping systems

Barnyard millet is mostly grown as a pure crop, followed by pulses like chickpea and cereals (wheat and barley). In Uttarakhand hills of India, barnyard millet is mostly grown

as an intercrop with rice bean, black soybean, and horse gram. The profitable barnyard millet intercropping system in hills has been found to be with soybean/rice bean in the ratio of 9:1 (Shalini et al., 2019). The number of studies on intercropping in barnyard millet is limited, and there is a need for extensive research on barnyard millet—based intercropping systems.

## 8. Bottlenecks in barnyard millet breeding

Barnyard millet is predominantly self-pollinating (Potvin, 1986) and self-compatible crop species. Because of the small size of the flowers, the early hours of flowering, the limited pollen viability duration, the nonavailability of the pollen grain, and the short period of flowers opening, emasculation, and artificial hybridization are challenging processes (Nirmala Kumari and Vetriventhan, 2009). Therefore, the success rate of making the cross is usually very low restraining the number of genetic studies and yield advancement in barnyard millet. As a result, most of the present-day's barnyard millet cultivars are the product of traditional mass selection and pure-line selection, and recombination breeding is still a daunting challenge (Joshi et al., 2021). Comprehending the floral biology and pollination pattern will aid in inventing suitable emasculation and crossing procedures (Gupta et al., 2012).

Barnyard millet grain requires dehulling before making it suitable for human consumption (Lohani et al., 2012). Therefore, another important breeding goal for reducing the drudgery of postharvest processing of barnyard millet is to be bred for easy dehulling type genotypes. Some researchers reported the occurrence of easy dehulling genotypes in local germplasm (Gupta et al., 2015), which may be utilized in breeding programs. Since barnyard millet is generally eaten like rice, therefore, the increased grain size is expected to increase market demand. Although, this rests a problem due to the trait's limited spectrum of variability. Besides these, nonshattering behavior is also very critical for yield improvement via reducing yield losses.

## 9. Major biotic and abiotic production constraints

The genus *Echinochloa* can withstand a variety of biotic and abiotic stresses with minimum yield losses. Although, cultivated species *E. frumentacea* is usually vulnerable to many biotic stresses mainly pests and diseases at different stages during crop growth (Jain et al., 1997; Jagadish et al., 2008). Therefore, the major breeding target traits to overcome biotic constraints in barnyard millet are resistance to grain smut, loose smut, sheath blight, shoot fly, stem borer, and aphid. Barnyard millet grain smut (Fig. 5.2) that is caused by *Ustilago panici-frumentacei* Brefeld is the main productivity constraint resulting in yield reduction up to 60.8% (Jain et al., 1997).



Figure 5.2

Smut in barnyard millet (A) grain smut (B) head smut. Source: Reproduced from Sood, S., Khulbe, R.K., Gupta, A.K., Agrawal, P.K., Upadhyaya, H.D., Bhatt, J.C., 2015a. Barnyard millet - a potential food and feed crop of future. Plant Breed 134:135–147. https://doi.org/10.1111/pbr.12243 with permission.

The flower ovaries of infected panicles are round and hairy, and they might expand 2-3 times than their normal size. Gall-like swellings can occasionally be seen on the nodes, axils of older leaves, and the stem. Likewise, *Ustilago crus-galli* Tracy and Earle is responsible for head smut condition (Nagaraja et al., 2007). This happens near the end of the growing season when the crop is about to mature.

The resistance source for grain smut has not been recognized in Indian barnyard millet, whereas Japanese barnyard millet confers strong resistance against it (Joshi et al., 2021). However, due to strong compatibility barriers between the two species, it is difficult to transfer resistance from Japanese barnyard millet to Indian barnyard millet (Sood et al., 2014). The alternate source for the improvement of Indian barnyard millet could be *E. colona*, the wild progenitor that is resistant toward grain smut and crossable with cultivated species. Loose smut caused by *Ustilagotritici* fungus is also the main disease that is responsible for affecting the grain development in both the cultivated species of *Echinochloa* genus (Jain et al., 1997; Gupta et al., 2010a).

On the other hand, Nagaraja and Mantur (2008) demonstrated that some *E. esculenta* accessions were immune to all smut diseases, allowing for the breeding of resistance lines. Gupta et al. (2010a) tested 257 barnyard millet accessions for grain smut tolerance, including advanced breeding lines, and clustered accessions based on response to smut infection, identifying highly resistant accessions. Similarly, *Helminthosporium* leaf blight,

which is a major concern in certain regions, should also be a focus of resistance breeding efforts (Jain et al., 1997).

Among the insect pest, pink stem borer (*Sesamia inferens*) (Fig. 5.3) and shoot fly (*Altherigona falcata*) are the most major insect pests of barnyard millet (Jagadish et al., 2008). The pink stem borer tunnels the stem resulting in dead heart/whitehead formation, while the shoot fly causes tunneling in the nursery stage. Therefore, *E. frumentacea* suffers a significant yield loss due to stem borer and shoot fly infection during the vegetative period.

In India, some promising germplasm accessions and barnyard millet varieties have been identified to be comparatively resistant to shoot fly (Murthi and Harinarayana, 1986). Subsequently, shoot fly and stem borer resistant genotypes have been identified in barnyard millet as DHBM 996 and TNEF-204 (Rawat et al., 2019).



Figure 5.3

Stem borer and its damage in barnyard millet. Source: Reproduced from Sood, S., Khulbe, R.K., Gupta, A.K., Agrawal, P.K., Upadhyaya, H.D., Bhatt, J.C., 2015a. Barnyard millet - a potential food and feed crop of future. Plant Breed. 134:135-147. https://doi.org/10.1111/pbr.12243 with permission. The pink stem borer tunnels the stem, culminating in the creation of a dead heart/white head, whereas the shoot fly tunnels the stem in the nursery stage. The crop has a better storage life and keeping quality, and it is pest-free.

