# "EFFECT OF DIFFERENT IPM COMPONENTS ON Helicoverpa armigera Hubner AND THEIR IMPACT ON NATURAL ENEMIES IN CHICKPEA"

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Thesis Submitted to the Acharya N.G. Ranga Agricultural University in partial fulfilment of the requirements for the award of the Degree of

# **Doctor of Philosophy in Agriculture**



# DEPARTMENT OF ENTOMOLOGY AGRICULTURAL COLLEGE BAPATLA - 522 101 June, 2001.

# **CERTIFICATE**

Mr. V. Visalakshmi has satisfactorily prosecuted the course of research and that the thesis entitled "EFFECT OF DIFFERENT IPM COMPONENTS ON *Helicoverpa armigera* Hubner AND THEIR IMPACT ON NATURAL ENEMIES IN CHICKPEA" submitted is the result of original research work and is of sufficiently high standard to warrant its presentation to the examination. I also certify that the thesis or part thereof has not been previously submitted by her for a degree of any university.

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## CERTIFICATE

This is to certify that the thesis entitled "Effect of different IPM components on Helicoverpa armigera (Hubner) and their impact on natural enemies in chickpea" submitted in partial fulfillment of the requirements for the degree of 'Doctor of phylosophy' of the Acharya N G Ranga Agricultural University, Hyderabad, is a record of the bonafide research work carried out by Mrs V.Visalakshmi under my guidance and supervision. The subject of the thesis has been approved by the student's Advisory Committee.

No part of the thesis has been submitted for any other degree or diploma. The published part has been fully acknowledged. All assistance and help received during the course of the investigation have been duly acknowledged by the author of the thesis.

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Date : 2.6.200

# **DECLARATION**

I, V. VISALAKSHMI, hereby declare that the thesis entitled "EFFECT OF DIFFERENT IPM COMPONENTS ON *Helicoverpa armigera* Hubner AND THEIR IMPACT ON NATURAL ENEMIES IN CHICKPEA" submitted to Acharya N.G. Ranga Agricultural University for the degree of Doctor of Philosophy in Agriculture in the major field of Entomology is the result of original research work done by me. I also declare that any material contained in the thesis has not been published earlier.

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Date : 2 6 200)

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### ABSTRACT

Name of the Author	:	V. VISALAKSHMI
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Degree	:	Doctor of Philosophy
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Investigations were carried out on the Effect of Different IPM Components on *Helicoverpa armigera* Hubner and their Impact on Natural Enemies in Chickpea" during *rabi* 1998-99 and 1999-2000 at ICRISAT Center. The various options of Integrated pest management (IPM) included, botanicals such as neem, insect pathogen (HaNPV), bird perches and chemical insecticides.

The pest infestation was observed throughout the cropping period in both the years with peak population during first week of January and last week of December, 1999. In both the years neem treatment effectively reduced the egg laying by *H.armigera* moths followed by IPM treatment which had neem as one of the components. During vegetative stage of the crop, endosulfan and HaNPV proved effective in reducing small and medium and large size larvae, respectively. But during the remaining period of the crop growth IPM maintained its superiority in reducing larvae of all age groups in both the years. Erecting bird perches was as effective as endosulfan in reducing large size larvae in the peak period of bird activity. Endosulfan was observed to be a more harmful IPM component in affecting the natural enemy fauna present on ground and also on foliage of the crop. Neem also reduced the natural enemy fauna to a lesser extent. No egg parasitism by *Trichogramma* was observed in both the years though a few dead *Trichogramma* adults were observed on chickpea plants. Up to 11 and 10% natural larval parasitism by *Campoletis chlorideae* Uchida was recorded during *rabi* 1998-99 and 1999-2000 seasons, respectively. Except endosulfan all the remaining IPM components proved to be safe to *C.chlorideae*. IPM treatment was proved to be more economical than individual components except bird perches with 1:2.30 & 1:3.76 cost benefit ratio's during *rabi* 1998-99 & 1999-2000 seasons, respectively. Plots treated with endosulfan were found to have residues in seed as well as in husk even at harvest stage.

Neem proved as effective oviposition deterrent on chickpea foliage under cage studies. The antifeedant effect of neem was also proved under choice and no choice situations in laboratory. Neem produced several abnormalities in *H.armigera* like mortality during larval stage, increased larval and pupal duration, reduced pupal weight, reduced effective oviposition period and fecundity when treated at larval stages, but these effects were more pronounced on early stages of larvae than later stages. However it had no effect on egg hatchability. Robin blue 1% proved to be a good ultraviolet ray protectant and increased the persistence of HaNPV up to six days under field conditions. HaNPV was found to have more impact on early stages of larvae than later stages. In addition to high larval mortality it was found to produce several ill effects on *H.armigera* like pupal abnormality, pupal death, reduced pupal weight and reduced adult emergence. HaNPV treatment during larval stage reduced the fecundity of emerged adults up to 20% and egg hatchability up to 30%.

#### INTRODUCTION

Pulses form an integral part of the vegetarian diet in the Indian sub-Besides being a very rich source of protein, pulses maintain soil continent. fertility through biological nitrogen fixation by bacteria prevalent in their root nodules, thus play a vital role in sustainable agriculture. Chickpea (Cicer arietinum L) is an important food legume crop in the production system of Semi Arid Tropics. Chickpea ranks second among the pulses in World. Of the 11 m. ha. of chickpea grown world wide, about 75% is grown in South India is the world's leading producer of chickpea with 68% of the Asia. total production. But the current productivity levels of pulses is low, 200-700 kg/ha. It's productivity, however is limited by a complex of interacting biotic and abiotic factors. Among biotic stresses insects are known to be the prime constraint in chickpea production throughout Asia. Of the several insect species that attack chickpea the gram pod borer Helicoverpa armigera Hubner is one of the major biotic constraints (Srivastava and Srivastava, 1990).

Gram pod borer is a prolific and wide spread pest, which feeds on at least 180 plant species spread across 47 botanical families (Pawar *et al.*,1986). The biological characteristics such as high degree of polyphagy, high mobility, facultative diapause, high fecundity and multigeneration, contribute directly to the pest status of *H. armigera* (Fitt, 1989).

So far, use of insecticides has been the major approach for controlling this pest in different crops in India and in most of the developing countries. Chemical control is one of the effective and quicker methods in reducing pest population, where farmer obtains spectacular results within a short period. However over reliance and indiscriminate use of pesticides for longer periods resulted in a series of problems, mainly risk of environmental contamination, loss of biodiversity which contributed to the development of insecticide resistant *H.armigera* population, resurgence, out breaks of the secondary pests into primary pest status, distruction of natural enemies, increase in inputs on chemicals and toxicological hazards due to pesticide residue etc., (Armes *et al.*, 1992).

Any single method of approach to pest control may not be feasible, hence the best alternative is Integrated Pest Management approach, which is based on the principles of managing the pest rather than aiming at complete eradication: In view of this, extensive studies are in progress to develop IPM combining all possible components like use of resistant varieties, cultural & mechanical control, biological suppression, chemical control, behavioural approaches etc., (Jayaraj, 1992) which ultimately reduce the negative influence of insecticides on the natural enemies, that are present in the suitable ecological niche and will protect the ecosystem and the environment from toxicological hazards.

A major contribution of IPM to agriculture has been to demonstrate the need to base all phases of the production system on sound ecological principles, with the ultimate goal of 'designing' agro- ecosystem that is economically and ecologically sustainable. The information available on cultural, varietal, biological and chemical methods of pest control has been critically reviewed in view of significant advances made so far in chickpea pest management strategies such as mixed or intercropping, host avoidance, use of sex pheromone traps, neem seed kernel extract and use of insect pathogen against the gram pod borer, *H.armigera* which have generated enough scope to begin with IPM in chickpea (Lal, 1992). Heavy use of highly toxic and persistent pesticides year after year reduced the population of a potent indigenous endoparasite *Campoletis chlorideae* Uchida culminating into heavy out breaks of *H.armigera* in several gram growing areas (Odak, 1982), but the availability of information on the effects of botanicals, viral pathogens and other IPM components individually and in combination on natural enemies present in chickpea crop is limited. Major obstacle in the use of insect viruses in field situations is the rapid inactivation by ultraviolet radiation (Ignoffo and Garcia, 1992). Robin blue is one of the popular UV rays protectant (Rabindra and Jayaraj, 1988), but the information on efficiency of robin blue in increasing the efficacy of HaNPV under field conditions is limited.

Hence, the present study is mainly devoted to generate information on the effect of different IPM strategies, cultural, biological and chemical, individually and in combination on the chickpea pod borer and its natural enemies, the various effects of neem and HaNPV on life cycle of *H.armigera* and the efficiency of robin blue as a UV rays protectant. The studies are contemplated with the following objectives.

- 1. To evaluate the effect of IPM components on Helicoverpa armigera.
- 2. To study the impact of IPM components on natural enemies in chickpea.
- 3. To study the effect of neem on oviposition deterrency and antifeeding activity of *Helicoverpa armigera*.
- 4. To evaluate the efficacy of Robin blue as UV protectant to improve the persistence of HaNPV.

## **REVIEW OF LITERATURE**

#### Helicoverpa armigera Hubner as a Pest of Chickpea

Among seventeen species placed in Helicoverpa, H.armigera, H.punctigera and H.assulta are exhibiting higher fecundity, wide host range and greater migratory tendencies (Barrett, 1967). H.armigera is widely distributed from Southern Europe through Africa, Asia and Australia to the south-western pacific islands (Hardwick, 1965). About 87% of the worlds chickpea crop is grown in South Asia (Jodha and Rao, 1987) and Helicoverpa armigera Hubner is one of the most serious pests of grain legumes, especially chickpea and causes up to 85% pod damage in different states with an average of 8% (Sithanantham et al., 1984) and up to 91.7% in Punjab (Chhabra, 1990). H.armigera attains peaks twice in a year i.e March-April and October (Chhabra, 1990) and the population was positively correlated with maximum and minimum temperatures and negatively correlated with Relative Humidity and % parasitism by Campoletis chlorideae Uchida (Yadava and Lal, 1988). According to Joginder Singh et al. (1990) the nondiapause type of H.armigera completed two generations between 5th November and 5th April compared with one generation for diapausing H.armigera.

#### Efficacy of IPM Components Against H.armigera

#### Neem

Azadirachtin, a tetrah (or) triterpenoid is the most active insecticidal component found in neem seeds and leaves (Butterworth and Morgan, 1968). This active component has a number of biological properties including repellency, feeding and oviposition deterrency, hormone like growth disrupting activity and low mammalian toxicity (Schmutterer, 1990). Neem seed extracts or their components have systemic property and are

translocated within plants (Saxena, 1987; Morian *et al.*, 1990). Unlike ordinary insecticides based on single active ingradients, derivatives of neem comprise a complex array of novel compounds which have diverse behavioural and physiological effects on insects (Saxena, 1989). Pesticides derived from neem tree *Azadirachta indica* A. Juss. appear to be promising for use in IPM programs and provide broad spectrum control of more than 200 species of insect pests (Ascher, 1993) and safe in pest control programe and may prevent several adverse effects caused due to application of synthetic insecticides (Rajasekaran and Kumaraswamy, 1985).

Odak (1982) tested different plant products and concluded that neem seed kernel extract 5% had lowest pod damage (3.1%) due to Harmigera in chickpea. Neem seed kernel extract 2% and 2% hot pepper fruit extract gave good protection of bean from Maruca testulalis (Geyer) and H. armigera and other important insect pests (Hongo and Karel, 1983). Thakur et al., (1988) reported that on the basis of grain yield, endosulfan 0.07% was the most effective treatment followed by monocrotophos 0.04% & neem leaf extract 5% and on the basis of profitability neem leaf extract 5% was the most effective followed by endosulfan 0.07% and monocrotophos 0.04% treatments. It was concluded that neem seed kernel extract can be used in place of highly toxic synthetic insecticides because of its safety to beneficial insects and its lower cost against H. armigera in chickpea. Sinha and Mehrotra (1988) reported that application of neem oil (at 375, 560, 750 & 3750 ml / ha) in chickpea field against H. armigera at flowering and 10 days after did not give a significant effect in reducing the larval population and the incidence of damage but gave higher yield of seed than untreated control.

According to Sehgal and Ujagir (1990) neem seed kernel extract at 5% was less effective against *H armigera* in chickpea than endosulfan 40° g at /ha but still significantly better than the control. According to Sachan and Lal (1993) neem seed kernel extract and neem leaf extract were effective for controlling the noctuid, *H. armigera* on chickpea and pigeonpea. Datkhile et al. (1992) reported that neem seed extract at 5% was the least effective on gram pod borer when compared to synthetic pyrethroids. Butani and Mittal (1993) reported that malathion, DDT and neem seed kernel suspension were all equally effective in controlling *H armigera* in chickpea and increasing grain yield. According to Sinha (1993) when infestation of *H armigera* in chickpea reached 20.5% (1986-87) and 12.5% (1988-89) spraying with diflubenzuron (0.05%), neem oil and kernel extract 5% at an interval of 10 days gave a 2-fold reduction in infestation compared with the untreated control.

Sinha (1993) also reported that during 1989 and 1990–91 neem emulsion & neem wp gave 40–60% control. Neem kernel extract 5% gave 40% reduction in infestation of *H.armigera* in chickpea and was comparable to endosulfan 0.07% (1989-91). It was reported that there is no significant difference in the seed yield in plots treated with neem emulsion 0.125%. neem kernel extract 5%, flufenoxuron 0.01% and endosulfan 0.07% against *H armigera* in chickpea. Sarode *et al.* (1995) reported that NPV and neem seed kernel extract gave better control of *H.armigera* on chickpea when applied in combination than when applied singly. Khan (1996) reported that neem seed extract 5% and Nimbecidine 0.2% recorded 21.9 Q/ha and 19.6 Q/ha seed yield of chickpea by reducing *H.armigera* infestation which are comparable with other treatments viz. cypermethrin + profenofos 0.088%. monocrotophos 0.04%, profenofos 0.2% and chlorpyriphos 0.05%. According to Ravi and Verma (1997)<sup>b</sup> azadirachtin was the least effective insecticide compared to fenvalerate, endosulfan and diflubenzuron in reducing *H* armigera in chickpea.

