

Pre-breeding efforts for widening the genetic base of cool-season food legumes

KUMAR SHIV¹, SHARMA SHIVALI², KUMAR JITENDRA³, MAALOUF FOUAD¹, HAMWIEH ALADDIN¹, GAUR POORAN² and SARKER ASHUTOSH¹

¹International Center for Agricultural Research in the Dry Areas (ICARDA), Beirut, Lebanon; ²International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Telangana State, India; ³ICAR-Indian Institute of Pulses Research, Kanpur, India; E-mails: sk.agrawal@cgiar.org; shivali.sharma@cgiar.org

Globally, cool-season food legumes (CSFL) mainly chickpea, field pea, lentil, faba bean, and grass pea are grown on ~33 million ha with a production of ~46 million tonnes, contributing 47% to the global pulse output. These crops hold important position in sustainable diversified agri-food systems. Average productivity of CSFL is 1380 kg/ha which is far below their genetic potential. Theaverage annual genetic gain reported in various legume crops is estimated 0.7% which is not enough to meet the growing demand of pulses. Despite many improved varieties. narrow genetic base has been identified as one of the major constraints for limited genetic gain in CSFL crops. This is basically due to the bottlenecks during the process of evolution and domestication, which has further been compounded using limited germplasm in national and international breeding programs. Consequently, many useful alleles/variants have been left behind in landraces and crop wild relatives (CWR). This scenario is changeable, provided we conserve, evaluate and utilize CWR in breeding these crops. Sizeable number of CWR accessions are available in the global collections held at ICARDA and ICRISAT, screening of them has resulted in identification of useful donors not only for resistance to major biotic and abiotic stresses but also for phenology, plant type and yield traits. Breeders have traditionally been reluctant about the use of CWR in breeding programs due to complex and unpredictable outcomes because of crossing barriers, linkage drag, andlong breeding cycle. There are now modern tools and techniques available to identify CWR with useful alleles that will enable breeders to introgress into desired agronomic back grounds more precisely and efficiently.

Introduction

Chickpea (Cicer arietinum L.), field pea (Pisum sativum L.), lentil (Lens culinaris Medikus subsp. culinaris), faba bean (Vicia faba L.), and grass pea (Lathyrus sativus L.) are the important cool-season food legume (CSFL) crops for sustainable diversified agri-food systems. These crops are currently grown on 33 million ha area with a production of 46 million tonnes. During the last five decades, CSFL crops, especially chickpea and lentil have experienced spectacular growth in global production, mainly because of their expansion in new areas along with enhanced productivity. However, the current average productivity of CSFL crops is estimated at 1380 kg/ha which is far below their genetic potential. The annual genetic gain reported in legume crops is estimated at 0.7% (Bogale et al. 2015, Belete et al. 2017, Idrissi et al. 2019). Despite many improved varieties, narrow genetic base has been identified as one of the major constraints for limited genetic gains in CSFL crops. This is basically due to the bottlenecks during the process of evolution and domestication, which was further compounded during the directed breeding programs. Pedigree analysis of chickpea and lentil varieties released in India confirmed the extensive and repetitive use of a handful of germplasm as parents in hybridization (Kumar et al. 2004). In lentil, for example, an early flowering exotic line Precoz (ILL4605) has been utilized extensively to tailor plant architecture having vigorous growth, early maturity, and large seeds, particularly for Indo-Gangetic plains (Kumar et al. 2014). One option to widen the genetic base of CSFL crops is to mainstream a more systematic and targeted use of crop wild relatives (CWR) in breeding programs (Dempewolf et al. 2017). CWR contains a wealth of genetically important traits/alleles due to their adaptation to a diverse range of habitats. Plant breeders have long recognized the potential value of CWR, but the absence of a simple means to determine which CWR might hold valuable genetic variation has severely limited their use. Now, with the advent of modern tools and techniques combined with systematic phenotyping, it is feasible to mainstream CWRs and prioritize their use for crop improvement.