Barnyard millet is well-adapted to abiotic stresses compared with most other cereal crops (Gupta et al., 2010b; Singh et al., 2010; Dwivedi et al., 2012). However, its production is hampered by the lodging, resulting in significant yield and quality losses. Under the high moisture conditions that prevail during the rainy season, barnyard millet can reach a height of 2 m or more making it vulnerable to lodging. This problem could be mitigated by reducing plant height 120 to 130 cm. The resulting decrease in fodder yield due to reduced height could be offset by creating genotypes with more number of basal tillers, which would increase grain yield as well. The length and number of spikes in barnyard millet are positively associated with grain yield. As a result, developing lodging-resistant cultivars with more and longer spikes in combination with good crop husbandry could be the common approach for increasing grain yield.

# **10.** Breeding advancements in barnyard millet 10.1 Conventional breeding strategies

In India, barnyard millet breeding gained interest at the national level in 1969, with the inception of the All India Coordinated Millets Improvement Project. Subsequently, five crop-specific lead research centers in the country were established with International Development Research Center (IDRC) assistance during 1978–79 providing a significant boost to small millets improvement (Seetharam, 1998). Among them, ICAR-VPKAS, Almora in Uttarakhand, was nominated for barnyard millet improvement. Conventional breeding strategies, which include pure line selection, pedigree selection, mass selection, and mutation, that are pertinent to other self-pollinating crops are also equally applicable in barnyard millet. In barnyard millet, most cultivars (about 18 in number) in India were released ensuing pure-line selection from local landraces/cultivars, accompanied with the aid of pedigree selection (Table 5.2) (AICSMIP, 2014).

#### 10.2 Mutation breeding

Mutation breeding has played a key role in self-pollinated crops, where hybridization is challenging to achieve variability (Sood et al., 2019). Mutation breeding was started to induce genetic polymorphism for yield traits. Inducing a mutant phenotype in polyploid species like *Echinochloa* is very challenging. Gamma irradiation treatment is reported to improve genetic variability for the seed yield and yield components such as the number of tillers, plant height, ear head, and length (Mehra et al., 1985).

S. No.	Name of cultivar	Silent features
1.	VL Madira172	Average grain yield is 2.2–2.3 t ha <sup>-1</sup> . Mature in 75–80 days.
2.	VL Madira181	Tolerant to grain smut and well suited to high cropping intensity. Average grain yield 1.6–1.7 t ha <sup>-1</sup> . Mature in 90–95 days.
		Profuse tillering.
3.	VLMadira 207	Average grain yield is $1.6-1.9$ t ha <sup>-1</sup> . Mature in 80–90 days.
		Pyramidical shaped panicles. Tolerant to grain smut. Also having
		improved plant type with high harvest index.
4.	CO(KV) 2	Average grain yield is 2.1–2.2 t $ha^{-1}$ . Mature in 95–100 days.
		Having branched panicles, nonlodging, profuse tillering, and
		suitable for contingency planting.
5.	ER 64 (Pratap sawan1)	Dual-purpose variety giving fodder yield $5.0-5.5$ t ha <sup>-1</sup> and
		average grain yield is $1.5-1.7$ t ha <sup>-1</sup> . Resistant to smut and
		tolerant to shoot fly
6.	RAU 11 (Sushrutha)	Average grain yield is $2.0-2.2$ t ha <sup>-1</sup> . Erect plant type, compact
		panicles. Profuse tillering, high yielding, and very early duration
		(75—80 days).
7.	DHBM 93-3	Average grain yield is 2.2–2.4 t $ha^{-1}$ . Mature in 90–95 days.
		Responsive to fertilizer application

## Table 5.2: Improved and popular cultivars of Indian barnyard millet developed in India (Sood et al., 2020).

## 11. Modern breeding approaches for genetic improvement 11.1 Genetic diversity studies through molecular markers

Initially, Hilu (1994) employed the random amplified polymorphic DNA (RAPD) markers to exploit the genetic divergence and phylogenic relationship between *Echinochloa* species and revealed that RAPD markers were working to differentiate at the genomic level both the cultivated as well as wild progenitors of the *Echinochloa* species. E. frumentacea species was shown to be more genetically diverse than *E. utilis* populations. To solve the problem of the low magnitude of polymorphism by RAPD markers, amplified fragment length polymorphism (AFLP) markers were employed to investigate the genetic divergence in *Echinochloa* species (Danquah et al., 2002; Tabacchi et al., 2009). The microsatellites and polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) were also employed to establish species relationships in genus *Echinochloa* (Danquah et al., 2002; Yamaguchi et al., 2005; Yasuda et al., 2006). Likewise, in E. colona, InDel markers were used for recognition of rbcL, matK, and ITS genes and also had been extensively used as a low-cost method for barcoding of the DNA in E. colona, E. oryzicola, and E. crus-galli species (Lee et al., 2014). Besides SSR markers, the genotyping-by-sequencing (GBS) approach has also been used to generate 21,000 SNPs in barnyard millet core collection germplasm (Wallace et al., 2015). Recently, Babu et al. (2018a, b) employed rice, finger millet, and maize genomic SSRs for cross-transferability analysis among barnyard millet cultivated and wild species for identification of

polymorphic markers, syntenic regions, genetic diversity, and population structure analysis. The SSRs of all the crops were highly transferable, polymorphic, and were able to distinguish barnyard millet genotypes clearly.

#### 11.2 Gene/QTL mapping

In barnyard millet, numerous SSR and SNP markers are recognized (Wallace et al., 2015; Chen et al., 2017; Manimekalai et al., 2018; Murukarthick et al., 2019) to expedite the linkage map creation and QTL mapping; however, no genetic linkage map is reported so far. Till now, only two mapping studies are available publicly in barnyard millet; one for waxy traits (Ishikawa et al., 2013) that used functional SNP markers and revealed that three loci, namely, EeWx1, EeWx1, and EeWx3 were responsible for controlling waxy traits Likewise, Renganathan et al. (2019) employed bulk segregants analysis (BSA) and 51 EST-SSR markers to study the genetics of anthocyanin pigmentation and revealed that the anthocyanin pigments in barnyard millet is closely linked to SSR marker, BMESSR 39. Barnyard millet genome mapping is still in its infancy. Therefore, advance experimental investigation on mapping studies is necessary for the exploitation of markerassisted selection strategy (Renganathan et al., 2020).