In a laboratory study Nimbecidine gave 20 2% egg mortality of *H* armigera where as endosulfan gave 41 1% (Usha and Patel, 1997). Jeyakumar and Gupta (1999) noticed ovicidal effect of NSKE in different age groups of eggs of *H.armigera* and mortality decreased with increase in age group of eggs. According to Ujagir *et al* (1997) azadirachtin (Nimbecidine 0.03%) did not show any yield increase by reducing the pod damage caused by *H.armigera* when compared to either HNPV or chemical insecticides in chickpea. Murugan *et al.* (1998) reported that the neem limonoids azadirachtin, salanin, deacetyl gedunin, 17-hydroxy azardiradione and deacetyl nimbin were found to be potent antifeedants and growth inhibitors to the cotton boll worm *H.armigera*. Padmaja and Rao (2000) recommended three plant oils including neem oil as a potential control measure for the management of the American bollworm, *H. armigera* on the basis of ED50 dose.

#### HaNPV

Anita Mistry *et al.* (1984) reported that five sprays of NPV @ 250 LE / ha / week gave satisfactory control of *H.armigera* in chickpea and increased in grain yield upto 47% over control. Dhamdhere and Khaire (1986) evaluated different doses of HNPV on *Cicer arietinum* L against *H.armigera* and concluded that two applications of 450 LE / ha at a 10 day interval were most effective in reducing the damage and resulted in the highest yield. Jayaraj *et al.* (1987) reported that *H. armigera* population in chickpea was significantly reduced with an application of 250 LE / ha HNPV and the virus was more effective when sprayed in the evening than in the morning. When the virus was applied

with 2% starch or 1% sugar there was no difference between morning and evening applications. Pawar et al. (1987) concluded that 2 sprays of HNPV @ 500 LE / ha were as effective as 2 sprays of 0.05% endosulfan in reducing infestation by *H. armigera* and pod damage in chickpea and in increasing seed yield. Bilapate et al. (1988) recorded 6.9, 1.9 and 24.5% mortality of *H. armigera* due to NPV during 1<sup>st</sup>, 2<sup>nd</sup> and 3<sup>rd</sup> generations, respectively

According to Pawar et al. (1990) the lowest pod damage and highest yield were observed in plots treated with 2 sprays of endosulfan 0.07% or 2 sprays of HNPV @ 500 LE / ha or with one spray of HNPV @ 500 LE / ha followed by one spray of endosulfan 0.07% against *H. armigera* in chickpea. Chundurwar and Pawar (1991) described mortality of *H. armigera* infesting chickpea in Maharashtra, India due to natural infection by a nuclear polyhedrosis virus. Rabindra *et al.* (1992) evaluated the effect of HNPV on different varieties of chickpea against *H.armigera* and concluded that control of *H.armigera* on chickpea with HNPV was significant on the highly susceptible or moderately susceptible (Co 2) varieties than on the tolerant variety(ICC 506). A single spray of 250 LE / ha of HNPV in 500 litres of water per hectare resulted in 97.2% mortality of *H.armigera* in 1987 and 25.4 to 78.8% larval mortality during 1988 in chickpea (Misra *et al.*, 1991)

Elcar (HNPV) and Dipel (*B.t.*) were not as effective as cypermethrin in controlling *H.armigera* infestation in chickpea but yield was significantly high in bio-insecticidal treated plot than control and was statistically on par with cypermethrin treated plot (Ibrahim Ali *et al.*, 1993). According to Sarode *et al.* (1995) HNPV @ 500 LE / ha recorded the lowest infestation of *H.armigera* on cotton followed by NSKE 6%, hence recommended in IPM system. NPV had greater effect on the density of *H.armigera* large larvae on susceptible chickpea genotypes than on ICC 506 variety and the yields were also significantly higher in NPV treated susceptible genotype compared to quinalphos or control plots (Cowgill and Bhagwat, 1996). Abhisek Shukla and Goydani (1996) reported that application of HNPV for the control of *H. armigera* infesting chickpea produced a significantly higher seed yield compared to control but low compared to plots treated with endosulfan Sharma *et al.* (1997) assessed different biopesticides and chemicals for control of *H. armigera* in chickpea and concluded that HNPV gave the best control compared to other biopesticides and chemicals.

#### **Biological control**

Achan et al. (1968) reported Campoletis chlorideae Uchida as a larval parasite of Harmigera. The activity of C chlorideae brings an appreciable reduction of Heliothis larvae and crop losses in chickpea (Bhatnagar and Davies, 1978). First record of C.chlorideae on H.armigera in Bihar to the extent of 14.3 to 58% was reported by Prasad and Chand (1986). Inundative release of Trichogramma chilonis Ishii to control the noctuid, H.armigera was ineffective in gram (Yadava et al., 1985). Parasitism by the ichneumonid C.chlorideae in H.armigera on chickpea was highest during December, lowest during February and almost nil during March (Yadava, 1990). The larval parasites of Harmigera, braconids. Apanteles sp., Bracon sp. and Microbracon sp., the ichneumonid, C.chlorideae and the eulophid Euplectrus euplexiae were reported by Singh and Balan (1986). Mehto et al. (1986) recorded eight species of natural enemies on H.armigera in chickpea, which included C.chlorideae, Araneae, Coccinella spp., Chrysopa spp and Pantala spp., ICRISAT (1987) reported parasitism of H.armigera in chickpea by C.chlorideae and Carcelia illota Curron in Andhra Pradesh. H.armigera population in chickpea was

negatively correlated with per cent parasitism by *C chlorideae* (Yadava and Lal. 1988). Srinivas (1989) studied seasonal incidence of *C chlorideae* and *Eriborus* sp. on *H. armigera* in chickpea, and found peak parasitization by *C.chlorideae* in first two weeks of December (43.9%) and by *Eriborus* sp during last week of January (43.8%). Early instars of *H armigera* were more prone to attack by various enemies (Srinivas and Jayaraj. 1989) Garg (1989) observed 25% parasitization of *H.armigera* by *C chlorideae* Shrivastava and Yadav (1991) recorded 61.9% and 16.66% parasitization of *H armigera* by *C. chlorideae* at Kawardhe and Amora areas of Madhya Pradesh, respectively in chickpea. The ichneumonid *C. chlorideae* and tachinid. *C.illota* and the braconid, *Apanteles* spp. played a key role in suppressing the larval population of *H. armigera* in chickpea during podding stage (Patnaik *et al.*, 1991; Mishra *et al.*, 1992 and Ahmed *et al.*, 1996).

A German ornithologist estimates that a single pair of tits with their progeny destroy annually at least 120 million insect eggs or 150,000 caterpillars and pupae (Ali, 1996). Ghode et al. (1988) observed the avian predation of gram pod borer H.armigera in Orissa and reported that the cattle egret (Bubulcus ibis Lin.) and river tern were feeding Harmigera on bengalgram in the third week of January. Due to the presence of the birds, the population of H.armigera was reduced from 5-10 larvae/plant in the middle of January to a negligible number (<1/plant) by the end of the Patel (1988) conducted studies on predation of Harmigera and month. Spodoptera litura Fab. by insectivorous birds with special emphasis on mynas Acridotheres tristis (Lin.). Joginder Singh et al. (1990) mentioned the importance of house sparrow and myna as natural enemies of H.armigera in Ludhiana. In Kota, Rajasthan the house sparrow reduced Helicoverpa population by 20 to 40% (ICAR, 1992). Wightman et al. (1993) suggested that predation by cattle egret might be increased by giving the birds easy access to the larvae by sowing on ridges or by optimizing row separation in a flat sowing.

#### Endosulfan

Daware and Dhanorkar (1981) reported that several conventional insecticides such as endosulfan, monocrotophos, quinalphos, etc. were found effective against *H. armigera*. According to Dhurve and Borle (1985) permethrin 0.01% followed by carbaryl 0.2% and endosulfan 0.05% were effective in reducing the damage caused by *H armigera* in chickpea and recorded significantly higher yields. Three sprays of 0.05% endosulfan or 0.05% quinalphos at 15 day intervals commencing at 50% flowering stage gave most effective and economic control of *H.armigera* in chickpea (Rizvi et al., 1986)

Jayaraj et al. (1987) compared efficiency of NPV, endosulfan and its combination in controlling *H. armigera* in chickpea and reported that mixture of NPV (125 LE / ha) and endosulfan (0.035%) resulted in maximum protection. But an application of virus @ 250 LE / ha followed by endosulfan 0.035% five days later was on par with 0.07% endosulfan. Two sprays of HNPV @ 500 LE / ha were as effective as two sprays of 0.05% endosulfan in reducing infestation by *H. armigera* larvae and pod damage and increased yield in chickpea (Pawar et al., 1987). Gunasekaran and Balasubramanian (1987) reported that endosulfan @ 525 g a.i. / ha resulted in reduction of 75.2, 87.6 & 98.2% of *H. armigera* at 1, 3 and 7 days after application on chickpea. Sanap and Deshmukh (1987) tested seven insecticides for the control of *H. armigera* on chickpea, among which 0.07% endosulfan resulted in the least pod damage (1.4%) and highest yield (1209 Kg / ha). According to Thakur et al. (1988) on the basis of grain yield and profitability endosulfan at 0.07% was the most effective treatment in

controlling Harmigera on gram followed by monocrotophos at 0.04% and neem kernel extract at 5%. On the basis of mean per cent damage at the dry pod stage and grain yield, fenvalerate 0.02% and endosulfan 0.07% were the most effective treatments against Harmigera on gram (Kaul et al., 1988). Among various insecticides tested endosulfan 0.07% spray gave maximum protection against H. armigera in standing crop of bengalgram (Jain and Singh, 1988). Parsai et al (1989) tested eleven insecticides against Harmigera on chickpea and reported that 0.05% monocrotophos and 0.07% endosulfan were highly effective One spray with endosulfan at flower bud formation to reduce H. armigera on chickpea achieved 61.1 to 81.1 % mean reduction of larvae at different locations and 60.0 to 87.5% avoidable loss in grain yield (Singla et al., 1989). According to Deka et al. (1989) endosulfan at 500 g a.i. / ha was the most effective of five insecticides tested in reducing larval population of H. armigera by 94.4% at 72 hrs after spraying and in obtaining yield increase of 159.03% compared to untreated control in C. arietinum. Endosulfan spray gave good control of H. armigera in chickpea (Ghosh et al., 1989).

Sehgal and Ujagir (1990) reported that endosulfan at 420 g a i. / ha significantly and consistently reduced pod damage by *Helicoverpa* to < 22.5% from 65.5% and increased grain yields to > 1.7 t / ha from 0.7 t / ha in chickpea and not shown any phytotoxic effect when applied during flowering stage (Parsai *et al.*,1990). Endosulfan 0.07% followed by 0.06% endosulfan were the most effective treatments against *H. armigera* in chickpea (Gupta *et al.*, 1990). The lowest pod damage due to *H.armigera* (3.8%) and highest yield (1379 Kg / ha) were observed in plots treated with 2 sprays of endosulfan in chickpea. According to Gupta and Thakur (1990) monocrotophos 0.05%, fenvalerate 0.01% and endosulfan 0.08% gave good control of *H.armigera* larvae in chickpea and increased yield by

67-70% in November sown crop and by 103 - 113% in December sown crop. ULV spray of endosulfan @ 1 I / ha in bengalgram reduced *H* armigera to 4.40 larvae / 5 plants and 12.61% pod damage. Patel *et al* (1990) recommended one spraying of monocrotophos 0 04% at 50% flowering followed by endosulfan 0.07% 15 days later for irrigated chickpea

According to Barkhade et al. (1991) pod damage due to Harmigera on chickpea was the least with 4% endosulfan dusting at 30 days after flowering but spraying with 0.05% endosulfan at 10 DAF, dusting with 4% dust at initiation of flowering and 2 sprays of 0.05% endosulfan at 15 and 30 DAF gave similar effect. Greatest yields were obtained on different cultivars of chickpea treated with two applications of endosulfan 0.07% during the pod formation stage (Chauhan and Dahiya, 1991). According to Gupta et al. (1991) among different spray schedules sequential spraying at the flowering followed by podding stage with endosulfan 0.07% is the most effective in terms of cost ; benefit ratio, 12 ; 1 in chickpea. Khan et al. (1993) tested different insecticides against noctuid Harmigera on gram. applied at pod formation and at 15 days later and concluded that endosulfan and cyfluthrin were most effective while endosulfan recorded the highest average yield of 32 Kg / plot compared to untreated control According to Sinha (1993) NSKE 5% was comparable to endosulfan 0.07% against H.armigera in chickpea during 1989-90, where as endosulfan 0.07% gave 72% control during 1990-91. In a laboratory study endosulfan 0.07% gave 100% inhibition of H. armigera egg hatching (Mala et al., 1993).

Two applications of endosulfan against the gram pod borer on *C.arietinum* recorded average larval population of 0.88 per plant as against 2.6 in control and yield was 1573 Kg / ha against 251 Kg / ha in no treatment (Noorani *et al.*, 1994). Two sprays of endosulfan at 50%

flowering followed by 2 sprays at the green pod stage effectively controlled *H.armigera* on chickpea (Giraddi *et al.*,1994). Endosulfan 0.07% significantly reduced *H.armigera* in chickpea and recorded the highest grain yield irrespective of sowing dates (Chaudhary and Sachan, 1995). According to Vyas and Lakhchaura (1996) endosulfan 0.07% applied twice was superior compared to monocrotophos and HNPV in controlling *H.armigera* on chickpea.

#### IPM

In the past 30 years the fundamental paradigm that emerged in plant protection is IPM. A major contribution of IPM to agriculture has been to demonstrate the need to basic all phases of the production system on sound ecological principles, with the ultimate goal of designing agroecosystems that are economically and ecologically sustainable. During the last two decades considerable amount of work has been carried out on the use of parasitoids (Nagarkatti, 1982), predators (Greathead and Girling, 1982; King *et al.*, 1982), microbial insecticides including nuclear polyhedrosis virus (Tinsley, 1979; Bell, 1982; Mc Kinley, 1982) and neem extracts (Thakur *et al.*, 1988; Rao *et al.*, 1990) in pest management. However no single method of control can be expected to provide an acceptable solution to pest management. The discipline of integrated pest management (IPM) has been built on the philosophy of total system consideration and multiple control techniques.