Present Status

ICARDA genebank holds 50,968 accessions of legume crops including 15,749 of chickpea, 14,597 of lentil, 10,034 of faba bean, 6,131 of peas and 4,457 of grasspea. The current holdings also include CWR of lentil (619), chickpea (547) and grasspea (1555). Despite having *Narbonens is* as a rich CWR, it is not accessible to faba bean improvement due to strong crossing barriers (Caracuta *et al.* 2016). However, four subspecies, namely *major*, *equina*, *minor* and *paucijuga* have been identified within *Vicia faba* for faba bean improvement. Similarly, ICRISAT genebank holds one of the largest collections of *Cicer* species wherein 20,764 accessions of cultivated and wild *Cicer* species are conserved for use in chickpea breeding programs. Despite large collections of these crops, there are major germplasm gaps at species and genotype levels, and a continuum is very much required to fill these gaps in wild genepool from the unrepresented areas of diversity in genebanks. For example, vast majority of *Cicer* collections are cultivated germplasm with many duplicates. A recent collection mission has expanded the available CWR collections of both *C. reticulatum* and *C. echinospermum* by over 10-fold (von Wettberg *et al.* 2018). Similarly, out of 187 species in the genus *Lathyrus*, global collection at ICARDA represents only 45 species.

Screening of CWR

Wild species are valuable sources of novel variation for yield traits, phenology, and resistance to biotic and abiotic stresses. Previous screening shows marked genetic variability for desired traits among landraces and CWR in lentil, chickpea, faba bean and grass pea. Wild Lens has shown to carry resistance/tolerance for key stresses including drought, cold, salinity, Ascochyta blight, Stemphylium bight, rust, Fusarium wilt, Sitona weevil, Orobanche, powdery mildew and Anthracnose. The highest frequency of resistance was observed for Stemphylium blight in L. lamottei followed by L. ervoides (Podder et al. 2013) and Sitona weevil in L. odemensis followed by L. ervoides (El-Bouhssini et al. 2008). Some wild accessions showing combined resistance to Ascochyta blight and Fusarium wilt or anthracnose diseases have also been identified (Bayya et al. 1995, Tullu et al. 2006). Wild relatives of lentil also offer drought tolerance in L. nigricans, L. odemensis and L. ervoides (Gupta and Sharma 2006) and cold tolerance in L. culinaris ssp. orientalis (Hamdi et al. 1996). Donors for yield traits like 100-seed weight and pods/plant have been observed in L. lamottei and L. culinaris ssp orientalis (Gupta and Sharma 2006. Singh et al. 2014). L. ervoides has been identified as a good source of growth habit, phenology, biomass and seed traits (Tullu et al. 2011, Tullu et al. 2013, Singh et al. 2013, Kumar et al. 2014). Singh et al. (2014) evaluated the global wild Lens taxa originating from 27 countries under diverse agro-climatic conditions in India for three cropping seasons, resulting in wide variations for many yield attributing traits including multiple disease resistance in L. nigricans and L. ervoides accessions. Nutritional quality traits have been measured in wild Lens with significant diversity for RFO, raffinose, and verbascose (Tahir et al. 2012) and mineral (Sengupta et al. 2016, Kumar et al. 2018) concentrations.

Annual wild *Cicer* species have great potential for chickpea improvement through genetic base broadening (von Wettberg *et al.* 2018) and by providing adaptive traits lost in the cultigen. These species have been screened for reaction to Ascochyta blight, Botrytis grey mold, dry root rot, Fusarium wilt, cyst nematode, leaf miner, seed beetle, and cold tolerance both at ICRISAT and ICARDA, and a high level of resistance to each stress has been identified. *Cicer judaicum* is reported to have resistance genes for Ascochyta blight, Fusarium wilt and Botrytis grey mould and *C. bijugum*, *C. pinnatifidum* and *C. reticulatum* for resistance to cyst nematode. For example, ILWC 7-1 of *C. bijugum* showed resistance to Ascochyta blight, Fusarium wilt, leaf miner, cyst nematode and cold, and ILWC 33/S-4 of *C. pinnatifidum* to Ascochyta blight, Fusarium wilt, seed beetle, and cyst nematode.

Screening of faba bean landraces has resulted in identification of germplasm with multiple disease resistance and heat, and herbicide tolerance in *minor*, *major* and *equina* subspecies with successful examples of introgression of Orobanche resistance to major types. Screening of 2000 landraces against Ascochyta blight, chocolate spot, and rust has resulted in identification of 424 accessions as resistant to chocolate spot, 136 to Ascochyta blight and 52 to rust (Maalouf *et al.* 2016). Similarly, screening of faba bean landraces have resulted in identification of tolerance to Orobanche (Maalouf *et al.* 2011) and herbicide (Maalouf *et al.* 2018).