#### 11.3 Genomic resources and utilization

The whole-genome sequencing approach is compulsory to comprehend the genome alignment and gene catalog of a crop and to find out key genes as well as pathways linked to traits of economic importance (Joshi et al., 2021). However, limited genomic information, such as DNA markers, genetic/linkage maps, and genome sequences, is the major constraint for the molecular characterization of barnyard millet crop. The whole genome sequencing has already been completed in other millets like sorghum, pearl millet, foxtail millet, finger millet, and proso millet (Zhang et al., 2012; Mace et al., 2013; Hittalmani et al., 2017; Varshney et al., 2017; Zou et al., 2019). Due to genome complexity and the dearth of exploration effort in barnyard millet, very restricted attempts have been made to ascertain the genomic organization as well as associated advancements. Genomic resources from closely related species where genome sequences are existing could be exploited toward supplementing genomic resources in barnyard millet.

## 12. Comparative genomics and synteny studies

Comparative genomics plays a critical role by using the synteny evidence between conserved regions among the crops plant belonging to the same family (Moore et al., 1995; Gale and Devos, 1998; Pattanayak et al., 2019). Owing to the scarcity of genome sequence information, comparative genomic studies will be an effective tool for genomic

studies in barnyard millet. A high amount of cross-transferability was revealed in barnyard millet by using the genomic microsatellite markers that were established via in silico mining for foxtail millet (Pandey et al., 2013; Kumari et al., 2013). The 100 intron-length polymorphic markers mined from the foxtail millet genome revealed cross transferability of 94% in Indian barnyard millet (Muthamilarasan and Prasad, 2014). Likewise, more than 70% cross transferability of rice genic SSR obtained from the calcium transporters and calcium kinase primers to Indian barnyard millet was demonstrated (Yadav et al., 2014). These markers would be of tremendous and profound value for studies of genetic diversity, creation of linkage map, and additionally their use in marker recognition for significant agro-morphological traits that are linked to QTLs in barnyard millet. Further, these recognized QTLs will be able to efficiently introgressed through marker-assisted selection in the barnyard millet genotypes that are locally adapted for enhancement of yield and stress regulation.

# **13.** Functional genomics approach for gene discovery 13.1 Transcriptomics

Gene expression profiling based on transcriptome analysis is a powerful functional genomics technique for identifying candidate genes that control a variety of biological processes (Kumar et al., 2016). Enormous transcript profiles for different characters accompanied with invasiveness and adaptations processes like resistance toward herbicides, photosynthesis, flooding, and other genes related to homeostasis, have been developed in the weedy *Echinochloa* species (Li et al., 2013a,b; Yang et al., 2013; Nah et al., 2015; Xu et al., 2015; Guo et al., 2017; Gao et al., 2018). However, transcriptome profiles related to traits of economic importance like disease resistance and nutritional quality are yet to be developed in the Indian barnyard millet.

#### 13.2 Proteomics and metabolomics

Proteomics is a functional genomics technique that infers to the inclusive analysis of a whole set of proteins in terms of organization, expression, and functional properties. The proteomics often employed various techniques such as 2-D gel electrophoresis, mass spectrophotometry, and gel-free shotgun (Matros et al., 2011). The speedy advancements toward proteomics techniques via proteome mapping, comparative proteomics, and the discovery of protein—protein interactions are enabling new perceptions into plant genomes (Varshney et al., 2009). However, the studies using proteomics with major emphasis to comprehend seed quality characters may not yet have been applied in barnyard millet. Metabolomics is a new "omics" technology that involves identifying, characterizing, and quantifying biomolecules having a low molecular weight in a biological environment (Kumar et al., 2016). These low-molecular-weight biomolecules are referring as

metabolomes. Various secondary metabolites such as polyphenols and flavonoids have been revealed to play a significant role in the nutraceutical belongings of barnyard grain. As a result, comprehensive metabolomics studies are needed to conclude the best genotypes for human consumption. However, barnyard millet grain metabolomics profiling is yet to be available.

#### 13.3 Genetic transformation methods

To satisfy the growing global demand for cereals, new methods, and techniques for developing new varieties are needed (Gaspairs et al. 2008; Jagga-Chugh et al., 2011). Development of transgenic plants with predictable traits is one of these effective approaches. In contrast to major cereals, effective transformation protocols are yet to be standardized in small millets including barnyard millet. However, efforts toward the development of a fast, effective, and repeatable regeneration protocol (Rajak et al., 2018) are encouraging efforts toward the standardization of genetic transformation barnyard millet. So far, there is only one report available regarding genetic transformation studies in barnyard millet, which is based on biolistic transformation for evaluating the efficacy of different promoters in GUS expression (Gupta et al., 2001). However, there has been no transformation attempt through the *Agrobacterium*-mediated method in barnyard millet.

## 14. Conclusion and future outlook

Barnyard millet breeding plans have come to a halt, owing to a lack of support in terms of capital and patronage from different funding and research organizations. Hence, greater efforts are required to evolve cultivars with economic traits that are preferred by the farmer as well as the consumer. In the future, more breeding plans are needed to take advantage of the genetic variability to improve the yield potential, stabilization of yield, salinity tolerance, pest and disease resistance, and superior nutritional worth, primarily micronutrient constitutes (Renganathan et al., 2020). Nevertheless, due to a scarcity of genetic and genomic resources in barnyard millet, the advances in breeding programs are very slow.

Current crop production is challenged especially due to climate change, which has resulted in enlarged drought, salinity, and heat problems (Mabhaudhi et al., 2019). Besides this, agricultural production resulted in an untenable way owing to dependence on expensive chemical fertilizers and pesticides, which are responsible for environmental degradation and increased production costs (Fernie and Yan, 2019). Under such circumstances, the use of underutilized and potential crops like barnyard millet, which can offer diversification for food and genetic resources, is one of the prospective ways to tackle these problems. Due to their diverse adaptation on marginal lands, low input requirements, reduced vulnerability to stresses, and nutritional supremacy as paralleled to main staple cereals. Increasing consumer's awareness of balanced diets, as well as the climate-resilient properties, highlights the requisite of putting more effort into research for barnyard millet. As there is a scarcity of germplasm and data on genetic multiplicity, their tangible use in crop improvement programs is restricted. Hence, highlighting the germplasm pool is vital for recognizing trait precise resources, genes and alleles, which can be employed in barnyard millet improvement. The yield hindrance in barnyard millet can be smashed through the use of male sterility and manipulating heterosis, and enhancement of genomics-assisted breeding, in conjugation using improved cultivation practices. To speed up barnyard millet improvement, genomics-aided breeding would expedite the credentials of novel alleles and genes with better agronomical efficiency as well as tolerance to biotic and abiotic stresses. To upscale the barnyard millet as a worthwhile crop to farmers, the researchers must need to be characterized available germplasm resources, utilization genetic and genomic tools designed for breeding high-yielding cultivars, and development of improved packages of practices for cultivation, as well as the processing and improving consumption through value addition.