Reed and Pawar (1982) reviewed the management strategies and approaches to manage *H.armigera* on chickpea which covered population studies through pheromone and light traps, use of insecticides, NPV, parasitoids, cultural practices and breeding for host plant resistance. Pawar *et al.* (1987) reported that population of *H.armigera* in chickpea is

the lowest in plots which received two applications of 0.05% endosulfan followed by those treated with virus only @ 500 LE / ha and a treatment in which application of 500 and 250 LE / ha were followed by endosulfan 0.05% sprays. According to Jayaraj et al. (1987) an application of HNPV @ 250 LE / ha followed by endosulfan 0.035% 5 days later was on par with 0.07% endosulfan for the control of Harmigera on chickpea. None of the various IPM components like natural enemies including C chlorideae. NPV, inter-cropping system and altering sowing dates were superior to recommended pesticides in controlling Harmigera on chickpea and pigeonpea (Mahajan et al., 1990). Pawar et al. (1990) observed the lowest pod damage of, 3.84% and highest yield of, 1379 Kg / ha of chickpea in plots treated with 2 sprays of endosulfan 0.05% alone or NPV @ 500 LE / ha or one spray of NPV @ 500 LE / ha followed by one spray of endosulfan 0.05%. Six years of experimentation revealed that NPV + two sprays of endosulfan (0.035%) at 1st and 3rd week of the crop recorded less pod damage and maximum yield against Harmigera in chickpea (Thakur, 1990).

Pimbert (1990) reported the themes that call for more research attention for IPM of *H. armigera* in chickpea like host plant resistance and G x E interaction, vegetation management and biological control, IPM and the selective use of plant diversity, biotechnology and pest control, group action to complement pest controls aimed at individual house holds and sustainability. Ahmed *et al.* (1990), Weigand & Tahhan (1990) and Sithanantham (1987) reviewed various aspects of *H.armigera* management on chickpea and also covered population studies through pheromone traps, insecticide use, use of bacteria, viruses and parasitoids, cultural practices and host plant resistance and breeding and integration of control methods According to King and Sawicki (1990) all the IPM desiderata of increased use of resistant or tolerant cultivars, timely pesticide applications targeted against neonate larvae based on scouting and economic thresholds and rotation of insecticides especially the synthetic pyrethroids can be used for Harmigera resistance management. ICRISAT, AICRIP and Directorate of Pulses Research conducted surveys and given overview of the biological and ecological aspects of the H.armigera in chickpea and pigeonpea and pest control measures which include use of pheromone traps, parasitoids (C chlorideae), predators (Delta spp.) and HNPV, breeding for HPR. advancing the sowing date or using early maturing varieties, mixed or intercropping with cereal / other legumes, use of phosphate fertilizers and application of insecticides (Sachan, 1990). According to Lal (1990) and Yadava (1990) H. armigera is an important pest of chickpea and pigeonpea in U.P. India. Use of insecticides, NSKE, pheromone traps, growing early maturing cultivars or advancing the sowing date, opting for resistant varieties, use of parasitoids and pathogens (NPV) were considered effective in controlling this pest in this state.

Sachan and Lal (1993) reported that use of 250-375 LE/ha NPV alone or incombination with endosulfan 0.035% has given 60-80% and NSKE at 5% has given 50–70% mortality of *H.armigera* in chickpea According to Jayaraj (1992) excessive reliance on chemical control method alone for the effective management of *H.armigera* has led to several problems and reported that the use of NPV in combination with Jaggery, teepol, etc., pheromone and light traps for monitoring, inundative release of parasites, application of NSKE 5% were good for control of *H. armigera* in <sup>-</sup> pulses. Sarode *et al.* (1995) reported that application of HNPV @ 500 LE/ ha plus the neem extract at 6% gave the maximum reduction in *H.armigera* larval number than when applied singly in chickpea crop. Three insecticidal applications during the season based on thresholds in an on-farm chickpea fields in susceptible (Annegiri & ICCC 37) and resistant (ICC 506) varieties resulted in a threefold increase in yield (Wightman *et al.*, 1995)

According to Sarode and Sarnaik (1996) *H.armigera* damage caused during flowering and podding stage results in substantial losses i.e. about 30 – 100% avoidable yield loss and also reported that adverse effects of chemical control led to switch onto the IPM programme, in which HNPV and NSKE were found to be effective, the addition of half doses of insecticides to these have been reported to improve their efficiency

Yadava (1996) conducted chickpea onfarm trials in Nepal during 1992-95 and compared improved agronomic package i.e., seed treatment with thiram + bavistin, hand weeding at 25 DAS and spraying Thiodan or Decis against Heliothis with farmers practices and reported high yield by 16-87% and significantly increased net returns. Sanap and Pawar (1998) conducted a field experiment in Maharashtra during 1993-96 for controlling H.armigera infesting gram. IPM treatment comprising endosulfan 0.07%. NSKE 5% and NPV @ 250 LE / ha were evaluated, and the results revealed that 3 spray applications starting from initiation of flowering and subsequent 2 sprays at 15 days interval with first 2 sprays either with NPV or NSKE followed by a 3rd spray with endosulfan were the most effective and resulted in a 26.9% and 27.3% increase in yield, respectively. According to Prasad and Singh (1997) chickpea sown on 25th September produced more grain yield and had a lower incidence of Harmigera compared to sowing on 10th October. According to Bhagwat (1997) an integrated pest management strategy using a botanical insecticide, host specific virus to protect chickpea from pod borer showed better efficacy of the approach over local farmers practices in onfarm situations.

The integrated pest management components *T.chilonis*, *Chrysoperla carnea* Stephens, HNPV. Nimbecidine, Dipel and synthetic chemicals were imposed at different interval on the basis of pheromone trap threshold level on a consolidated block of 40 ha cotton fields at two locations. Shankaraband and Kurlagundi. The results demonstrated a significant superiority of the IPM strategy in terms of both cost versus benefit and environmental safety over that used in the farmers fields where only conventional control methods were followed (Reddy and Manjunatha, 2000)

#### Effect of IPM Components on Natural Enemies

#### Neem

Parmar (1993) recommended use of neem in IPM as it was found relatively safe to natural enemies. Li et al. (1986) tested 29 insecticides including B.t and neem oil in order to study their side effects on Trichogramma japonicum Ashmead and concluded that neem oil and Bt were the safest pesticides for the parasitoid. Aqueous NSKE 2% had no influence on oviposition of the egg parasitoid Telenomus remus Nixon (Joshi Neem products showed little affect on T chilonis (Malathi et et al., 1982). al., 1999). Markandeya and Diwakar (1999) reported that when Harmigera eggs were treated with Margosan 1500 ppm 10 ml/l, T. chilonis parasitised 45% eggs as compared to 97.4% in control but not affected hatching Neem seed oil at 0.3% deterred oviposition (parasitization) by the parasitoid T.chilonis (Raguraman and Singh, 1999). Spraying of high concentration of AZT-VR-K on adult braconids and their contact with sprayed cabbage leaves . for 2 days has no obvious effect on the wasps (Schmutterer, 1992). Cano and Gladstone (1994) studied the influence of the NSK based extract NIM-20 on parasitization of eggs of the H.zea in a melon field by T. pretiosum and concluded no negative effect.

Neem oil was the safest pesticide for spiders, mainly Lycosa pseudoannulata (Bosenberg & Strand) as compared with three synthetic products (Wu, 1986) and in comparision with endosulfan (Fernandez et al. Serra (1992) also did not observe any adverse effects of a 4% 1992) aqueous NSKE on unidentified spiders in tomato fields The commercial products Margosan-O <sup>™</sup>, Azatin <sup>™</sup> and RD9 Repellin showed no toxicity to the spider fauna (Mansour et al., 1993). Breethaupt (1995) in corn fields and Saucke (1995) in cabbage fields reported no harmful effects to spider Oxyopes papuanus when NSKE 2% or Neem Azal-S applied Markandeva and Diwakar (1999) also reported that Margosan, a neem product did not affect the survival of wolf spider, Lycosa pseudoannulata. Feeding of the adults of the earwig Doru taeniatum using larvae of Spodoptera frugiperda (S. & A.) confined for 2 -4 days on corn leaves treated with AZT-VR-K a neem product, did not cause mortality of the predators (Hellpap, 1985). The cricket Metioche vittaticollis was not affected by neem seed bitters containing Aza and other active ingredients at 10,000 ppm in field trial (Lamb and Saxena, 1988). According to Fernandez et al. (1992) in a trial with 3% neem oil, 5% aqueous NSKE, endosulfan and water as control, all the mirid bugs died in endosulfan treatment but no mortality was recorded in the other treatments.

In a laboratory experiment, adults of the coccinellid, kept on neem oil treated glass plates according to IOBC/WPRS guidelines did not show increased mortality or reduction of fecundity when compared to control, but metamorphosis of the larvae was interrupted (Schmutterer, 1981). Predaceous coccinellids survived when a formulation with high neem oil content was sprayed whereas the target pest, sorghum aphid was successfully controlled (Srivastava and Parmar, 1985). Treatment with Neemix 4.5 EC caused several abnormalities throughout the life cycle and

even in emerged adults also in *C. septempunctata* treated at immature stages but the LC50 values were much higher than recommended rate for pest control, hence can be safely used in IPM programmes (Banken and Stark, 1997and 1998) Maragosom 1500 ppm 10 ml / I. gave 6.7 and 5% mortality due to contact to grubs and adults of *Menochilus sexmaculatus* (Fabricius) (Markandeya and Diwakar, 1999).

Eisenlohr et al (1992) reported that the number of syrphid larvae was not reduced in the field after spraying with Neem Azal – F on peach trees infested by *Myzus persicae* (Sulzer), but the number of adults derived from larvae collected in the field on treated trees were reduced. Isman et al (1992) showed that neem had no detrimental effects on predatory syrphids. Lowery and Isman (1995) reported that the number of larvae of predacious cecidomyilds was reduced in the field after application of NSKE 14% and neem oil 1% as compared with control.

AZT-VR-K 1000 ppm (Kaethner, 1991) and Neem Azal – F (Vogt, 1993) did not show any side effects on the broad spectrum predator *C.carnea*. Schulz *et al.* (1997) indicated no negative effects of Neem Azal-T/S @ 3 I / ha on *C. carnea* and to honey bees. According to Srinivas and Sundara Babu (2000) various neem products caused egg and grub mortality of *C.carnea* and also affected longevity of adults.

Margosan – O proved to be nontoxic to honey bee workers up to a concentration of 4418 ppm AZ / ha (Schmutterer and Holst, 1987). Honey 'bee larvae are less susceptible to azadirachtin than most pest species (Neumann and Isman, 1996).
#### HaNPV

A parasite C chlorideae was found to transmit the NPV virus both directly (100%) and indirectly (50%) (Odak et al., 1982) Bijjur et al (1991) reported that Apis cerana indica did not show any signs of abnormal development when treated with NPV of H. armigera Ruberson et al (1991) found that a nabid predator Nabis raseipennis Reuter fed with NPV infected soybean looper larvae did not affect survival rate but had a shorter developmental time than those fed with healthy prey. Heinz et al. (1995) indicated that two common predators C carnea and Orus insidosus were not adversely affected by feeding on larvae of Heliothis virescens infected with recombinant Autographa californica (Speyer) NPV. Sajap et al. (1999) reported that when an assasin bug Sycanus leucomesus Walk was fed on NPV infected larvae of S. litura, it appeared normal but with smaller size of head capsule & shorter tibial lengths with 10% reduced survival, 12 days prolonged pre-oviposition period, reduced longetivity and fecundity of adults.

#### Endosulfan

In a laboratory study spraying of endosulfan 0.07%, monocrotophos 0.05%, phosalone 0.1% on *Trichogramma* parasitized eggs of *H.armigera*, the emergence of adult parasitoids were not affected (Santharam and Kumaraswami, 1985). Malathi *et al.*(1999) reported that endosulfan was relatively toxic on emergence of *T.chilonis*, oviposition behaviour but not on the further development. Heavy use of highly toxic and persistent pesticides year after year reduced the population of a potent indigenous endoparasite *C.chlorideae* culminating into heavy out breaks of *H.armigera* in several gram growing districts of Madhya Pradesh (Odak, 1982). Pawar *et al.*(1989) reported that the parasitism by *C.chlorideae* was lower in pesticide treated area compared to untreated control. Ravi and Verma (1997)<sup>b</sup>

recommended endosulfan for the control of Harmigera which showed little effect on the larval parasitoid C.chlorideae. Krishnamoorthy (1995) reported that endosulfan, dicofol, monocrotophos, phosalone, methyl demeton, phosphamidon, dimethoate, sulphur and dithane M-45, were found toxic to both larvae and adults of C carnea in a laboratory study. Both spider and ground beetle populations were known to be reduced by regular application of insecticides (Pfrimmer, 1964; Luff, 1987) Pyrethroid insecticides suppressed web building frequency and web size and building accuracy of spider Aroneus diadematus (Samu and Vollrath, 1992). Some group of invertebrates such as ground beetles (Carabidae) and spring tails (Collembola) had decreased substantially and persistently under the high input (prophylactic) pesticide regime (Cilgi et al., 1993). Kostandy (1995) reported that the population density of the predators decreased obviously in the fields treated with insecticides for controlling cotton bollworms. Frequent use of fungicides and insecticides reduced the abundance, activity and species diversity of spiders (Rayner et al., 1996). Endosulfan (367.5 g a 1 / ha) and dimethoate (120 g a.i. / ha) caused less reduction of theridiid spider and lacewing larvae, several coccinellids and Hemiptera compared to thiodicarb and methomyl (Wilson et al., 1998). According to Van den Berg et al. (1998) natural enemies generally have a high impact on Lepidoptera in unsprayed fields in Indonesia, but generalist predators seem to recover more slowly after insecticidal application than lepidopterans leads to more dependency on insecticides for the control.