Wild gene pool is a rich reservoir of rare alleles for grass pea improvement which have been evaluated for ODAP content and Orobanche tolerance. Evaluation of 142 accessions of *L. cicera* showed a range of 0.073–0.513% for



ODAP content which is much lower than that in the cultivated species (Kumar et al. 2013). Accessions of L. cicera are also good source of earliness, Orobanche tolerance and cold tolerance. Evaluation of 490 CWR accessions of Lathyrus revealed significant variation for ODAP content ranging from 0.024 to 0.456%. On an average, Lathyrus annuus, L. cicera, and L. gorgoni showed low ODAP content as compared to the cultivated species. Results of 286 accessions representing 13 species showed good sources of resistance to Orobanche and low ODAP content in CWR (Fig. 1).

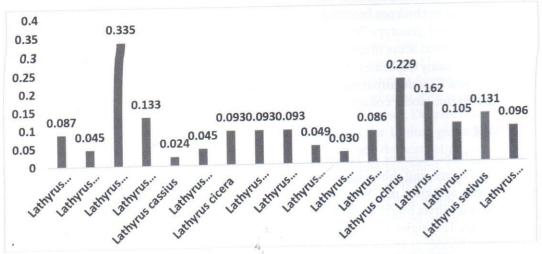


Fig 1. Screening of Lathyrus CWR for low ODAP content during 2017

Use of CWR in breeding programs

Generally, high yielding lines are crossed to secure a high mean performance of the resulting progenies. To identify superior progenies, to ensure genetic gain in the next selection cycle, and to maintain long-term selection gain, it is important that the cross also generates high genetic variance. Mainstreaming CWR in breeding programs help to ensure high genetic variance among resultant progenies and maintain long term selection gains in a breeding program. However, the use of wild germplasm in breeding programs has been restricted mainly due to difficulties in access to exotic germplasm, extreme regional specificity of adaptation, large number of uncharacterized accessions, crossing barriers, linkage drags, and perception that wide crosses would disturb favorable combinations fixed over a period in cultivated germplasm and result in inferior recombinants. For example, interspecific crosses among Lathyrus species are extremely difficult with limited success. Similarly, transferring Orobanche resistance from faba bean minor to fababean major is difficult due to strong negative association between large type and Orobanche tolerance. Within the cultivated lentils, extreme specificity of adaptation limits the scope of direct introduction of exotic landraces in lentil breeding program. South Asian landraces are generally early maturing small seeded red lentils and West Asian landraces are late maturing large seeded mostly yellow lentils. Similarly, chickpea has a narrow genetic base, and this reduces the range of adaptive alleles, therefore it limits the potential solutions sought for by breeders to make adaptation selection for a wide range of agro-ecological zones. To widen the genetic base, pre-breeding efforts at ICRISAT and ICARDA breeding programs include parents of diverse origins to combine traits contributing to yield, appropriate phenology, adaptation to major biotic and abiotic stresses and market preferred traits. Derivatives from crosses between South and West Asian parents in lentil have generally shown higher yields mainly due to larger seed size introduced from the West Asian parents in the typical short duration background of South Asian genotypes (Shrestha et al. 2005).

Sources of extra earliness in lentil CWR, ILWL118 having less than 90 days maturity and high iron and zinc content in ILWL74 and ILWL80 have encouraged breeders to biofortify short-duration lentil through pre-breeding (Kumar *et al.* 2017). ICARDA in collaboration with NARS partners in India has implemented a pre breeding project in lentil, which has been instrumental in introgressing useful genes in mainstream breeding. Transgressive segregants



for agronomically important traits have been mined from wide crosses (Kumar *et al.* 2011, Singh *et al.* 2013, Kumar *et al.* 2014). Recent development in lentil improvement has been the successful hybridization of the cultivated lentil with *L. ervoides* using embryo rescue (Tullu *et al.* 2013) and introgression of resistance to *Orobanche crenata* and anthracnose (Fiala *et al.* 2009, Tullu *et al.* 2011). Recently, successful use of *Lens orientalis and L. ervoides* as a source of genes for resistance to key diseases, phenology, micro-nutrients and possibly for plant habit, and other important agronomic traits has been demonstrated with the development of elite prebred lines of lentil at ICARDA. These pre-bred lines not only performed well with >40% yield advantage over the best check (Bakaria) but also rich in micronutrients and fit in short season windows of 80-100 days (Fig. 2).