Thus, there is an urgent need to develop a roadmap to upkeep barnyard millet cultivation, through various efforts like the development of distinct cultivars for explicit environments and pursuing attributes upgrading through modern breeding and genomic tools. These strategies would aid in the fighting against starvation and malnutrition and also conjointly benefiting to the farmers and other players tangled in barnyard millet cultivation in the face of climate change.

## References

- AICSMIP, 2014. Report on the Compendium of Released Varieties in Small Millets [Internet]. Bengaluru, India. http://www.dhan.org/smallmillets/docs/report/Compendium of Released Varieties in Small millets. pdf (Accessed 13 March 2021).
- Anju, T., Sarita, S., 2010. Suitability of foxtail millet (*Setaria italica*) and barnyard millet (*Echinochloa frumentacea*) for development of low glycemic index biscuits. Malays J Nutr 16, 361–368.
- Anuradha, N., Patro, T.S.S.K., Bhanu, U.K., Madhuri, J., Sowjanya, A., 2014. Multivariate analysis in barnyard millet (*Echinochloa frumentacea*). J. Food Agricul. Veter. Sci. 4 (2), 194–199.
- Arora, S., Srivastava, S., 2002. Suitability of millet-based food products for diabetics. J. Food Sci. Technol. 39, 423–428.
- Babu, B.K., Rashmi, C., Sood, S., 2018a. Cross transferability of finger millet and maize genomic SSR markers for genetic diversity and population structure analysis of barnyard millet. Indian J. Genet. 78 (3), 364–372.
- Babu, B.K., Sood, S., Kumar, D., Joshi, A., Pattanayak, A., Kant, L., Upadhyaya, H.D., 2018b. Cross genera transferability of rice and finger millet genomic SSRs to barnyard millet (Echinochloa spp.). 3 Biotech 8, 95. https://doi.org/10.1007/s13205-018-1118-1.
- Bajwa, A., Jabran, K., Shahid, M., Ali, H.H., Chauhan, B., Ehsanullah, 2015. Eco-biology and management of *Echinochloa crus-galli*. Crop Protect. 75, 151–162. https://doi.org/10.1016/j.cropro.2015.06.001.
- Bhat, B.V., Arunachalam, A., Kumar, D., Tonapi, V.A., Mohapatra, T., 2019. Millets in the Indian Himalaya. Indian Council of Agricultural Research, New Delhi, p. 84.

- Chauhan, B., Johnson, D.E., 2011. Ecological studies on *Echinochloa crus-galli* and the implications for weed management in direct-seeded rice. Crop Protect. 30, 1385–1391. https://doi.org/10.1016/ j.cropro.2011.07.013.
- Chen, G., Zhang, W., Fang, J., Dong, L., 2017. Identification of massive molecular markers in *Echinochloa phyllopogon* using a restriction-site associated DNA approach. Plant Divers 39, 287–293. https://doi.org/ 10.1016/j.pld.2017.08.004.
- Clayton, W.D., Renvoize, S.A., 2006. Genera Graminum: Grasses of the World in Kew Bulletin Additional Series XIII, Royal Botanical Gardens Kew. University of Chicago Press, Chicago, IL.
- Danquah, E.Y., Hanley, S.J., Brookes, R.C., Aldam, C., Karp, A., 2002. Isolation and characterization of microsatellites in *Echinochloa* (L.) Beauv. spp. Mol. Ecol. Notes 2, 54–56. https://doi.org/10.1046/j.1471-8286.2002.00144.x.
- Doggett, H., 1989. Small millets a selective overview. In: Seetharam, A., Riley, K.W., Harinarayana, G. (Eds.), Small Millets in Global Agriculture. Oxford and IBH Publ. Co. Pvt. Ltd, Janpath, New Delhi, pp. 3–18.
- Dwivedi, S., Upadhyaya, H.D., Senthilvel, S., Hash, C.T., Fukunaga, K., Diao, X., et al., 2012. Millets: genetic and genomic resources. Plant Breed. Rev. 35, 247–375.
- FAO, 1995. Sorghum and Millets in Human Nutrition. FAO Food and Nutrition Series No. 27. Food and Agricultural Organization, Rome, Italy.
- Farrell, W., 2011. Plant Guide for Billion-Dollar Grass (*Echinochloa frumentacea*). USDA-Natural Resources Conservation Service.
- Fernie, A.R., Yan, J.B., 2019. *De novo* domestication: an alternative route toward new crops for the future. Mol. Plant 12, 615–631.
- Gale, M.D., Devos, K.M., 1998. Comparative genetics in the grasses. Proc Nat Acad Sci USA 95, 1971–1974.
- Gao, Y., Li, J., Pan, X., Liu, D., Napier, R., Dong, L., 2018. Quinclorac resistance induced by the suppression of the expression of 1-aminocyclopropane-1-carboxylic acid (ACC) synthase and ACC oxidase genes in *Echinochloa crus-galli* var. zelayensis. Pestic. Biochem. Physiol. 146, 25–32. https://doi.org/10.1016/ j.pestbp.2018.02.005.
- Gaspairs, S., Bregire, C., Orczyk, W., Nadolska-Orczyk, A., 2008. Agrobacterium mediated transformation of oat (*Avena sativa* L.) cultivars via immature embryo and leaf explants. Plant Cell Rep. 27, 1721–1729.
- Gopalan, C., Ramshashtri, B.V., Balasubramanian, S.C., 1999. In: Narasinga Rao, B.S., Deosthale, Y.G., Pant, K.C. (Eds.), Nutritive Value of Indian Foods, second ed. National Institute of Nutrition, Indian Council of Medical Research, New Delhi.
- Gopalan, C., Rama Shastri, B.V., Balasubramanian, S.C., 2009. Nutritive Value of Indian Foods. National Institute of Nutrition, Indian Council of Medical Research, Hyderabad, p. 99.
- Guo, L., Qiu, J., Ye, C.-Y., Jin, G., Lingfeng, M., Zhang, H., 2017. *Echinochloa crus-galli* genome analysis provides insight into its adaptation and invasiveness as a weed. Nat. Commun. 8, 1031. https://doi.org/ 10.1038/s41467-017-01067-5.
- Gupta, P., Raghuvanshi, S., Tyagi, A.K., 2001. Assessment of the efficiency of various gene promoters via biolistics in leaf and regenerating seed callus of millets, *Elusine coracana* and *Echinochloa crus-galli*. Plant Biotechnol. 18, 275–282.
- Gupta, A., Joshi, D., Mahajan, V., Gupta, H.S., 2010a. Screening barnyard millet germplasm against grain smut (*Ustilago panici-frumentcei* Brefeld). Plant Genet. Resour. Charact. Util. 8 (1), 52–54.
- Gupta, A., Mahajan, V., Gupta, H.S., 2010b. Genetic resources and varietal improvement of small millets for Indian Himalaya. In: Tewari, L.M., Pangtey, Y.P.S., Tewari, G. (Eds.), Biodiversity Potentials of the Himalaya. Gyanodaya Prakashan, Nainital, India, pp. 305–316.
- Gupta, A., Sood, S., Agrawal, P.K., Bhatt, J.C., 2012. Floral biology and pollination system in small millets. Eur. J. Plant Sci. Biotechnol. 6 (2), 80–86.
- Gupta, A., Sood, S., Agrawal, P.K., Bhatt, J.C., 2015. B 29: an easy dehulling barnyard millet (*Echinochloa frumentacea* Link) genotype. Natl. Acad. Sci. Lett. 38, 21–24. https://doi.org/10.1007/s40009-014-0295-7.
- Gururani, K., Sood, S., Kumar, A., Joshi, D.C., Pandey, D., Sharma, A.R., 2021. Mainstreaming Barahnaja cultivation for food and nutritional security in the Himalayan region. Biodivers. Conserv. 30, 551–574.