#### Residues

Pandey et al. (1977)<sup>b</sup> with two sprays of 0.07% endosulfan at 600 l. / ha on bengalgram at pod formation stage for the control of the pod borer, observed that its residues were much higher than the tolerance limit even 25 days after spraying both on the plant and in the grain. Pandey

10-1-5-1-5 2 hours

et al (1977)<sup>4</sup> with two sprays of 0.07% endosulfan at 500 I / ha to pea crop at the pod formation stage, recorded 5.90 ppm residue at the time of harvest in the plant. There was an increase in the residues in the grain from 1.95 to 3.30 ppm starting from 3<sup>rd</sup> to 15<sup>th</sup> day after first application which was attributed to the translocation and accumulation of the toxicant in the grain. 3.3 ppm of residue recorded in the grain at the time of harvest

Verma (1983) studied the persistence of carbaryl, endosulfan, monocrotophos and chlorfenvinphos applied for control of pest complex of bengalgram and reported that endosulfan residue fell below the tolerance limit in 12 days and residues were persistent in leaves than in pods Gopal *et al.* (1988) analyzed the residue of stereo isomers of endosulfan and its toxic metabolites by GLC when applied at 0.07 and 0.14% and reported that the residue did not exceed the maximum tolerance level on the edible plant parts of brinjal and gram at the time of harvest. According to Singh *et al.* (1988) when endosulfan was applied @ 0.5 Kg a.i. / ha on sorghum, the initial deposit was 3.14 ppm this degraded to 1.17 ppm (below tolerance limit of 2ppm) within 5 days and fell below the detectable level after 20 days. The half life of endosulfan in sorghum was 11.95 days.

Singh et al. (1990) reported residues in soybean crop as 0.137. 0.913 and 1.947 ppm when treated with 0.05, 0.1 and 0.2% concentrations of endosulfan 8 days after treatment, but at the time of harvest the residue was below detectable limit both in grain and haulms. According to Parihar et al. (1990) in Jaipur, Rajasthan, waiting periods of 1.3 and 2.33 days should be observed before the green pods of chickpea can be consumed safely after having been sprayed with endosulfan at 500 and 1000 g a.i. / ha, respectively. The residue fell below the detectable limit 5 days after spraying at both doses. Dethe and kale (1991) analyzed residues of endosulfan by gas liquid chromatography in seeds of chickpea sprayed at twice than recommended dose and reported that the residues were at undetectable level. The residues of endosulfan resulting from three sprays in mustard seed at harvest by GLC was 0.065 and 0.145 mg/ kg for the recommended and double dosages, respectively (Udean *et al.*1991) Endosulfan could be used for the control of insect pests of pigeonpea without problem of excessive residue in the grains (Chawla and Joia, 1992) According to Senapathi *et al.* (1992) residues were concentrated in the husk than in the grain and also recommended that neither grains nor husk are consumed following application of quinalphos and monocrotophos, but the grains may be safely consumed after treatment with endosulfan

Gopal and Mukharjee (1993) determined the residue of endosulfan on egg plant, mustard and chickpea and reported that alpha isomer was degraded more rapidly than the beta isomer, Beta isomer accumulated during the first 3 days following treatment. The total endosulfan residues in seeds from the treated mustard was 0.08 to 0.12 mg / Kg and were at or below the limits of detection (0.02 mg / Kg) in chickpea seeds following harvest. Ravi and Verma (1997)<sup>a</sup> recommended mixture of diflubenzuron with endosulfan for the control of *H.armigera* in chickpea and suggested safe interval before consumption for endosulfan on chickpea as 4 days Naseema Beevi *et al.* (1997) reported that when endosulfan @ 0.07% and 0.14% during flowering and pod formation stage was applied in cowpea the residues were dissipated to below detectable level on 15<sup>th</sup> day in low dose while it was 0.3 mg / kg in higher dose. According to Tanwar and Honda (1998) the half lives of foliar application of endosulfan at 350 and 700 g

a i / ha during rainy season were 42 and 47, days respectively on foliage and 5-8 and 5 days, respectively on pods. Terminal residues at harvest on pigeonpea pods and pod covers were 1.73 and 2.45  $\mu$ g / gram at recommended and double concentrations, respectively, corresponding values for grain were 0.43 and 0.79  $\mu$ g / gram, respectively.

#### Cost Benefit ratio

Parsai *et al.* (1989) tested eleven insecticides against *H armigera* on chickpea and reported that the highest cost benefit ratio (1.5.1) was obtained with endosulfan. According to Datkhile *et al.* (1996) endosulfan 0.07% recorded 5.3 and NPV 250 LE / ha 2.6. Neemark 0.2% 2.7 and NSKE 1.2 cost benefit ratios in chickpea against gram pod borer. Patel *et al.* (1997) reported that endosulfan 0.035% gave the highest incremental cost benefit ratio (1:14.1), followed by endosulfan wp 0.035% (1.12.9) and chlorpyriphos 0.02% (1:12.1). NSKE suspension 3% gave 1.11.7 cost benefit ratio which was less effective and economical for controlling *H.armigera* in pigeonpea.

#### **Oviposition Deterrency Effect of Neem**

Fagoonee (1981) reported that crude alcoholic extracts of dried neem leaf repelled *Crocidolomia binotalis* Zeller female from treated cabbage leaves at a distance of about 25 cm. In *H. armigera*, the volatiles of neem seed kernels and their aqueous distillates offered at a distance prevented contact and repelled the moths (Schmutterer, 1990). Saxena and Rembold (1983) found that azadirachtin alone neither repelled *H* armigera adults nor deterred egg laying but due to presence of organosulphur compounds including dipropyl disulphide helped in repelling adults and deterred egg laying (Balandrin *et al.*, 1988). Oviposition deterrency effect of azadirachtin was reported by Schmutterer (1990)

According to Murugan *et al.* (1995) neem extract had both antifeedant and antiovipositional effects on the *Helicoverpa* moths. Jeyakumar and Gupta (1999) reported that neem seed kernel extract 10 and 7.5% treatments reduced oviposition to 60.9 and 59% compared to control

In the sheep blowfly, *Lucilia sericata* neem oil and the formulated NSKE AZT-VR-K were powerful ovipositional deterrent agents especially AZT-VR-K provided 100% repellence at 0.02% (Rice *et al.*, 1985) According to Neumann and Isman (1995) 1% crude neem oil emulsion significantly reduced the proportion of eggs laid by *S.litura* on treated cabbage plants

#### Neem Effects on H.armigera

Neem azadirachtin besides being an antifeedant has been shown to interfere with growth, moulting and ecdysis (Rembold and Sieber 1981), reproduction (Dorn *et al.*, 1986) and mortality (Rembold *et al.*, 1981) of various insects. NSKE at higher concentrations reduced egg production and hatching percentage of many insects (Brattson, 1983). Neem has adverse effects on ovarian development and fecundity and fertility of adults (Karnavar, 1987), effective against nearly 200 insects and mite species (Saxena, 1989). Neem affected growth and feeding rates at 5-20 ppm in many laboratory experiments against a variety of insect species (Mordue and Blackwell, 1993).

Reduced fertility/fecundity in *H.armigera* due to NSKE was observed by Joshi and Sitaramiah (1979). Neem extracts have been observed to effect the morphology and development of chickpea pod borer *H.armigera* (Jabbar *et al.*, 1988). According to Tahir Anwar *et al.*, (1993) topical application of neem oil 0.001% on thoracic region of the *H.armigera* larvae had no significant effect on the longevity of both the sexes. Murugesan and Jacob (1994) reported that with increase in NSKE on *Heliothis armigera* and *S.litura* food intake gets reduced, growth becomes slower and moulting is inhibited.

The El<sub>95</sub> Values for azadirachtin (doses for 95% ecdysis inhibitory activity) for first instar larvae of *H.zea* and *H.virescens* were 2ppm, for *S.frugiperda* 1ppm and for *Pectinophora gossypiella* Saund 10ppm (Kubo and Klocke, 1982).

According to EL-Sayeed (1985) 0.2-0.5% suspension of ground neem seeds caused 100% mortality of *S.littoralis* by the end of the larval period, and also caused pupal mortality and adult deformity. Antifeedant and growth inhibitory effects of neem limnoids in *S.litura* were repoted by Murugan and Jeyabalan, (1995); Koul *et al.* (1996). Application of azadirachta extract affected behaviour and vitality of larvae and adults of *S.frugiperda* (Breuer and Schmidt, 1996).

#### Neem Antifeedant Effects

Pradan et al. (1962) were among the first to report that extracts from seeds of neem Azadirachta indica A.Juss. were antifeedant to the desert locust Schistocerca gregaria Forskal.

According to Schoonhoven *et al.* (1987) contact with azadirachtin makes disruption in food intake and increases the locomotory activity of insects. Neem extracts had both antifeedant and antiovipositional effects on the *Helicoverpa* moths (Murugan *et al.*, 1995). In studies on the feeding behaviour of larvae of lepidopterous insects such as *S.littoralis*, *S.frugiperda*, *S.exempta*, *H.virescens*, *H.zea*, *H.armigera*, *Trichoplusia ni* (Hb.) and *M.brassicae* azadirachtin reduced feeding (Schmutterer, 1990).

Neem oil possesses distinct antifeedant activity against cotton bud worm *S.littoralis* (Koul, 1987). Chen *et.al.* (1996) reported that neem seed kernel extract at concentrations ranging from 0.2-4.0% reduced the number of eggs from 87.5 to 99.2% compared with eggs in untreated guava fruit in choice test by the oriental fruit fly *Bactrocera dorsalis*. Prabal Saikia and Rameswaran (2000) conducted free choice method to test the repellent action of plant derivatives against *Cnaphalocrosis medinalis* (Guen) and reported that neem oil 60 EC 1% when used 38.3% larvae moved towards neem treated and 61.6% towards control leaves after settlement observed

#### Use of Robin blue in HaNPV

Efficacy of entomopathogens can be maximized by conserving their stability in the environment (Ignoffo and Falcon, 1978). Major obstacle in the use of insect viruses in field situations is the rapid inactivation by ultraviolet radiation (Gudauskas and Canerday, 1968; Jaques, 1985, Ignoffo and Garcia, 1992).

Rabindra and Jayaraj (1988) reported that when HNPV was applied to *C.arietinum* plant at 1,00,000 polyhedral bodies / ml and exposed to field conditions, its persistence was increased with 1% Robin blue and Tinopal According to Rabindra *et al.* (1989) addition of whole milk (20%), whole egg homogenate (10%), Ranipal (0.5%), Robin blue (0.5%) and cotton seed oil (5%) to ULV sprays of HNPV in chickpea effectively controlled the noctuid *H.armigera* and reduced pod damage and significantly increased yields.

## Effect of HaNPV on Different Age Groups of H.armigera

Phenomenon of maturation immunity where the larvae can not be infected by a pathogen beyond a particular age has been reported in *Harmigera* (Whitlock, 1977). Ignoffo (1966) reported that as the age of the *Hizea* and *Hivrescens* larvae increases their susceptibility to the NPV decreases, the same was also reported by Allen and Ignoffo (1969) in case of *Hizea*. According to Boucias *et al.* (1980) as the age of velvet caterpillar. Anticarsia gemmatalis (Hubner) larvae increases the time taken for NPV infected larvae to die increases. Evans (1981) showed that 90 per cent of the variability of *Memestra brassicae* (Linnaeus) to NPV susceptibility could be accounted for the increase in body weight. Smits and Vlak (1988) reported that the time for NPV infected *Spodoptera exigue* (Hubner) larvae to die increases with age of the larvae.

Prasad and Ramakrishnan (1993) found that S.litura larvae after certain age could not be infected by nuclear polyhedrosis virus (NPV) Jayachandran and Chaudhari (1996) reported that in case of S.litura the mortality due to NPV infection increases due to increased dosage and decreases with host age. According to Ingalhalli et al. (1995) during NPV infection in the armyworm Mythimna separata (Walk.) the fat body, gut and integument indicated hypoglycemia, where as the haemolymph demonstrated the hypertrehalosemia and hyperglycemia. These changes were similar to the ones observed during starvation, but hypertrehalosemia was more evident than hyperglycemia during starvation. The growth rate, gross and net efficiency of food utilization for body matter were observed to be decreased during the course of NPV infection to H. armigera (Kencharaddi and Jayaramaiah, 1997). It was also reported that LC50 of HaNPV is 2.9 x 104, 5.33 x 104 and 2.7x105 PIBS / mI for first, third and fifth instars of H. armigera, and the LT50 is 4,8 days at 2.0 x 10<sup>5</sup> PIB / mI for 1<sup>st</sup> and 3rd instars, respectively. Chaudhary (1997) calculated LC50 value for 4 and 12 day old larvae of Spilosoma obliqua Wlk. as 2.6 x 104, 2.96 x 105 PIB / ml, respectively and suggested that virus application at early stages of the

larvae will be more effective for maximum foliar protection with early death of the larvae. Increasing virus dosage slightly increased speed of kill in respect of HzSNPV against *H. zea* and for AfMNPV against *S. frugiperda* (Farrar and Ridway, 1999).

Patil *et al.* (1989) studied the sublethal effects of the LC25 and LC50 NPV treatments on *M.seperata* parent, F, generations and the results revealed that in both the generations weight of the larval, pupal and adult stages and the adult longevity decreased, while developmental duration for the larval and pupal stages increased significantly. Further, pupation and adult emergence rates, growth index, fecundity, average egg production, per cent egg hatchability declined considerably. The above results were also demonstrated in spruce bud worm (Morris, 1977) and in the cotton boll worm *H. zea* (Luttrell *et al.*, 1982).

#### MATERIALS AND METHODS

Studies on the "Effect of different IPM components on *Helicoverpa armigera* Hubner and their impact on natural enemies in Chickpea " were conduted at the International Crops Research Institute for Semi-Arid Tropics (ICRISAT). Patancheru, during two chickpea seasons (post rainy season 1998-99 (November to February) and 1999-2000 (September to February) The materials used and methods employed in conducting these experiments are elucidated in this chapter.

#### 3 1 FIELD EXPERIMENTS

The influence of IPM components on *H.armigera* and their impact on natural enemies in chickpea was investigated by conducting field experiments in ICRISAT farm, Patancheru during post rainy season (*rabi*) 1998-99 and 1999-2000.