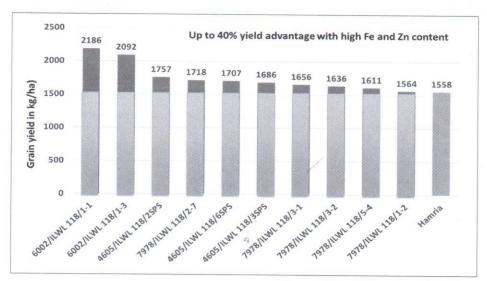


Fig. 2. Yield performance of prebred lines of lentil (Lens culinaris x L. orientalis) at Marchouch in Morocco during 2017.

Development of improved chickpea germplasm with high and stable yields and market-acceptability adapted to the Mediterranean production systems is a major goal of ICARDA Kabuli-chickpea program. Recently, good progress has been made in terms of desirable trait introgression from wild species. During the last seven years, about 10% of 996 crosseshave *C. reticulatum* as one of the parents to introgress resistance to cold, drought, heat and fusarium wilt into cultivated chickpea. This has resulted in the development of improved lines with high potential yield and resistance/tolerance to major diseases and abiotic stresses (FLIP06-165C, FLIP06-166C, FLIP06-167C, FLIP06-168C, FLIP06-173C). Grain yield of 20 breeding lines developed from interspecific crosses indicated the potential of wild *Cicer* in yield increase (Fig. 3).

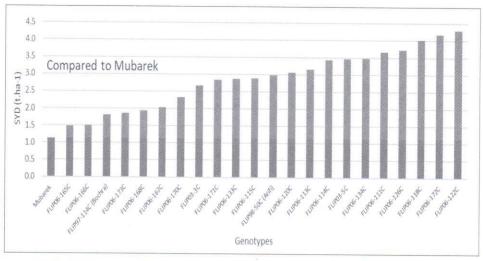


Fig 3. Performance of prebred lines of chickpea in yield trials at Terbol, Lebanon



At ICRISAT, precise evaluation of wild *Cicer* species has resulted in identification of accessions having high levels of resistance for Ascochyta blight, Botrytis grey mould and dry root rot. These accessions have been utilized in transferring resistance genes into popular chickpea cultivars following interspecific hybridization. Major focus of these activities is to combine heat tolerance with dry root rot resistance and short duration with botrytis grey mold resistance. To meet these objectives, simple (bi-parental) and complex (3-way and 4-way cross crosses) cross approaches are followed for creating new variability in the primary genepool. Using complex 3-way crosses [*C. arietinum* × (*C. reticulatum* × *C. echinospermum*)], advanced backcross populations were developed in *desi* and *kabuli* chickpea genetic backgrounds (Sharma *et al.* 2016). Considerable variability for morpho-agronomic traits was observed in these populations (Saran 2019). Promising introgress lines having high-yield, early maturity, erect growth habit, high 100-seed weight, and better agronomic performance under rainfed environments (Verma 2019) have been identified for use in chickpea breeding programs (Fig. 4). Similarly, *Cicer echinospermum* has been used as a source for *Ascochyta* resistance, particularly in the Australian breeding program (Knights *et al.* 2008).

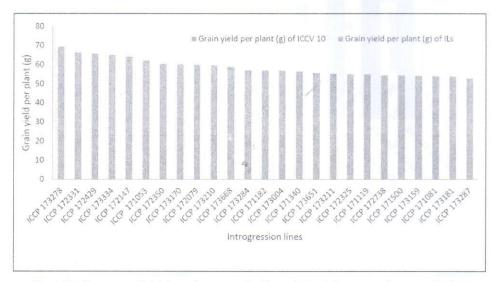


Fig 4. Performance of chickpea introgression lines derived from complex cross in desi background during 2018-19 post-rainy season at ICRISAT, Patancheru, India

A set of introgression lines in *desi* background were screened for botrytis grey mold resistance at hot spot (GBPUA&T, Pantnagar). After two years of evaluation (2017-18 and 2018-19), three introgression lines, ICCP171033, ICCP171418, and ICCP172606 were found resistant (score ≤3.0) and five introgression lines, ICCP171312, ICCP171452, ICCP171454, ICCP171711, ICCP172468, ICCP172527 moderately resistant (score 4.0-5.0) to BGM. These pre-bred lines derived from wild *Cicer* species will provide new and diverse variability for important traits for further use in chickpea improvement programs to develop new cultivars with a broad genetic base.