- Han, X., Shen, T., Lou, H., 2007. Dietary polyphenols and their biological significance. Int. J. Mol. Sci. 8 (9), 950–988.
- Hilu, K.W., 1994. Evidence from RAPD markers in the evolution of *Echinochloa* millets (*Poaceae*). Plant Syst. Evol. 189 (3–4), 247–257.
- Hittalmani, S., Mahesh, H.B., Shirke, M.D., Biradar, H., Uday, G., Aruna, Y.R., 2017. Genome and Transcriptome sequence of Finger millet (*Eleusine coracana* (L.) Gaertn.) provides insights into drought tolerance and nutraceutical properties. BMC Genom. 18, 465. https://doi.org/10.1186/s12864-017-3850-z.
- Holm, L.G., Plunknett, D.L., Pancho, J.V., Herberger, J.P., 1991. The World's Worst Weeds. Distribution and Biology. Krieger Publishing Company, Malabar, FL, p. 609.
- IIMR, 2018. Annual Report 2017-18. Indian Institute of Millets Research, Hyderabad.
- Ishikawa, G., Seimiya, Y., Saito, M., Nakamura, T., Hoshino, T., 2013. Molecular characterization of spontaneous and induced mutations in the three homoeologous waxy genes of Japanese barnyard millet [*Echinochloa esculenta* (A. Braun) H. Scholz]. Mol. Breed. 31, 69–78. https://doi.org/10.1007/s11032-012-9769-9.
- Jagadish, P.S., Mohapatra, H.K., Chakravarthy, M.K., Srivastava, N., Nangia, N., 2008. A Compendium of Insect Pests of Finger Millet and Other Small Millets. All India Coordinated Small Millets Improvement Project, GKVK, Bangalore, p. 60.
- Jagga-Chugh, S., Kachhwaha, S., Sharma, M., Kothari-Chajer, A., Kothari, S.L., 2011. Optimization of factors influencing micro projectile bombardment-mediated genetic transformation of seed-derived callus and regeneration of transgenic plants in *Eleusine coracana* (L.) Gaertn. Plant Cell Tissue Organ Cult. 109, 401–410.
- Jain, A.K., Jain, S.K., Yadava, H.S., 1997. Assessment of yield losses due to grain smut in barnyard millet. Indian Phytopathol. 50, 49–52.
- Jayaraman, N., Suresh, S., Nirmala Ganeshan, N.M., 1997. Genetic enhancement and breeding strategies in small millets. In: National Seminar on Small Millets: Current Research Trends and Future Priorities as Food Feed and in Processing for Value Addition. 23–24 April, 1997, Bangalore, India, pp. 19–21.
- Joshi, D.C., Sood, S., Lakshmi Hosahatti, R., Kant Pattanayak, A., Kumar, A., Yadav, D., Stetter, M.G., 2018. From zero to hero: the past, present and future of grain amaranth breeding. Theor. Appl. Genet. 131, 1807–1823.
- Joshi, D.C., Chaudhari, G.V., Sood, S., Kant, L., Pattanayak, A., Zhang, K., Fan, Y., Janovska, D., Meglic, V., Zhou, M., 2019. Revisiting the versatile buckwheat: reinvigorating genetic gains through integrated breeding and genomics approach. Planta 250, 783–801.
- Joshi, D.C., Meena, R.P., Chandora, R., 2021. Genetic resources: collection, characterization, conservation and documentation. In: Singh, M., Sood, S. (Eds.), Millets and Pseudocereals. Woodhead publishing, pp. 25–31.
- Kumar, A., Metwal, M., Kaur, S., Gupta, A.K., Puranik, S., Singh, S., Singh, M., Gupta, S., Babu, B.K., Sood, S., Yadav, R., 2016. Nutraceutical value of finger millet [*Eleusine coracana* (L.) Gaertn.], and their improvement using omics approaches. Front. Plant Sci. 7, 1–14.
- Kumar, A., Tripathi, M.K., Joshi, D., Kumar, V., 2021. Millets and Millet Technology. Springer, Singapore, p. 438.
- Kumari, K., Muthamilarasan, M., Misra, G., Gupta, S., Alagesan, S., Parida, S., 2013. Development of eSSR-Markers in *Setaria italica* and their applicability in studying genetic diversity, cross-transferability and comparative mapping in millet and non-millet species. PLoS One 8, e67742. https://doi.org/10.1371/ journal.pone.0067742.
- Lee, J., Park, K., Lee, I., Kim, C., Kown, O., Park, T., 2014. Simple sequence repeat analysis of genetic diversity among Acetyl-CoA carboxylase inhibitor-resistant and -susceptible *Echinochloa crus-galli* and *E. oryzicola* populations in Korea. Weed Res. 55, 90–100. https://doi.org/10.1111/wre.12119.
- Li, G., Wu, S., Cai, L., Wang, Q., Zhao, X., Wu, C., 2013a. Identification and mRNA expression profile of glutamate receptor-like gene in quinclorac resistant and susceptible *Echinochloa crus-galli*. Gene 531, 489–495. https://doi.org/10.1016/j.gene.2013.09.013.