#### 3.1.1 Experimental Design

The research was conducted in black percision (BP) 7A field of ICRISAT farm with an area of 8000 Sq. m during *rabi* 1998-99, and BP14 field of ICRISAT farm with an area of 9000 Sq. m during *rabi* 1999-2000 The area was divided into 24 plots, each plot measuring 288 Sq. m (18 x16 m during *rabi* 1998-99 and 24 x 12 m during *rabi* 1999-2000), to conduct the experiment with six treatments in four replications each Randomized block design was used to conduct the trial (Plate.1).

#### 3.1.2 Sowing

A high yielding, desi, medium duration variety ICCC 37 (kranti) seed was obtained from ICRISAT. To reduce the incidence of seed borne diseases such as collar rot, and root rot the seeds were treated with Mancozeb @ 2 g / Kg of seed. The treated seeds were sown on 11th November during *rabi* 1998-1999 and 22nd October during *rabi* 



Plate 1 : Field view of the experimental plot



Plate 2 : Bird perch in the experimental plot.

1999-2000 with the spacing of 60 cm between rows and 15 cm with in a row

## 3 1 3 Efficacy of Different Treatments Against Gram pod Borer and Their Natural Enemies

A field experiment was conducted in a randomized block design with six treatments and four replications. The experiment was conducted in 288 Sq m area plots with ICCC 37 chickpea variety. The following treatments were used to study the effect of treatments on gram pod borer and their natural enemies.

- T<sub>1</sub> Neem (AZA 3%) 0.006% during *rabi* 1998-99 and (Nivaar 1500 ppm) @ 1750 ml / ha during *rabi* 1999-2000.
- T, Heliothis Nuclear Polyhedrosis Virus 250 LE / ha
- T, Fixing bird perches @ 1 perch / plot.
- T Endosulfan 35 EC 0.07%.
- $T_{1}$  Integrated Pest Management  $(T_{11}, T_{22}, T_{3} \text{ and } T_{4})$
- T<sub>c</sub> Control

#### 3.1.3.1 Neem

During *rabi* 1998-99 30,000 ppm neem product AZA3% was supplied by Dr. Baliga, Technical Consultant, Mumbai. This AZA 3% was obtained through ICRISAT and used in this experiment. 20 ml of AZA 3% in 10 liters of water was mixed to obtain 60 ppm or 0.006% used for spraying. During *rabi* 1999-2000 neem product Nivaar (1500 ppm) was obtained from ICRISAT sprayed at recommended rate i.e., 1750 ml / ha (50 ml / 10 l of water). This spray fluid requirement was standardized before spraying by using water at 10 liters per plot.

### 3132 Heliothis armigera nuclear polyhedrosis virus (HaNPV)

Heliothis armigera Nuclear Polyhedrosis virus was produced at ICRISAT-NPV laboratory and used for the studies. The HaNPV stock solution was prepared such that 1 ml of HaNPV solution equals to one larval equivalent (LE) containing 6 x 10° POBs. Since, virions were sensitive to ultra violet rays of sunlight, the spraying was done in evening hours. In order to protect the polyhedron particles from ultra violet rays robin blue was mixed in the spray solution @ 1 ml/l of spray fluid HaNPV was used @ 250 LE / ha.

#### 3133 Bird perches

Dried up tree branches were cut to create a natural tree like appearance in the field, to be used as a perch by the visiting bird (Plate.2). The vertical height of the perch was about 1.5 m from ground, a suitable height for insectivorous birds to rest and search for the larvae in the chickpea canopy. The perches were installed and maintained @ one perch per plot from 21 DAS and 32 DAS during 1998-99 and 1999-2000, respectively, till crop harvest.

#### 3.1.3.4 Endosulfan

Endosulfan 35 EC was obtained from ICRISAT and used in the experiment. To prepare 0.07% concentration, 2 ml of the stock solution was mixed in a liter of water.

The treatments were given five times during *rabi* 1998 – 99 and 1999 – 2000 at 15 days interval during cropping period. The sprays were - initiated after pest population was above ETL (2 small size larvae/ plant) The sprays were imposed on 21,37, 52, 67 and 84 DAS during *rabi* 1998 – 99 and 32, 47, 62, 78 and 94 DAS during *rabi* 1999 – 2000. The variation of dates of spraying during the two years of study was due to the variations in incidence, stage of the crop and condition of the field.

T<sub>2</sub> (HaNPV) treatment at all the above mentioned days and in IPM  $(T_{s})$  at 37 and 84 DAS during *rabi* 1998-99 and 47 and 94 DAS during *rabi* 1999-2000 received HaNPV spray @ 250 LE / ha after 4 p m mixed with UV protectant

In all the replications of  $T_3$ , bird perches were installed @ one perch per plot on the day when first spray of other treatments was given and retained in the plot till last observation was taken.

In  $T_s$  (Integrated Pest Management) plot, bird perches were installed @ one perch per plot on the day of first spraying and kept untill the last observation was taken. At the same day  $T_s$  received neem spray. Second and third were HaNPV 250 LE / ha and endosulfan 0.07%, respectively. Once again neem and HaNPV were given as fourth and fifth spray, respectively, to manage *H.armigera* throughout the crop period, in  $T_s$ 

#### 3135 Methods of observation

From each plot twenty plants were randomly selected for recording observations. In each plant the number of eggs, small size (first & second instars), medium size (third & fourth instars) and large size (fifth & sixth instars) larvae were counted at weekly interval. The observations were taken 15 DAS onwards during *rabi* 1998-99 and 24 DAS during *rabi* 1999-2000 with weekly interval till crop maturity. The mean number of larvae/eggs per plant at different crop stages in different treatments were worked out.

The data were subjected to square root transformation for analysis in randomized block design.

#### 314 Effect of Different Treatments on Natural Enemies

## 3 1 4 1 Monitoring the activity of soil inhabiting natural enemies in different treatments.

For monitoring soil inhabiting natural enemies pitfall traps were used (Plate 3). One litre plastic containers were used as pitfall traps. These containers were placed in the soil by burrying to the ground level at the rate of three traps per plot. These traps were installed at 21 DAS during *rabi* 98 – 99 and 30 DAS during *rabi* 1999 – 2000 at random in the plot. These jars acted as traps to moniter soil dwelling natural enemies

One ml of formaldehyde and 1 ml of soap water were mixed with one litre of water and poured into the trap up to 3/4 the volume, so that natural enemies falling into the trap will be killed immediately after falling and preserved well in the trap without spoilage upto observation.

#### Methods of observations

Observations were taken once in 10 days till crop maturity. Individual traps were removed from the soil and then formaldehyde, soap water mixtures along with collected insects were poured into a filter to separate the insects from the collected fluid. From the collection, individual insects were separated using camel hair brush/forceps and were identified. Thus, observations were made in all the treatments across the trial. The traps were cleaned with water and replaced once again in pits at ground level with formaldehyde & soap water solution.

The total number of natural enemies in all traps of a treatment was worked out. The mean number of natural enemies present during different crop stages in different treatments were calculated. The data collected were analyzed in randomised block design after transforming into square root



Entrall trapitor collecting soil inhabiting natural energies



We for collecting aerial catural energies

values to get the effect of different treatments on the activity of soil inhabiting natural enemies.

#### 3142 Effect of different treatments on the aerial natural enemies

De Vac (Plate 4) was used to assess the activity of various predators and parasitoids in different treatments, which were inhabiting on the crop canopy (aerial natural enemies). Due to the vacuum created inside the trap, the insects found in the crop canopy were captured inside the trap. This trap was operated twice during *rabi* 1998 – 99 i.e. at 22 DAS and 54 DAS. During *rabi* 1999 – 2000 this trap was not operated due to machine failure. But it was substituted with hand operated trap, however due to stickiness of foliage because of presence of malic and oxalic acids most of the aerial natural enemies were stuck to the foliage.

At the time of De Vac operation, operator walked twice on any of the two rows in a plot for one minute by carrying De Vac on his back and holding mouth of the trap near the crop canopy. The collected material was transferred into a polythene cover and labelled. The counting and identification was carried out in the laboratory.

Total number of natural enemies in different treatments were calculated and subjected to randomized block design analysis to assess the effect of different treatments on the natural enemies in the crop canopy.

# 3 1 4 3 Efficacy of different treatments on egg, larval and pupal parasitoids

To evaluate the percentage egg parasitism, 100 eggs along with leaves @ one egg per leaf per vial for different treatments were collected and observed daily till larvae or parasitoid hatched out For observing larval or larval-pupal parasitism 100 larvae per treatment were collected, and released in individual glass vials and observed daily for parasitism. Larvae were fed with soaked chickpea seeds and the feed was changed at alternate days. Observations were made upto adult emergence

Two such collections were made on 25 DAS and 57 DAS during rabil 1998-99 and 36 and 73 DAS during rabi 1999-2000

Total number of parasitised eggs and larvae and pupae were counted separately and the percentage parasitisation was worked out. The data were subjected to randomized block design analysis after transforming the percentage values into arc sin values.

#### 315 Pod Damage

Net plot area (14 x 6m) was marked and plant population in net plot area was counted before harvesting.

From net plot area 20 plants were removed randomly and all the pods were removed and collected in a cover and labelled. In the laboratory, number of healthy and pod borer damaged pods were counted and percentage pod damage was worked out for all the plots using the formula given below.

#### 316 Yield

Net plot area was separately harvested and threshed 3 days after harvesting. Threshed grains were cleaned and weighed. The pods collected from 20 plants were removed from net plot for working out per cent

pod damage and also threshed, cleaned and weighed and was added to the net plot yield

The data were subjected to RBD analysis to know the effect of different treatments on yield.

#### 3 1 7 Residue Analysis

Pesticide residues present in husk and seed of chickpea in endosulfan treatment and IPM treatment plots were analyzed for residues of endosulfan

#### 3171 Method of sample collection

To determine the residues of endosulfan in chickpea plots treated with endosulfan and endosulfan received IPM treatment, ten chickpea plants were collected at random in all the four replications at the time of harvest, air dried and preserved in the refrigerator.

#### 3172 Extraction and clean up

The chopped and blended chickpea husk, seed, 200g and100g, respectively was taken from the composite sample and extracted with 300ml mixture of n-hexane:Isopropanol (2.1). The filtered extract was washed with distilled water and the aqueous phase was discarded. The hexane layer was collected through anhydrous sodium sulphate. A drop of keeper was added and extract was concentrated.

The concentrated extract was dissolved in 45-50 ml of hexane:acetone (9:1) and little quantity of Darco G 60 (activated charcoal) was added with occasional shakings. This was filtered through filter paper and residues were washed with 3X15 ml of hexane:acetone (9.1) mixture making it ready for GC analysis.

#### **3**173 Determination

The residue estimation was done using GCECD with the following parameters GC ÷ Packard 437A Detector Electron Capture Detector Column 4%SE 30+6% OV 210 Column (Oven) 190 Temperature(°C) 250 Detector temperature(°C) Injector temperature(°C) 270

Carrier gas flow (ml/min)		N <sub>2</sub> 60
Retention time (min)		
Endosulfan I	•	7.3
Endosulfan II	:	10.5
Endosulfan sulphate	:	2.8

#### 3174 Recovery

Chickpea plants were collected from control plots and after chopping and blending, were transferred to the reagent bottles at the rate of 100 g A known amount of standard solution (equals to 1ppm) was added. After shaking the contents, the samples were subjected to clean up for the determination of residue. The recovery obtained was 99 per cent for endosulfan I&II and 97.5 per cent for endosulfan sulphate.

#### **32 LABORATORY STUDIES**

#### 3 2 1 Effect of Neem on H. armigera Oviposition

Sixty. 30 days old potted chickpea plants were used for this study Two pots in each treatment and fifteen replications were maintained in a cage Neem @ 0.006% was sprayed with hand operated sprayer as one treatment and water spray as another treatment. Fifteen pairs of *H armigera* adults less than 12 hrs old were released into the cage immediately after spraying 10% honey solution on cotton swabs was provided in this cage as adult food. The number of eggs per pot were counted and recorded in treated and untreated pots daily until the death of adults. Total number of eggs per pot was calculated.

The data were tested by using two sample t-test to know the significance of the treatments.

#### 322 Larval Preference for Neem

#### Choice test

In large petriplates (15 cm diameter) neem (azadirachtin) sprayed chickpea leaves on one side and water sprayed chickpea leaves on opposite side were kept. Ten first instar larvae were released in the middle Their movement towards neem treated or control chickpea leaves were observed at hourly interval until they have settled for continuous feeding Ten replications were maintained and the same process was repeated for third instar larvae also. But for fifth instar larvae, due to cannabalism, in each petri plate one larva was released in middle and a total of ten petri plates constituted one replication, and for 10 replications 100 petri plates were maintained and their preference was observed.

#### No choice test

In large petri plates neem (azadirachtin) sprayed chickpea leaves were kept and ten first instar larvae were released on them Larval searching behaviour was observed at hourly interval until they have settled for continuous feeding. Ten replications were maintained. The same process was repeated for third instar larvae. But for fifth instar in each petri plate one larva was released, a total of ten petri plates constituted one replication. Hundred petri plates were maintained for ten replications Larval preference was recorded and expressed as per cent settled on neem and tried for another food source.

Two sample t-test was used to know the significance of preference between neem and water sprayed chickpea leaves for different instars

#### 3.2.3 Effect of Neem on the Different Age Group of H. armigera

Neem 0.006% was sprayed on field collected chickpea leaves and pods Then air dried for ½ hour. Then the leaves and pods were fed to three age groups viz. first & second, third & fourth, fifth & sixth instars. For each age group ten replications with 12 larvae in each replication were maintained. For each age group control was also maintained separately Larval mortality was recorded from 24 hrs after treatment upto pupation Larval duration, pupal period and pupal weight were recorded. Adult mortality was also observed.

Pupae were collected from larvae that received neem treatment. Adults emerged from these pupae were tested for their fecundity. The fecundity was observed by releasing adults of *H.armigera* female:male in 1:2 ratio in oviposition chambers which were provided with egg laying tissue papers and 10% honey in cotton swabs. The number of eggs was recorded daily from third day after release upto their death Oviposition period was also recorded. Total number of eggs per female was calculated.