In faba bean, several varieties with high level of resistance to chocolate spot using landraces have been released in Ethiopia. Prominent among them are Moti, Gebelcho, Obsie and Walki (Temesgen *et al.* 2015). Recently, variety named Gora (ILB2717- 1 × R878-1) has been released in Ethiopia with higher degree of resistance to chocolate spot and larger seed size.

New Approaches for mainstreaming CWR

During domestication and directed breeding, many alleles are inadvertently left behind in landraces and wild species. Introgression of the unattended alleles using innovative breeding tools could provide prebred lines for further use in main breeding programs. There are now genetic procedures to identify CWR with adaptation to local biotic and abiotic stresses and introgress them with ease. For example, application of population genomic scans can detect loci with exceptionally high population Fst values, possibly indicating loci with divergent selection for local adaptation (Baute *et al.* 2015). Conventionally, breeders have used CWR in their breeding schemes typically as sources of



resistance to various biotic and abiotic stresses with limited success. Because of linkage drag, many undesired traits persist in segregating generations which need to be removed at an early stage via repeated backcrossing with recipient parent accompanied by testing at every stage to recover the recurrent genotype with desired traits only. With the availability of high-density genome-wide molecular markers and linkage maps, we can use molecular markers as foreground and background selection for elimination of unwanted plants and recovery of recipient genotype at an early stage of the breeding procedure. Over 90% of the recurrent parental genotype can be recovered within two generations when a suitable number of markers and an adequate number of progenies are used for background selection (Tanksley *et al.* 1989). This represents a substantial saving in time compared to conventional backcross breeding. The procedure of introgression can further be fastened by integrating rapid generation advancement to advance introgression lines quickly as shown in an F₂ population derived from a cross between *Lens culinaris* Medik. and *Lens ervoides* (Lulsdorf and Banniza 2018). A recent international collaboration has built a large introgression resource from the newly expanded collection of wild diversity in chickpea (von Wettberg *et al.* 2018). Advanced introgression lines, currently in F₄ to F₆ stages, have been developed, using 20 *C. reticulatum* and 6 *C. echinospermum* parents into five different cultivated accessions (von Wettberg *et al.* 2018, Shin *et al.* 2018).

Conclusion

SS

ıd

ed

ot

И.

or

ed

ty

ld

er nd

CI

on

of

12)

The successful use of crop wild relatives holds promise as a source of genes for resistance to diseases, phenology, nutritional quality, and possibly for plant habit, biomass production, and other important agronomic and market traits. With rapid advances in embryo rescue techniques, speed breeding, genomic tools and high throughput phenotyping techniques, the prospect of transferring useful traits from wild gene pools in CSFL crops has brightened. Wide genetic base of cultivated varieties provides a type of insurance against the epidemics of diseases and insect pests besides, of course, making the cultivated germplasm more amenable to breeding advances.