- Li, G., Wu, S.G., Yu, R.X., Cang, T., Chen, L.P., Zhao, X.P., 2013b. Identification and expression pattern of a glutathione S-transferase in Echinochloa crus-galli. Weed Res. 53, 314–321. https://doi.org/10.1111/ wre.12031.
- Lohani, U.C., Pandey, J.P., ShahiNC, 2012. Effect of degree of polishing on milling characteristics and proximate compositions of barnyard millet (*Echinochloa frumentacea*). Food Bioprocess Technol. 5 (3), 1113–1119. https://doi.org/10.1007/s11947-011-0518-6.
- Mabhaudhi, T., Chibarabada, T., Chimonyo, V., Murugani, V., Pereira, L., Sobratee, N., Govender, L., Slotow, R., Modi, A., 2019. Mainstreaming underutilized indigenous and traditional crops into food systems: a South African perspective. Sustainability 11, 172. https://doi.org/10.3390/su11010172.
- Mace, E.S., Tai, S., Gilding, E.K., Li, Y., Prentis, P.J., Bian, L., 2013. Whole-genome sequencing reveals untapped genetic potential in Africa's indigenous cereal crop sorghum. Nat. Commun. 4, 2320. https:// doi.org/10.1038/ncomms3320.
- Majid, M.A., Hamid, M.A., Mannujan, 1989. Importance, genetic resources and breeding of small millets in Bangladesh. In: Seetharam, A., Riley, K.W., Harinarayana, G. (Eds.), Small Millets in Global Agriculture. Oxford & IBH Publishing Co., New Delhi, pp. 71–76.
- Manidool, C., 1992. Echinochloa crus-galli (L.) P. Beauv. In: t' Mannetje, L., Jones, R.M. (Eds.), Plant Resources of South-East Asia. No. 4. Forages. Pudoc Scientific Publishers, Wageningen, the Netherlands, p. 303.
- Manimekalai, M., Dhasarathan, M., Karthikeyan, A., Murukarthick, J., Renganathan, V.G., Thangaraj, K., 2018. Genetic diversity in the barnyard millet (*Echinocholafrumentacea*) germplasms revealed by morphological traits and simple sequence repeat markers. Curr. Plant Biol. 14, 71–78. https://doi.org/10.1016/ j.cpb.2018.09.006.
- Matros, A., Kaspar, S., Witzel, K., Mock, H.P., 2011. Recent progress in liquid chromatography-based separation and label-free quantitative plant proteomics. Phytochemistry 72, 963–974.
- Maun, M.A., Barrett, S.C., 1986. The biology of Canadian weeds. 77. Echinochloa crus- galli (L.) Beauv. Can. J. Plant Sci. 66, 739–759.
- Mehra, H.S., Joshi, H.C., Chikara, J., 1985. Induced mutations in Japanese millet. Indian J. Agric. Sci. 55, 294–295.
- Mitich, L.W., 1990. Intriguing world of weeds. Barnyard grass. Weed Technol. 4, 918-920.
- Moore, G., Devos, K.M., Wang, Z., Gale, M.D., 1995. Grasses, line up and form a circle. Curr. Biol. 5, 17-23.
- Muldoon, D.K., 1985. The effect of photoperiod on the growth and development of *Echinochloa* spp. Millets. Aust. J. Exp. Agric. 25, 428–433.
- Muldoon, D.K., Perason, C.J., Wheleer, J.L., 1982. The effect of temperature on growth and development of *Echinochloa* millets. Ann. Bot. 50, 665–672.
- Murthi, T., Harinarayana, G., 1986. Insect pests of small millets and their management in India. In: Seetharam, A., Riley, K.W., Harinarayana, G. (Eds.), Small Millets in Global Agriculture. Oxford and IBH publishing, New Delhi, India, pp. 255–270.
- Murukarthick, J., Manimekalai, M., Karthikeyan, A., Perumal, S., Dhasarathan, M., Kandasamy, T., 2019. Transcriptomes of Indian barnyard millet and barnyard grass reveal putative genes involved in drought adaptation and micronutrient accumulation. Acta Physiol. Plant. 41, 66. https://doi.org/10.1007/s11738-019-2855-4.
- Muthamilarasan, M., Prasad, M., 2014. Advances in *Setaria* genomics for genetic improvement of cereals and bioenergy grasses. Theor. Appl. Genet. 128, 1–14. https://doi.org/10.1007/s00122-014-2399-3.
- Nagaraja, A., Mantur, S.G., 2008. Evaluation of barnyard millet entries for grain smut resistance and yield. Mysore J. Agric. Sci. 42, 375–377.
- Nagaraja, A., Kumar, J., Jain, A.K., Narasimhudu, Y., Raghuchander, T., Kumar, B., Hanumanthe Gowda, B., 2007. Compendium of Small Millets Diseases. All India Coordinated Small Millets Improvement Project, GKVK, Bangalore, p. 80.
- Nah, G., Im, J.H., Kim, J.W., Park, H.R., Yook, M.J., Yang, T.J., 2015. Uncovering the differential molecular basis of adaptive diversity in three *Echinochloa* leaf transcriptomes. PLoS One 10, e0134419. https:// doi.org/10.1371/journal.pone.0134419.