The egg hatching was observed by keeping a small piece of egg laying tissue paper along with the eggs in closed plastic boxes. The number of eggs hatched were recorded and hatching per cent was calculated

The data were tested by using two sample t-test.

#### 3 2 4 Persistence of HaNPV

Persistence of HaNPV under field conditions is one of the main drawbacks in the use as a bio-insecticide. Of the several locally used UV protectants Robin blue is the most popular one. Hence its efficiency was tested to increase HaNPV persistence under field conditions. The treatments were

- T, HaNPV @ 6 x 10° POB /I alone.
- T, HaNPV @ 6 x 10° POB / I +1% Robin blue
- T, Control (water spray)

The treatments were applied on chickpea crop in three different 100 m apart patches. From each treatment leaves and pods were collected immediately after treatment, 1, 2, 3, 4, 5, 6 and 7th day after treatment. Eighty four laboratory reared third instar larvae / treatment (12 larvae / replication) were used. Seven replications were maintained. Larvae were starved for one day before treatment. Mortality of larvae was recorded every day until pupation and mortality per cent was calculated. This experiment was repeated for 3 times.

The data were subjected to arc sin transformation and analyzed by Randomized Block Design.

325 Effect of HaNPV on three different age groups of H. armigera

Field collected chickpea leaves and pods were sprayed with HaNPV @ 6 x 10<sup>9</sup> POB / I.. air dried for ½ hr., and then HaNPV treated leaves and pods were fed to three age groups. After 24 hrs the feed was changed with fresh untreated soaked chickpea seeds. Mortality of the larvae was recorded from 24 hrs after treatment up to pupation. Pupal abnormality and adult emergence were also recorded.

## 3.2.6 Effect of HaNPV Treatment During Larval Stage on Fecundity and Egg Hatching of *H.armigera*.

Pupae were collected from the population that received HaNPV treatment during larval stage. For this experiment four treatments were used (1) female from HaNPV treated population and male from normal population, (2) male from HaNPV treated population and female from normal population. (3) both male and female from HaNPV treated population and lastly (4) male and female from normal population. The moths were released in the egg laying chambers made of plastic which were provided with egg laying tissue papers and 10% honey solution on cotton swabs. The egg laying was recorded from third day after release until the death of adults. Five replications were maintained, and the experiment was repeated three times. The data were subjected to square root transformation for analysis.

From the same treatments a small piece of egg laying tissue paper containing more or less hundred eggs was kept for egg hatching. The number of eggs hatched was recorded and per cent hatch was calculated. The per cent of eggs hatched were subjected to arc sin transformation for analysis.

The observations were analyzed by using Completely Randomized Block Design.

#### RESULTS

#### **4 1 FIELD STUDIES**

#### 411 Population Fluctuations of H.armigera during rabi 1998-99

The moth activity of *H.armigera* was seen throughout the crop period with peaks at 43 DAS (100.7 moths/trap), 71DAS (55 moths/trap) and 9? DAS (58.3 moths/trap).

Observations on the number of eggs during 1998-99 season showed an average of 1.22 eggs per plant at vegetative stage and 2.00 eggs/ plant at flowering stage with a peak of 3.36 eggs/plant at 50 DAS (Appendix I) There was little difference in mean number of eggs/plant during podding and preharvest stages, indicating a similar moth activity throughout that period.

Data on small size larval counts indicated 1.5, 1.9, 1.5 and 1.3 larvae per plant during vegetative, flowering, pod formation and preharvest stages of the crop, respectively. This clearly showed uniform occurrence of small larvae throughout the crop period which was the result of continuous adult activity

Maximum number of medium size larvae were observed at pod formation stage (1.45 larvae/plant at 71DAS) (Appendix.III) The medium size larval population in the other stages of the crop was around 0.8 larva/plant

Large size larval peak was noticed at preharvest stage with 0.39 larva per plant, and the population during the other stages was around 0.3 larva per plant. Fig. 1 : Population fluctuations of *H.armigera* in chickpea during *rabi* 1998-99



The pooled larval data showed the peak activity with 3.0 larvae per plant during the flowering stage of the crop, later the population declined slightly and reached 2.3 larvae/plant by preharvest stage of the crop

### 4 1 2 Efficacy of the Treatments on Ovipositional Behaviour of *H.armigera* during rabi 1998-99

In order to assess the efficacy of different treatments on the ovipositional behaviour of *H. armigera*, studies were conducted in chickpea crop during *rabi* 1998 – 99. The results are presented in Table 1. The number of eggs per plant were recorded at weekly interval in different treatments on 20 random plants and the mean number of eggs per plant during different stages of crop are presented. The crop received a total of five sprays during the entire cropping period, two sprays (21 DAS, 37 DAS) during vegetative stage, one spray during flowering stage (52 DAS) and two sprays during pod formation stage (67 DAS, 84 DAS).

#### Vegetative stage

The plots treated with neem and IPM treatment which received neem as first and HaNPV as second spray were found highly effective in reducing egg laying by *H.armigera* with lower number of eggs per plant (0.65 and 0.69 egg/plant, respectively) during vegetative stage. The remaining treatments, HaNPV (0.82), endosulfan (0.84) and bird perches (0.94) were on par with significantly less number of eggs per plant than control (1.22).

#### Flowering stage

During flowering stage also neem (1.33 eggs/plant), HaNPV (1.37 eggs/plant) and IPM (1.40 eggs / plant) which received endosulfan as third spray were on par and found superior in keeping the egg number at low level. But and were as effective as neem with and, respectively. Control (1.99) recorded more number of eggs. Bird

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage	(Mea	an number of	eggs per plant)	
Neem 0.006%	0.65	1.33	0.58	0
(AZA 3%)	(0.8060)*	(1.1526)ª	(0.7635)*	(0.2236)"
HaNPV 250LE/ha	0.82	1.37	0.73	0.07
	(0.9063)ªb	(1.1688)ª	(0.8561) <sup>60</sup>	(0 3446) <sup>bc</sup>
Bird perches one/plo	0.94	1.69	0.73	0.06
	(0.9681) <sup>6</sup>	(1.3005) <sup>₀</sup>	(0.8537) <sup>bc</sup>	(0.3354) <sup>te.</sup>
Endosulfan 0.07%	0.84	1.74	0.68	0.02
	(0.9183)ªº	(1.3195)º	(0.8239) <sup>ab</sup>	(0.2622) <sup>ab</sup>
IPM	0.69	1.40	0,58	0.01
	(0.8326)"	(1.1812)"⁵	(0.7579)"	(0 2500)**
Control	1.22	1.99	0.87	0.09
	(1.1038)⁰	(1.4090)⁰	(0.9328)°	(0.3791) <sup>,,</sup>
S.Ed.	0.064	0.057	0.043	0.052
CD	0.135	0.120	0.090	0 111

 Table 1: Effect of the treatments on oviposition behaviour of H. armigera during rabi 1998-99.

(Figures in parenthesis are square root transformed values)

\*Values followed by same letters in each column are statistically not significant

perches (1.69) and endosulfan (1.74) showed no effect on *H armigera* ovipositional behavior and were on par with control

#### **Pod** formation stage

It is one of the most critical stages of the crop, where the infestation had direct effect on yield. IPM (0.58) which received neem as fourth and HaNPV as fifth spray and neem (0.58) showed good effect in minimizing the oviposition of *H.armigera*. Endosulfan (0.68) treatment was also found as effective as IPM and neem treatments. Control plot had the highest number of eggs throughout the pod formation stage with 0.87 egg per plant HaNPV (0.73) and bird perch (0.73) treatments showed little effect on repelling the ovipositing adults, being on par with control.

#### Preharvest stage

During this period neem treatment (0) provided maximum repellency for *H.armigera* oviposition. IPM (0.01) and endosulfan (0.02) treatments also recorded less number of eggs and were on par with neem. Bird perches (0.06) and HaNPV (0.07) were on par with control treatment (0.09)

### 4.1.3 Efficacy of the Treatments on Small Size (first & second instars) Larvae of *H.armigera* during *rabi* 1998-99

Studies were conducted to assess the efficacy of different IPM components in managing the small size larval population. Observations were recorded at weekly interval on 20 random plants per plot and the mean number of larvae per plant at different crop stages are described below (Table 2).

#### Vegetative stage

During this stage of the crop two sprays were given with the **res**pective treatments, once at 21 DAS and another at 37 DAS. The plots **sprayed** with endosulfan recorded the lowest number of larvae per plant (0.91) but it was on par with HaNPV (1.09). IPM which received neem as first and HaNPV as second spray and neem and bird perches (1.12, 1.18 and 1.25 larvae/plant respectively) were statistically on par and significantly **superior** to control (1.47) in bringing down the larval population of *H.armigera* during vegetative stage of the crop.

#### Flowering stage

The sprays in the respective treatments were given at 52DAS coincided with flowering stage of the crop. The plots with IPM treatment which received endosulfan as third spray recorded the lowest number of larvae per plant (1.29). However the plots treated with endosulfan (1.34) and HaNPV (1.38) were also on par with IPM. Neem (1.43) recorded next in the order of efficacy. The control plot recorded the highest number of larvae (1.88). Bird perches showed no effect on managing small larvae and it was on par with control.

#### Pod formation stage

During podding stage the crop received two sprayings at 67 DAS and 84 DAS. The IPM treatment which received neem as fourth spary and HaNPV as fifth spray registered the lowest number of larvae (1.16). The plots sprayed with HaNPV (1.23), neem (1.28) and endosulfan (1.28) were<sup>-</sup> also on par with IPM and recorded less number of larvae compared to control. Control recorded the highest number of larvae (1.49). Bird perches plot showed no effect in managing small size larvae by recording 1.33 larvae/plant and was on par with control.

Treatment/ crop stage	Vegetative Flowering Pod formation (Mean number of larvae per plant)			Preharvest
Neem 0.006%	1.18	1.43	1.28	0.97
(AZA 3%)	(1.0852) <sup>ь</sup>	(1.1971)⁵	(1.1308)⁰⁵	(0.9840) <sup>ь</sup>
HaNPV250LE/ha	1.09	1.38	1.23	0.95
	(1.0422) <sup>սե</sup>	(1.1741) <sup>ııb</sup>	(1.1084) <sup>sb</sup>	(0.9745) <sup>ь</sup>
Bird perches one/plot	1.25	1.84	1.33	1.00
	(1.1165)⁵	(1.3554)°	(1.1528) <sup>⊳c</sup>	(1.0003) <sup>₅с</sup>
Endosulfan 0.07%	0.91	1.34	1.28	0.94
	(0.9517)ª	(1.1563)ª	(1.1326)ª⁵	(0.9680)⁵
IPM	1.12	1.29	1.16	0.27
	(1.0560) <sup>⊾</sup>	(1.1361)ª	(1.0778)³	(0.5180)ª
Control	1.47	1.88	1.49	1.25
	(1.2118)⁰	(1.3692)⁰	(1.2212)⁰	(1.1179)⁰
S.Ed.	0.045	0.019	0.033	0.056
CD .	0.095	0.040	0.069	0.118

 Table 2 : Efficacy of the treatments in managing small size larvae of H.armigera

 during rabi 1998-99

(Figures in parenthesis are square root transformed values)

\*Values followed by same letters in each column are statistically not significant

#### Preharvest stage

The crop received fifth spray at 84 DAS during pod formation but its effect was measured at preharvest stage. Based on the results, IPM plot was the most efficient by recording the lowest population with 0.27 larva per plant and significantly superior to the rest of the treatments. Endosulfan (0.94) stood next in the order of efficacy but was on par with HaNPV (0.95), neem (0.97) and bird perches (1.00). Control registered the highest number of larvae (1.25) but was at par with bird perches.

## 4.1.4 Efficacy of the Treatments on Medium size (third & fourth instars) Larvae of *H.armigera* during *rabi* 1998-99

To evaluate the effect of different IPM components in managing the medium (III & IV instars) size larval population, an experiment was conducted with a total of five sprays during the crop period, two sprays during vegetative, one spray during flowering, and two sprays during pod formation stage. The number of larvae per plant in different treatments were recorded at weekly interval and the mean number of larvae per plant at each crop stage are presented in Table 3.

#### Vegetative stage

Neem (0.48 larva/plant) and HaNPV (0.49 larva/plant) on par and were found to be the most effective and significantly superior to the rest of the treatments by recording the lowest number of medium size larvae. Endosulfan (0.56), IPM which received neem as first spray and HaNPV as second spray (0.56) and bird perches (0.62) were on par and significantly superior to control (0.79) in bringing down *H.armigera* medium size larval population during vegetative stage of the crop.

#### Flowering stage

During flowering stage IPM was the most effective treatment which received endosulfan as third spray with 0.58 larva per plant. HaNPV (0.62), neem (0.62) and endosulfan (0.64) were also as effective as IPM treatment in managing the medium size larvae at flowering stage. Bird perch plot also gave a significant reduction in the number of larvae per plant (0.72) compared to control (0.89).

#### Pod formation stage

IPM that received neem as fourth and HaNPV as fifth spray was significantly superior to the rest of the treatments in managing medium size larvae with 0.68 larva per plant. Birds were active in plots with bird perches during this stage and recorded 0.81 larva per plant and stood next in second position. But HaNPV (0.83) and endosulfan (0.89) were on par with bird perches. Neem was the least effective with 0.90 larva per plant among the treatments while control plot recorded significantly high larval population (1.00 larva per plant).

#### Preharvest stage

At preharvest stage also IPM showed its significant effect in managing the medium size larvae compared to other treatments by recording the lowest number of larvae (0.49). There was no significant difference in the number of larvae in the plots treated with HaNPV, bird perches and endosulfan (0.58, 0.61, 0.61, respectively). Once again neem was the least effective with 0.68 larva per plant among the treatments. Control plot experienced significantly higher larval population (0.84 larva per plant).