References

- 1. Baute GJ, Dempewolf H and Reisenberg LH. 2015. Using genomic approaches to unlock the potential of CWR for crop adaptation to climate change. Pp 268-280. In: Crop wild relatives and climate change, (Redden R, Yadav SS, Maxted M, Dulloo E, Guarino L and Smith P Eds). Wiley-Blackwell, USA.
- 2. Bayaa B, Erskine W and Hamdi A. 1995. Evaluation of a wild lentil collection for resistance to vascular wilt. Genetic Resources and Crop Evolution 42:231–235.
- Belete T, Mekbib F and Eshete M. 2017. Assessment of genetic improvement in grain yield potential and related traits of kabuli type chickpea varieties in Ethiopia (1974-2009). Advances in Crop Science Technology 5:3 Doi:10.4172/2329-8863.1000284.
- Bogale DA, Mekibib F and Fikre A. 2015. Genetic improvement of lentil (*Lens culinaris* Medikus) between 1980 and 2010 in Ethiopia. Malaysian Journal of Medicine and Biology Research 2:284-297.
- 5. Caracuta V, Weinstein-Evron M, Kaufman D, Yeshurun R, Silvent J and Boaretto E. 2016. 14,000-year-old seeds indicate the Levantine origin of the lost progenitor of faba bean. Scientific Reports6: 37399.
- 6. Dempewolf H, Baute G, Anderson J, Kilian B, Smith C and Guarino L. 2017. Past and future use of wild relatives in crop breeding. Crop Science 57:1070-1082.
- 7. El-Bouhssini M, Sarker A, Erskine W and Joubi A. 2008. First sources of resistance to *Sitona* weevil (*Sitona crinitus* Herbst) in wild *Lens* species. Genetic Resources and Crop Evolution 55:1-4.
- 8. Fiala JV, Tullu A, Banniza S, Séguin-Swartz G and Vandenberg A. 2009. Interspecies transfer of resistance to Anthracnose in lentil (*Lens culinaris* Medic.). Crop Science 49:825–830.
- Gupta D and Sharma SK. 2006. Evaluation of wild Lens taxa for agro-morphological traits, fungal diseases and moisture stress in northwestern Indian hills. Genetic Resources and Crop Evolution 53:1233-1241.
- 10. Hamdi A, Küsmenoglu I and Erskine W. 1996. Sources of winter hardiness in wild lentil. Genetic Resources and Crop Evolution 43:63–67.
- 11. Idrissi O, Sahri A, Houasli C and Nsarellah N. 2019. Breeding progress, adaptation, and stability for grain yield in Moroccan lentil improved varieties. Crop Science 59:925-936.

37



- 12. Knights EJ, Southwell RJ, Schwinghamer MW and Harden S. 2008. Resistance to *Phytophthora medicaginis* Hansen and Maxwell in wild *Cicer* species and its use in breeding root rot resistant chickpea (*Cicer arietinum* L.). Australian Journal of Agricultural Research 59:383-387.
- 13. Kumar J, Srivastava E, Singh M, Kumar S, Nadarajan N and Sarker A. 2014. Diversification of indigenous genepool by using exotic germplasm in lentil (*Lens culinaris* Medikus subsp. *culinaris*). Physiology and Molecular Biology of Plants 20:125–132.
- 14. Kumar S, Choudhary AK, Rana KS, Sarker A and Singh M. 2018. Bio-fortification potential of global wild annual lentil core collection. PLoS ONE, 13(1): e0191122.
- 15. Kumar S, Gupta P, Barpete S, Sarker A, Amri A, Mathur PN and Baum M. 2013. Grass pea. Pp 269 292. In: Genetic and Genomic Resources for Grain Legume Improvement (Singh M, Upadhyay HD and Bisht IS Eds.), Elsevier.
- 16. Kumar S, Gupta S, Chandra S and Singh BB. 2004. How wide is the genetic base of pulse crops. Pp 211-221. In: Pulses in New Perspective (Ali M, Singh BB, Kumar S and Vishwa Dhar Eds.), Indian Society of Pulses Research and Development, Kanpur.
- Kumar S, Imtiaz M, Aditya Pratap and Gupta S. 2011. Distant Hybridization and Alien Gene Introgression in Food Legumes. Pp 81-110. In: Biology and breeding of legume crops (Aditya Pratap and Kumar J. Eds), CAB International, UK.
- Kumar S, Kumar J, Dikshit HK and Sarker A. 2017. Mainstreaming Pre-breeding to enhance Genetic Gains in Lentil. In: Abstract - National symposium on "Pulses for Nutritional Security and Agricultural Sustainability" at ICAR-Indian Institute of Pulses Research (ICAR-IIPR), Kanpur, India. Dec 2-4, 2017.
- 19. Lulsdorf MM and Banniza S. 2018. Rapid generation cycling of an F2 population derived from a cross between *Lens culinaris* Medik. and *Lens ervoides* (Brign.) Grande after aphanomyces root rot selection. Plant Breeding 137:486-491.
- Maalouf F, Ahmed S and Somanagouda P. 2018. Developing improved varieties of faba bean. In: Achieving sustainable cultivation of grain legumes Volume 2: Improving cultivation of grain legumes (Sivasankar, S. et al. eds.), Burleigh Dodds Science Publishing, Cambridge, UK.
- 21. Maalouf F, Ahmed S, Shaaban K, Bassam B, Nawar F, Singh M and Amri A. 2016. New faba bean germplasm with multiple resistances to Ascochyta blight, chocolate spot and rust diseases. Euphytica 211:157-167.
- 22. Maalouf F, Khalil S, Ahmed S, Akintunde AN, Kharrat M, El Shama'a K and Malhotra RS. 2011. Yield stability of faba bean lines under diverse broomrape prone production environments. Field Crops Research124:288–294.
- 23. Podder R, Banniza S and Vandenberg A. 2013. Screening of wild and cultivated lentil germplasm for resistance to Stemphylium blight.Plant Genetic Resources 11:26-35.
- Saran D. 2019. Studies on genetic variability and identification of drought tolerant introgression lines from distant crosses of chickpea. M.Sc. dissertation. JNKVV Jabalpur.
- 25. Sengupta D, Thavarajah D, McGee RJ, Coyne CJ, Kumar S and Thavarajah P. 2016. Genetic diversity among cultivated and wild lentils for iron, zinc, copper, calcium and magnesium concentrations. Australian Journal of Crop Science10:1381-1387.
- 26. Sharma Shivali, Upadhyaya HD, Roorkiwal M and Varshney RK. 2016. Inter-specific hybridization for chickpea (*Cicer arietinum* L.) improvement. Pp 446-471. In: Polyploidy and Hybridization for Crop Improvement (Mason AS, ed.), CRC Press.
- 27. Shin MG, Bulyntsev SV, Chang PL, Korbu LB, Carrasquila-Garcia N, Vishnyakova MA, Samsonova MG, Cook DR and Nuzhdin SV. 2019. Multi-trait analysis of domestication genes in *Cicer arietinum–Cicer reticulatum* hybrids with a multidimensional approach: Modeling wide crosses for crop improvement. Plant Science 285:122-131.
- Shrestha R, Siddique KHM, Turner NC, Turner DW and Berger JD. 2005. Growth and seed yield of lentil (*Lens culinaris* Medikus) genotypes of West Asian and South Asian origin and crossbreds between the two under rainfed conditions in Nepal. Australian Journal of Agricultural Research 56:971-981.
- 29. Singh M, Bisht IS, Kumar S, Dutta M, Bansal KC, Karale M, Sarker A, Amri A, Kumar S and Datta SK. 2014. Global wild annual lens collection-A potential resource for lentil genetic base broadening and yield enhancement. PLOS ONE 9(9): e107781.