- Nirmalakumari, A., Vetriventhan, M., 2009. Phenotypic analysis of anther and pollen in diversified genotype of barnyard millet (*Echinochloa frumentaceae*) floral characters. ICFAI Univ. J. Genet. Evol. 2, 12–16.
- Padulosi, S., Bhag, M.C., Bala, R.S., Gowda, J.V., Gowda, K.T.K., Shanthakumar, G.C., 2009. Food security and climate change: role of plant genetic resources of minor millets. Indian J. Plant Genet. Resour. 22, 1–16.
- Pandey, G., Misra, G., Kumari, K., Gupta, S., Kumar Parida, S., Chattopadhyay, D., 2013. Genome-wide development and use of microsatellite markers for large-scale genotyping applications in foxtail millet [Setaria italica (L.)]. DNA Res. 20, 197–207. https://doi.org/10.1093/dnares/dst002.
- Parihar, M., Kumar, A., Bisht, J.K., Bhinda, M.S., Shyamnath, Meena, R.P., Mondal, T., Joshi, D.C., Bijarniya, H., Singh, S., Kant, L., 2021. Reviving the forgotten food network of potential crops to strengthen nutritional and livelihood security in North Western Himalayas. Indian J. Agron. 66, 44–59.
- Pattanayak, A., Roy, S., Sood, S., Iangrai, B., Banerjee, A., Gupta, S., Joshi, D.C., 2019. Rice bean: a lesser known pulse with well-recognized potential. Planta. https://doi.org/10.1007/s00425-019-03196-1.
- Poongodi, T., Vijayakumar, J.B.M., Nazni, P., Rajeshwari, M., 2003. Value addition for minor millets and its glycemic load among normal and type 2 diabetic subjects. In: Paper presented at the 39th national Nutritional Conference of NSI, Hyderabad.
- Potvin, C., 1986. Biomass allocation and phenological differences among southern and northern populations of the C<sub>4</sub> grass *Echinochloa crus-galli*. J. Ecol. 74, 915–923.
- Rahn, E.M., Sweet, R.D., Vengris, J., Dunn, S., 1968. Life history studies as related to weed control in the Northeast. In: Barnyardgrass, 5(368). University of Delaware, Agricultural Research Station, Bulletin, pp. 1–46.
- Rajak, K., Tiwari, N., Kumari, R., Rathore, S., 2018. Standardize protocol for callus induction and plant regeneration in barnyard millet using different combination of plant growth regulators. Int. J. Curr. Microbiol. App. Sci. (Special Issue-6), 2590–2596.
- Rao, B.D., Bhaskarachary, K., 2017. Nutrition and Health Benefits of Millets. ICAR- Indian institute of millets research, ISBN 81-89335-68-5.
- Rao, B.D., Bhaskarachary, K., Christina, G.D.A., Devi, G.S., Tonapi, V.A., 2017. Nutritional and Health Benefits of Millets. ICAR- Indian Institute of Millets Research (IIMR), Hyderabad, India.
- Rawat, L., Nautiyal, A., Bisht, T.S., Prasad, S., Naithani, D., Makhloga, K., 2019. Screening of barnyard millet germplasm against shoot fly and stem borer damage under field conditions. Int. J. Curr. Microbiol. Appl. Sci. 8, 1221–1226. https://doi.org/10.20546/ijcmas.2019.802.142.
- Renganathan, V.G., Vanniarajan, C., Ramalingam, J., 2019. Genetic analysis and identification of molecular markers linked to the anthocyanin pigmentation in barnyard millet [*Echinochloa frumentacea* Roxb (Link)]. In: Proceedings of the Neglected and Underutilized Crop Species for Food, Nutrition, Energy and Environment. NIPGR), New Delhi, p. 43.
- Renganathan, V.G., Vanniarajan, C., Karthikeyan, A., Ramalingam, J., 2020. Barnyard millet for food and nutritional security: current status and future research direction. Front. Genet. 11, 500. https://doi.org/ 10.3389/fgene.2020.00500.
- Saleh, A.S.M., Zhang, Q., Chen, J., Shen, Q., 2013. Millet grains: nutritional quality, processing, and potential health benefits. Compr. Rev. Food Sci. Food Saf. 12 (3), 281–295.
- Seetharam, A., 1998. Small millets research: achievements during 1947-97. Indian J. Agric. Sci. 68, 431–438. Seetharam, A., Gowda, J., Halaswamy, J.H., 2003. Small millets. In: Chowdhury, S.K., Lal, S.K. (Eds.),
- Nucleus and Breeder Seed Production Manual. Indian Agricultural Research Institute, New Delhi, India, pp. 54–67.
- Shahidi, F., Chandrasekara, A., 2013. Millet grain phenolics and their role in disease risk reduction and health promotion: a review. J. Funct.Foods 5 (2), 570–581.
- Shalini, B., Didal, V.K., 2019. Sustaining minor millet production in hilly areas of Uttarakhand through intercropping of minor millets and pulses- A review. Int. J. Curr. Microbiol. App. Sci. 8 (11), 397–406.
- Sharma, A., Sood, S., Khulbe, R.K., 2013. Millets food for the future. Biotech Today 3, 52. https://doi.org/ 10.5958/j.2322-0996.3.1.010.