Treatment/ Crop stage	Vegetative Flowering Pod formation (Mean number of larvae per plant)			Preharvest
Neem 0.006%	0.48	0.62	0.90	0.68
(AZA 3%)	(0.6890)ª	(0.7878)ª	(0.9483)º	(0.8215)⁰
HaNPV250LE/ha	0.49	0.62	0.83	0.58
	(0.7026)°	(0.7850)ª	(0.9126) <sup>⊳</sup>	(0.7622)⁵
Bird perches one/plot	0.62	0.72	0.81	0.61
	(0.7865)⁵	(0.8464)⁵	(0.9012) <sup></sup>	(0.7785) <sup>bc</sup>
Endosulfan 0.07%	0.56	0.64	0.89	0.61
	(0.7498)⁵	(0.7982) <sup>ab</sup>	(0.9417) <sup>bc</sup>	(0.7824) <sup>be</sup>
IPM	0.56	0.58	0.68	0 .49
	(0.7497)⁵	(0.7606)*	(0.8214)ª	(0.7023)*
Control	0.79	0.89	1.00	0.84
	(0.8890)°	(0.9419)⁰	(1.0003)₫	(0.9184)⁴
S.Ed.	0.018	0.024	0.022	0.025
CD	0.039	0.051	0.046	0.053

## Table 3 : Efficacy of the treatments in managing medium size larvae of H.armigera during rabi 1998-99

(Figures in parenthesis are square root transformed values)

\*Values followed by same letters in each column are statistically not significant

## 4.1.5 Effect of the Treatments on Large Size (fifth & sixth instars) Larvae of *H.armigera* during *rabi* 1998-99

The efficacy of different IPM components individually and in combination in managing large size larvae of *H. armigera* was tested and the results are presented in Table 4. Five sprays were given during the crop period, two at vegetative, one at flowering and the remaining two at pod formation stage.

#### Vegetative stage

Endosulfan stood first in controlling large size larvae by recording the lowest number (0.17) per plant during vegetative stage. IPM that received neem as first and HaNPV as second spray in addition to bird perches was the next effective treatment with 0.18 larva/plant and it was found on par with HaNPV (0.19), neem (0.19) and bird perches (0.21). All the treatments were significantly superior to control which recorded the highest larval population (0.32).

#### Flowering stage

IPM that received endosulfan as third spray was more effective with the lowest mean number of larvae per plant (0.081) during flowering stage, but endosulfan (0.10) was as effective as IPM treatment. Neem (0.12), HaNPV (0.13) and bird perches (0.14) were on par in managing the large size larvae. Control recorded significantly the highest larval number (0.28).

#### Pod formation stage

Larvae per plant during podding stage was the least in IPM (0.15) and HaNPV (0.15). Endosulfan (0.17) was on par with IPM and HaNPV managing large sized larvae. Bird perches (0.19) and neem (0.21) were on par in managing large size larvae but significantly superior over control (0.36).
Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage		(Mean numbe	er of larvae per pla	ant)
Neem 0.006%	0.19	0.12	0.21	0.05
(AZA 3%)	(0.4400) <sup>ab</sup>	(0.3413) <sup>₅</sup>	(0.4562) <sup>ь</sup>	(0.2235)ª
HaNPV250LE/ha	0.19	0.13	0.15	0.06
	(0.4398) <sup>sb</sup> ·	(0.3591)°	(0.3816)°	(0.2369) <b>°</b>
Bird perches one/plo	t 0.21	0.14	0.19	0.08
	(0.4608) <sup>ь</sup>	(0.3706)⁰	(0.4328) <sup>ь</sup>	(0.2849)*
Endosulfan 0.07%	0.17	0.10	0.17	0.06
	(0.4105)ª	(0.3093)ªb	(0.4080)ª <sup>b</sup>	(0.2498)ª
IPM	0.18	0.08	0.15	0.06
	(0.4291) <sup>ab</sup>	(0.2900)ª	(0.3816)ª	(0.2497)ª
Control	0.32	0.28	0.36	0.39
	(0.5644)°	(0.5257)⁴	(0.5985)⁰	(0.6223)⁵
S.Ed.	0.015	0.017	0.023	0.029
CD	0.032	0.037	0.049	0.062

## Table 4 : Efficacy of the treatments in managing large size larvae of *H.armigera* during *rabi* 1998-99

(Figures in parenthesis are square root transformed values)

#### Preharvest stage

All the treatments neem (0.05), HaNPV (0.06), IPM (0.06), endosulfan (0.06) and bird perches (0.08) except control were on par and registered uniformly less population but significantly superior to control (0.39) in managing large size larvae at pre harvest stage.

### 4.1.6 Efficacy of the Treatments on the Total Larval Load of *H.armigera* During *rabi* 1998-99

In order to assess the efficacy of different IPM components in managing the *H.armigera* larval population, an experiment was conducted and the results are presented in Table .5. The number of larvae in all the experimental plots were recorded at weekly interval and the data were compiled to get the mean number of larvae at different crop stages.

#### Vegetative stage

The interruptions were made in the respective treatments at 21 DAS and 37 DAS which coincided with vegetative stage of the crop. The results revealed a significant reduction in larval number in the plots treated with endosulfan with the lowest number of (1.58) larvae/plant. The plots treated with HaNPV with 1.78 larvae/plant stood next in the order of efficacy. Neem and IPM which received neem as first spray and HaNPV as second spray were on par (1.85, 2.00 larvae/plant, respectively). The plot with bird perches was the least effective among the treatments with 2.10 larvae/plant but significantly superior over control (2.52).

#### Flowering stage

During flowering stage the crop received one spray with the respective treatments at 52 DAS. IPM which received endosulfan as third spray was the most effective treatment with the lowest number of larvae per plant

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Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage		(Mean numbe	of larvae per plar	nt)
Neem 0.006%	1.85	2.17	2.39	1.70
(AZA 3%)	(1.3588) <sup>₀</sup>	(1.4718)ª	(1.5449)°	(1.3036)⁴
HaNPV250LE/ha	1.78	2.13	2.22	1.59
	(1.3320)⁵	(1.4575)"	(1.4900) <sup></sup>	(1.2521) <sup>»</sup>
Bird perches one/plo	t 2.10	2.68	2.34	1.68
	(1.4501) <sup>d</sup>	(1.6378)⁵	(1.5300) <sup>⊳</sup>	(1.2965) <sup>cd</sup>
Endosulfan 0.07%	1.58	1.68	2.34	1.60
	(1.2560)ª	(1.4416)ª	(1.5285) <sup>₀</sup>	(1.2646) <sup>₅c</sup>
IPM	2.00	1.95	2.00	1.31
	(1.4140) <sup>od</sup>	(1.3965)ª	(1.4140)ª	(1.1426)ª
Control	2.52	3.00	2.89	2.28
	(1.5879)•	(1.7318)⁵	(1.7006) <sup>d</sup>	(1.5081)"
S.Ed.	0.027	0.046	0.026	0.018
CD	0.056	0.096	0.054	0.038

Table 5:	Efficacy of the treatments in managing total larval load of H.armigera
	during <i>rabi</i> 1998-99

(Figures in parenthesis are square root transformed values) \*Values followed by same letters in each column are statistically not significant

(1.95) but was at a par with endosulfan (2.08), HaNPV (2.13) and neem (2.17). Bird perches was less effective with 2.68 larvae per plant and was on par with control (3.0).

#### Pod formation stage

The sprays were given at 67 DAS and 84 DAS during pod formation stage of the crop. IPM treatment which received neem as fourth and HaNPV as fifth spray recorded significantly the lowest number of larvae per plant (2.00). HaNPV with 2.22 larvae per plant stood next in the order of efficacy. The plots treated with endosulfan, bird perches and neem (2.34, 2.34, 2.39 larvae/plant respectively) were on par but significantly superior compared to control which recorded the highest number of larvae per plant (2.89).

#### Preharvest stage

The results revealed a significant reduction in the number of larvae in the plots which received IPM treatment by recording the lowest number (1.31) of larvae per plant. The plots treated with HaNPV and endosulfan were on par and stood next with 1.59, 1.60 larvae per plant respectively. Bird perches (1.68) and neem (1.70) were on par and less effective among the treatments but significantly superior to control (2.28).

## 4.1.7 Population Fluctuations of *H.armigera* in Chickpea During *rabi* 1999-2000

Moth activity of *H.armigera* was observed throughout the crop period. Three peaks were observed, with the first peak during the initiation of flowering i.e 47 DAS (14.33 moths/trap), 2<sup>nd</sup> peak at 90 DAS (30.6 moths/ trap) and 3<sup>rd</sup> peak at 97 DAS (22.3 moths/trap) (Plate.5 & Figure.2).



Plate 5 Pheromone trap for recording adult activity of *H. armigera*.



Plate 6 Eggs of *H. armigera* on chickpea leaf.

During *rabi* 1999-2000 observations on egg population showed an average of 0.29 egg per plant at vegetative stage and there was an increase in the number at flowering stage (0.34 egg/plant) with a peak egg number of 0.48 (Appendix VI) at 54DAS. Later the number reached 0.07 egg per plant by preharvest stage of the crop (Plate.6).

The data on small size larval counts indicated 0.93, 1.33, 1.03 and 0.90 larvae per plant during vegetative, flowering, pod formation and preharvest stages of the crop, respectively with a peak at 54DAS (Appendix.VII).

The medium size larval peak was noticed at 83DAS (Appendix.VIII) which coincided with pod formation stage. The highest number of medium size larvae was observed during pod formation stage (1.18 larvae/plant). The medium size larval population in the other stages of the crop was in the range of 0.63 to 0.81 larva per plant.

The large size larval peak (0.58 larva per plant) was noticed at 90DAS (Appendix IX) which coincided with pod formation stage. The population was 0.10, 0.22, 0.40 and 0.36 larva per plant during vegetative, flowering, pod formation and preharvest stages, respectively.

The data on pooled larval population showed that moderate population (1.65 larvae per plant) was recorded at vegetative stage of the crop and there was gradual increase in the number by flowering stage (2.26 larvae/plant). The population reached peak (2.6 larvae per plant) at pod formation stage and there after the larval number declined slightly to 1.84 larvae per plant by preharvest stage of the crop.

6**2** 



Fig. 2 : Population fluctuations of H.armigera during rabi 1999-2000

## 4.1.8 Efficacy of the Treatments on Oviposition Behaviour of *H.armigera* During rabi 1999-2000

In order to assess the efficacy of IPM components on oviposition behaviour of *H. armigera* an experiment was conducted during *rabi* 1999-2000 and the results are presented in Table. 6. The number of eggs were observed at weekly interval and the mean number of eggs per plant at different crop stages were calculated.

#### Vegetative stage

The crop received one spray in the respective treatments at 32 DAS during vegetative stage. The plots treated with neem recorded significantly the lowest number of eggs per plant (0.04). The next best was IPM treatment (0.08) which also received neem as first spray. There was no significant difference in the number of eggs recorded in the treatment with HaNPV and endosulfan with 0.16 and 0.20 egg per plant, respectively. Plots with bird perche (0.25) showed no effect and was on par with control (0.29).

#### Flowering stage

During flowering stage the interruptions were made in the respective treatments at 47 DAS and at 62 DAS. The results revealed that the highest reduction in egg number was in the plots treated with neem (0.11) which was also on par with endosulfan (0.19). There was no significant difference in the number of eggs recorded between the bird perches, IPM treatment receiving HaNPV as second spray and endosulfan as third spray and HaNPV spray alone (0.22, 0.23, 0.28, respectively). These were also found to be on par with control (0.34).

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage	(N	lean number o	of eggs per plant)	
Neem 1750 ml/ha	0.04	0.11	0.03	0
(Nivaar 1500 ppm)	(0.2039)ª	(0.3352)*	(0.1741)"	(0.2236)"
HaNPV250LE/ha	0.16	0.28	0.09	0.03
	(0.4029)°	(0.5281) <sup>∞</sup>	(0.3059) <sup>bc</sup>	(0.2760) <sup>ab</sup>
Bird perches one/pl	ot 0.25	0.22	0.10	0.07
	(0.4956) <sup>d</sup> ⁰	(0.4652) <sup>bc</sup>	(0.3102)″	(0.3445) <sup>ad</sup>
Endosulfan 0.07%	0.20	0.19	0.08	0.03
	(0.4469) <sup>cd</sup>	(0.4375) <sup>ab</sup>	(0.2789) <sup>bc</sup>	(0.2848) <sup>bc</sup>
IPM	0.08	0.23	0.06	0
	(0.2955) <sup>ь</sup>	(0.4784) <sup>bc</sup>	(0.2404) <sup>ab</sup>	(0.2236)ª
Control	0.29	0.34	0.23	0.07
	(0.5359)*	(0.5806)°	(0.4754)⁴	(0.3487)⁴
S.Ed.	0.042	0.055	0.032	0.029
CD	0.088	· 0.116	0.068	0.061

## Table 6 : Effect of the treatments on oviposition behaviour of *H.amigera* during *rabi* 1999-2000

(Figures in parenthesis are square root transformed values)

#### Pod formation stage

During pod formation stage the treatments were applied at 78 DAS. The results revealed that the highest reduction in number of eggs was in plots treated with neem (0.03 egg/plant) but it was on par with IPM which received neem as fourth spray with 0.06 egg per plant. There was no significant difference in the number of eggs recorded in endosulfan, HaNPV and bird perches treatments (0.08, 0.09, 0.10, respectively), being on par but were found to be significantly superior compared to control (0.23) in recording less number of eggs per plant.

#### Preharvest stage

During this stage the crop was sprayed with respective treatments at 94DAS. No egg was recorded in the plots treated with neem and IPM treatment which received HaNPV as 5<sup>th</sup> spray. There was no significant difference between the number of eggs recorded in the HaNPV and endosulfan (0.03 both) which came next in the order. Bird perches (0.07) showed no effect and was found to be on par with control (0.07).

### 4.1.9 Efficacy of the Treatments on Small Size (first & second instars) Larvae of *H.armigera* During *rabi* 1999-2000

To find the effect of various IPM components individually and in combination for managing small size larvae of *H.armigera*, an experiment was conducted and the pest population was recorded at weekly interval and the mean number of larvae per plant at different crop stages in different treatments were analyzed and presented in Table.7. A total of five sprays were given, one during vegetative (32 DAS), two during flowering (47 & 62 DAS), one during pod formation (78 DAS) and one during preharvest (94 DAS) stages of the crop.