- 30. Singh M, Rana MK, Kumar K, Bisht IS, Dutta N, Gautam NK, Sarker A and Bansal C. 2013. Broadening the genetic base of lentil cultivars through inter-sub-specific and interspecific crosses of Lens taxa. Plant Breeding 132:667–675.
- 31. Tahir M, Båga M, Vandenberg A and Chibbar RN. 2012. An assessment of raffinose family oligosaccharides and sucrose concentration in genus *Lens*. Crop Science52:1713-1720.
- 32. Tanksley SD, Young ND, Paterson AH and Bonierbale MW. 1989. RFLP mapping in plant breeding: new tools for an old science. Biotechnology 7:257-264.
- 33. Temesgen T, Keneni G, Sefera T and Jarso M. 2015. Yield stability and relationships among stability parameters in faba bean (*Vicia faba* L.) genotypes. Crop Journal 3:258–268.
- 34. Tullu A, Banniza S, Bett K and Vandenberg A. 2011. A walk on the wild side, exploiting wild species for improving cultivated lentil. Grain Legumes 56:13–14.
- 35. Tullu A, Bett K, Banniza S, Vail S and Vandenberg A. 2013. Widening the genetic base of cultivated lentil through hybridization of *Lens culinaris* 'Eston' and *L. ervoides* accession IG 72815. Canadian Journal of Plant Science 93:1037-1047.
- 36. Tullu A, Buchwaldt L, Lulsdorf M, Banniza S, Barlow B, Slinkard AE, Sarker A, Tar'an B, Warkentin TD and Vandenberg A. 2006. Sources of resistance to anthracnose (*Colletotrichum truncatum*) in wild *Lens* species. Genetic Resources and Crop Evolution 53:111-119.
- 37. Verma V. 2019. Tapping wild *Cicer* species for enriching variability in cultivated genepool for chickpea improvement. M.Sc. dissertation. JNKVV, Jabalpur.
- 38. Von Wettberg EJB, Chang PL, Başdemir F, et al. 2018. Ecology and genomics of an important crop wild relative as a prelude to agricultural innovation. Nature Communications 9(1):649.