- Sharma, A., Sood, S., Agrawal, P.K., Kant, L., Bhatt, J.C., Pattanayak, A., 2016. Detection and assessment of nutraceuticals in methanolic extract of finger (*Eleusine coracana*) and Barnyard Millet (*Echinochloa frumentacea*). Asian J. Chem. 28, 1633–1637. https://doi.org/10.14233/ajchem.2016.19790.
- Singh, K.P., Mishra, H.N., Saha, S., 2010. Moisture-dependent properties of barnyard millet grain and kernel. J. Food Eng. 96, 598–606. https://doi.org/10.1016/j.jfoodeng.2009.09.007.
- Sood, S., Khulbe, R., Saini, N., Gupta, A., Agrawal, P.K., 2014. Interspecific hybrid between *Echinochloaesculenta* (Japanese barnyard millet) and *E. frumentacea* (Indian barnyard millet) - a new avenue for genetic enhancement of barnyard millet. Electron. J. Plant Breed. 5, 248–253.
- Sood, S., Khulbe, R.K., Gupta, A.K., Agrawal, P.K., Upadhyaya, H.D., Bhatt, J.C., 2015a. Barnyard millet a potential food and feed crop of future. Plant Breed. 134, 135–147. https://doi.org/10.1111/pbr.12243.
- Sood, S., Khulbe, R.K., Kant, L., 2016. Optimal yield related attributes for high grain yield using ontogeny based sequential path analysis in barnyard millet (*Echinochloa* spp.). J. Agric. Sci. Technol. 18, 1933–1944.
- Sood, S., Joshi, D.C., Chandra, A.K., Kumar, A., 2019. Phenomics and genomics of finger millet: current status and future prospects. Planta. https://doi.org/10.1007/s00425-019-03159-6.
- Sood, S., Joshi, D.C., Pattanayak, A., 2020. Breeding advances in barnyard millet. In: Wani, S.H. (Ed.), Accelerated Plant Breeding Volume 1 Cereal Crops (Gosal SS. springer, Germany, pp. 391–409.
- Sundararaj, D.P., Thulasidas, G., 1976. Botany of Field Crops. Macmillan Publisher, New Delhi, India, p. 509.
- Surekha, N., Ravikumar, S.N., Mythri, S., Rohini, D., 2013. Barnyard millet (*Echinochloa frumentacea* Link) cookies: development, value addition, consumer acceptability, nutritional and shelf life evaluation. IOSR J. Environ. Sci. Toxicol. Food Technol. 7, 01–10. https://doi.org/10.9790/2402-0730110.
- Tabacchi, M., Mantegazza, R., Spada, A., Ferrero, A., 2009. Morphological traits and molecular markers for classification of *Echinochloa* species from Italian rice fields. Weed Sci. 54, 1086–1093. https://doi.org/ 10.1614/WS-06-018R1.1.
- Ugare, R., 2008. Health Benefits, Storage Quality and Value Addition of Barnyard Millet (*Echinochloa frumentacaea* link) (M.Sc thesis). University of Agricultural Sciences, Dharwad.
- Upadhyaya, H., Dwivedi, S.L., Singh, S.K., Sube, S., Vetriventhan, M., Sharma, S., 2014. Forming core collections in Barnyard, Kodo, and little millets using morpho-agronomic descriptors. Crop Sci. 54, 2673–2682. https://doi.org/10.2135/cropsci2014.03.0221.
- Varshney, R.K., Nayak, S.N., May, G.D., Jackson, S.A., 2009. Next-generation sequencing technologies and their implications for crop genetics and breeding. Trends Biotechnol. 27, 522–530.
- Varshney, R.K., Shi, C., Thudi, M., Mariac, C., Wallace, J., Qi, P., 2017. Pearl millet genome sequence provides a resource to improve agronomic traits in arid environments. Nat. Biotechnol. 35, 969–976. https://doi.org/10.1038/nbt.3943.
- Veena, S., Chimmad, B.V., Naik, R.K., Shanthakumar, G., 2005. Physicochemical and nutritional studies in Barnyard millet. Karnataka J. Agric. Sci. 18, 101e105.
- Vijayakumar, T.P., Mohankumar, J.B., Jaganmohan, R., 2009. Quality evaluation of chapati from millet flour blend incorporated composite flour. Indian J. Nutr. Diet 46, 144–155.
- Wallace, J.G., Upadhyaya, H.D., Vetriventhan, M., Buckler, E.S., Hash, C.T., Ramu, P., 2015. The genetic makeup of global barnyard millet germplasm collection. Plant Genome 8 (1), 1–7.
- Xu, W., Di, C., Zhou, S., Liu, J., Li, L., Liu, F., 2015. Rice transcriptome analysis to identify possible herbicide quinclorac detoxification genes. Front. Genet. 6, 306. https://doi.org/10.3389/fgene.2015.00306.
- Yabuno, T., 1987. Japanese barnyard millet (Echinochloautilis, Poaceae) in Japan. Econ. Bot. 41, 484-493.
- Yadav, S., Gaur, V., Jaiswal, J.P., Kumar, A., 2014. Simple sequence repeat (SSR) analysis in relation to calcium transport and signaling genes reveals transferability among grasses and a conserved behavior within finger millet genotypes. Plant Syst. Evol. 300, 1–8. https://doi.org/10.1007/s00606-014-0982-3.
- Yamaguchi, H., Utano, A.Y.A., Yasuda, K., Yano, A., Soejima, A., 2005. A molecular phylogeny of wild and cultivated *Echinochloa* in East Asia inferred from non-coding region sequences of trnT-L-F. Weed Biol. Manag. 5, 210–218. https://doi.org/10.1111/j.1445-6664.2005.00185.x.

- Yang, X., Yu, X.Y., Li, Y.F., 2013. De novo assembly and characterization of the barnyard grass (*Echinochloa crus-galli*) transcriptome using next-generation pyrosequencing. PLoS One 8, e69168. https://doi.org/ 10.1371/journal.pone.0069168.
- Yasuda, K., Yano, A., Nakayama, Y., Yamaguchi, H., 2006. Molecular identification of *Echinochloa oryzicola* Vasing. and *E. crus-galli* (L.) Beauv. using a polymerase chain reaction-restriction fragment length polymorphism technique. Weed Biol. Manag. 2, 11–17. https://doi.org/10.1046/j.1445-6664.2002.00041.x.
- Zhang, G., Liu, X., Quan, Z., Cheng, S., Xu, Pan, S., 2012. Genome sequence of foxtail millet (*Setaria italica*) provides insights into grass evolution and biofuel potential. Nat. Biotechnol. 30, 549–554. https://doi.org/ 10.1038/nbt.2195.
- Zou, C., Li, L., Miki, D., Li, D., Tang, Q., Xiao, L., 2019. The genome of broomcorn millet. Nat. Commun. 10, 436. https://doi.org/10.1038/s41467-019-08409-5.

#### Further reading

Sood, S, Khulbe, R.K., ArunKumar, R., Agrawal, P.K., Upadhaya, H.D., 2015b. Barnyard millet global core collection evaluation in the submontane Himalayan region of India using multivariate analysis. Crop J. 3, 517–525. https://doi.org/10.1016/j.cj.2015.07.005.