#### Vegetative stage

Endosulfan gave less than 50% control of small size larvae during vegetative stage with the lowest mean number of larvae per plant (0.47), but HaNPV (0.52) was also as efficient as endosulfan in managing small size larvae. IPM (0.56) which received neem as first spray in addition to bird perches and neem (0.61) were on par and came next in the order of efficacy. Bird perches alone showed no effect (0.90 larva per plant) and was on par with control plot which recorded the highest larval population (0.93).

#### Flowering stage

Neem was recorded lowest mean number of larvae per plant (0.79) but endosulfan (0.92) and IPM (0.95) which received HaNPV as second spray and endosulfan as third spray were also as effective as neem in managing small larvae. HaNPV recorded 0.97 larva/plant and stood next in the order of efficacy. But bird perches (1.30) showed no effect being on par with control. In control the highest larval population was recorded (1.33).

#### Pod formation stage

Control and bird perches plots had the highest population i.e., 1.03 and 0.98 larvae per plant, respectively during the pod formation stage. IPM that received neem as fourth spray was superior with the lowest mean number of small larvae during this stage (0.62). Endosulfan (0.67), HaNPV (0.71) and neem (0.73) were also as effective as IPM treatment.

#### Preharvest stage

IPM plot continued its efficiency in keeping small larvae at the lowest level (0.43 larva per plant). HaNPV (0.52) and endosulfan (0.52) showed

Treatment/ Ve Crop stage	egetative	Flowering (Mean number o	Pod formation f larvae per plant)	Preharvest
Neem 1750 ml/ha	0.61	0.79	0.73 .	0.54
(Nivaar 1500 ppm)	• (0.7821)°	(0.8895)ª	(0.8512)*	(0.7329)⁵
HaNPV250LE/ha	0.52	0.97	0.71	0.52
	(0.7214) <sup>ab</sup>	(0.9829) <sup>6</sup>	(0.8394)ª	(0.7177)ª <sup>b</sup>
Bird perches	0.90	1.30	0.97	0.79
one/plot	(0.9506)⁴	(1.1381)⁰	(0.9835) <sup>ь</sup>	(0.8907)°
Endosulfan 0.07%	0.47	0.92	0.67	0.52
	(0.6829)°	(0.9594) <sup>nb</sup>	(0.8168)ª	(0.7221) <sup>ab</sup>
IPM	0.56	0.95	0.62	0.43
	(0.7469) <sup>⊳</sup>	(0.9744) <sup>ab</sup>	(0.7855)*	(0.6587)"
Control	0.93	1.33	1.03	0.90
	(0.9638) <sup>d</sup>	(1.1545)°	(1.0122)⁵	(0.9488)⁰
S.Ed.	0.028	0.042	0.042	0.032
CD	0.060	0.089	0.088	0.067

## Table 7 : Efficacy of the treatments in managing small size larvae of H.armigera during rabi 1999-2000

(Figures in parenthesis are square root transformed values)

almost equal efficiency and were on par with IPM. Neem (0.54) came next in the order. Bird perches (0.79) showed no effect and was found to be on par with control (0.90).

# 4.1.10 Efficacy of the Treatments on Medium Size (third & fourth instars) Larvae of *H.armigera* During *rabi* 1999-2000

To assess the efficacy of different IPM components in managing medium size larvae of *H.armigera*, an experiment was conducted and the number of larvae per plant were recorded at weekly interval. The mean number of larvae per plant in different treatments at various crop stages were calculated and presented in Table.8.

#### Vegetative stage

The crop was sprayed with the respective treatments by 32 DAS which coincided with vegetative stage. The results revealed that the highest reduction in the number of larvae was in the plots treated with HaNPV (0.33). IPM (0.34) and neem (0.36) were also found to be statistically on par with HaNPV. Endosulfan (0.38) stood next in the order of efficacy. Bird perches (0.55) was the least effective among the treatments but significantly superior over control (0.63).

#### Flowering stage

During flowering stage the crop received two sprays at 47 & 62 DAS. IPM treatment which received HaNPV as second spray and endosulfan as third spray recorded the lowest number of larvae per plant (0.20) but it was on par with HaNPV with 0.23 larva per plant. The number of larvae per plant in the treatments endosulfan and neem were found to be on par (0.25, 0.29, respectively) and stood next. Bird perches with 0.48 larva per plant was the least effective among the treatments but significantly superior compared to control (0.71).

Treatment/ Crop stage	Vegetative Me	Flowering Flannumber of la	Pod formation Irvae per plant	Preharvest
Neem 1750 ml/ha	0.36	0.29	0.58	0.32
(Nivaar 1500 ppm)	(0.6018)ª <sup>b</sup>	(0.5400)°	(0.7580)ªb	(0.5642) <sup>ь</sup>
HaNPV250LE/ha	0.33	0.23	0.54	0.26
	(0.5735)ª	(0.4741) <sup>ab</sup>	(0.7315)ª	(0.5120)ª⁵
Bird perches one/p	lot 0.55	0.48	0.95	0.74
	(0.7415)°	(0.6920)⁴	(0.9719)⁰	(0.8580) <sup>d</sup>
Endosulfan 0.07%	0.38	0.25	0.66	0.57
	(0.6188) <sup>ь</sup>	(0.5039) <sup>₀</sup>	(0.8121) <sup>ь</sup>	(0.7539)⁰
IPM	0.34	0.20	0.60	0.23
	(0.5849)ª⁵	(0.4414)ª	(0.7760)ªb	(0.4838)*
Control	0.63	0.71	1.18	0.81
	(0.7902)ª	(0.8404)°	(1.0872) <sup>d</sup>	(0.8977)ª
S.Ed.	0.021	0.029	0.028	0.029
CD	0.045	0.062	0.059	0.062

Table 8 : Efficacy of the treatments in managing medium size larvae of *H. armigera* during rabi 1999-2000

(Figures in parenthesis are square root transformed values) \*Values followed by same letters in each column are statistically not significant

#### Pod formation stage

The crop was sprayed with the respective treatments at 78 DAS coinciding with pod formation stage. The results revealed that the highest reduction in the number of larvae was in the plots treated with HaNPV (0.54 larva per plant) but it was at a par with neem and IPM treatments which received neem as fourth spray, with 0.58, 0.60 larva/plant, respectively. Endosulfan treatment (0.66) stood next in the order of efficacy. Bird perches (0.95) showed little but significantly superior to control (1.18).

#### Preharvest stage

The crop was sprayed with the respective treatments by 94 DAS during preharvest stage. IPM was the most effective treatment with the lowest number of larvae per plant (0.23). HaNPV with 0.26 larva per plant was as effective as IPM. Neem treatment (0.32) came next in the order. Endosulfan (0.57) although less effective compared to the earlier treatments, was significantly superior to bird perches which showed no effect (0.74) and was found to be on par with control (0.81).

## 4.1.11 Effect of the Treatments on Large Size (fifth & sixth instar) Larvae of *H.armigera* During *rabi* 1999-2000

The effect of IPM components individually and in combination in managing large size larvae of *H.armigera*, was evaluated during 1999-2000. Five sprays were given, one at vegetative stage, two at flowering stage, one at podding and one at preharvest stage. The larval population was observed at weekly interval and the mean number of larvae per plant in different treatments at different stages was assessed and presented in Table 9.

#### Vegetative stage

For managing large size larvae HaNPV was found to be the most effective with the least number of larvae per plant (0.03) but was on par with bird perches (0.04). Birds activity was more during this stage (Table 10). IPM plots that received neem as first spray (0.058) and neem (0.058) and endosulfan (0.06) were being on par and registered significantly low late instar larval population compared to control (0.10).

#### Flowering stage

IPM plots that received HaNPV as second spray and endosulfan as third spray and HaNPV recorded lowest number of large size larvae (0.02). Neem stood next in the order of efficacy with 0.03 larva per plant. Endosulfan (0.06) and bird perches (0.08) were found to be on par and significantly superior compared to control (0.22).

#### Pod formation stage

IPM that received neem as fourth spray continued its efficiency in managing large size larvae (0.26), however it was found on par with endosulfan (0.27), HaNPV (0.27) and bird perches (0.28). Neem with 0.30 larva per plant came closely behind and was significantly superior to control (0.40).

#### Preharvest stage

IPM that received HaNPV as fifth spray was superior in managing large size larvae with 0.02 larva per plant but was on par with HaNPV (0.04) and bird perches (0.05). Endosulfan (0.09) and neem (0.13) were on par and came next in the order of efficacy and were significantly superior over control (0.36).

Treatment/ Vege	tative Flov	wering Poc	l formation	Preharvest
Crop stage	(Mean n	umber of larv	ae per plant)	
Neem 1750 ml/ha	0.58	0.03	0.30	0.13
(Nivaar 1500 ppm)	(0.3289) <sup>bc</sup>	(0.2885) <sup>bc</sup>	(0.5470)⁵	(0.4256)⁰
HaNPV250LE/ha	0.03	0.02	0.27	0.04
	(0.2885)ª	(0.2660) <sup>ab</sup>	(0.5189)ª <sup>b</sup>	(0.3009) <sup>ab</sup>
Bird perches one/plot	0.04	0.08	0.28	0.05
	(0.2956) <sup>sb</sup>	(0.3534)⁴	(0.5331) <sup>ab</sup>	(0.3210)²⁵
Endosulfan 0.07%	0.06	0.06	0.27	0.09
	(0.3353)⁰	(0.3353)∝d	(0.5188)⁵⁵	(0.3758) <sup>bc</sup>
IPM	0.06	0.02	0.26	0.02
	(0.3289)∞	(0.2234)ª	(0.5092)*	(0.2678)*
Control	0.10	0.22	0.40	0.36
	(0.3817)⁴	(0.5229) <sup></sup>	(0.6301)"	(0.8764) <sup>d</sup>
S.Ed.	0.016	0.025	0.017	0.037
CD	0.035	0.052	0.037	0.079

## Table 9 : Efficacy of the treatments in managing large size larvae of *H.armigera* during *rabi* 1999-2000

(Figures in parenthesis are square root transformed values)

ומחום		זאבו אבת זון חוב הווהעאב		
SI.No.	Common Name	Scientific Name	Food Habits	Remarks
-	Black Drongo/King Crow	Dicrurus adismilis (Bech.)	Grass hoppers and other insects, essentially needs a perch to watch and sally for the prey	Highly beneficial to agriculture by the vast quantities of insects it destroys
2	Roller/Blue Jay	Coracias benghalensis (Lin)	Large insect, frog or lizard on the ground, re- turning to either to the same perch or flying leisurely across to another near by.	Highly beneficial to agriculture, since it destroys vast quantities of injurious insects.
ю	Cattle Erget	Bubulcus ibis (Lin.)	Chiefly grasshoppers, blue bottles, cicadas and caterpillars; also frogs, lizards, fish etc.	Highly beneficial in both dry and wet agro ecosystems
4	Paddy Field Pipit	Anthus novaeseel- andiae (Lin.)	Weevils and other small insects. Runs about briskly inspurts, searching for the prey.	Mostly seen during and after ploughing in the agricultural fields.
ۍ	Common Swallow	Hirundo rustica (Lin.)	Hawks winged insects high up in air or close to the ground.	Beneficial to agriculture.
9	Indian Myna	Acridothers tristis (Lin.)	Omnivorus. Eats fruits, insects, kitchen scraps.	Follows the plough for insects, pupae. etc.
7	Red Winged Bush Lark	Mirafa erythroptera (Blyth)	Seeds of grass and weeds, insects.	Beneficial to Agriculture.
80	Red Wattled Lapwing	Vanellus indicus (Bodd.)	Insects, their eggs and larvae.	Affects open country, ploughed fields. graz- ing land, tanks and puddles. A beneficial bird in Natural pest control.
თ	lora	Aegithina tiphia (Lin.)	insects, grubs. mollusks, etc	Arboreal. affects village outskirts and sec- ondary jungle. Beneficial in agriculture and horticulture.
10	Baya Weaver Bird	Ploceus hilippinus (Lin.)	Gleans paddy and other grain-harvested fields. Also eats insects.	Seen abundantly in ICRISAT, Patancheru campus, Beneficial in insect pest control
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Table 10: List of birds observed in the chickpea field-their identification and habits

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## 4.1.12 Efficacy of the Treatments on the Total Larval Load of *H.armigera* During rabi 1999-2000

To assess the effect of different IPM components in managing *H.armigera* larval population on chickpea, an experiment was conducted with six treatments. The number of larvae were recorded at weekly interval in different experimental plots. The mean number of larvae of all stages per plant in different treatments during different crop stages was calculated and presented in Table 11.

#### Vegetative stage

The crop received one spray with the respective treatments by 32 DAS during vegetative stage. The results revealed the highest reduction in the number of larvae in the plots treated with HaNPV (0.88) but it was on par with endosulfan (0.91). IPM which received neem as first spray in addition to bird perch with 0.94 larva per plant, came next and was found to be significantly superior to the remaining treatments. Next were neem (1.02) and bird perches (1.05) being on par and gave significantly superior control of *H.armigera* larvae compared to control plots (1.65).

#### Flowering stage

The crop received two sprays at 47 & 62 DAS during flowering stage. IPM (1.21 larvae/ plant) which received HaNPV as second spray and endosulfan as third spray, HaNPV (1.21 larvae/plant), and endosulfan (1.24 larvae/plant) were the best effective treatments being on par and significantly superior to the remaining treatments. Neem stood next with 1.33 larvae per plant in the order of efficacy. Bird perch was the least effective with 1.85 larvae per plant among the treatments but it was significantly superior compared to control which recorded 2.26 larvae per plant.

Treatment/	Vegetative Flowering Pod formation Preharvest					
Crop stage	(Mean number of larvae per plant)					
Neem 1750 ml/ha	1.02	1.33	1.60	0.90		
(Nivaar 1500 ppm)	(1.0101)º	(1.1545)⁵	(1.2644) <sup>b</sup>	(0.9501) <sup>b</sup>		
HaNPV250LE/ha	0.88	1.21	1.51	0.82		
	(0.9397)ª	(1.1009)ª	(1.2284)ª	(0.9047)⁵		
Bird perches one/plo	ot 1.05	1.85	2.20	1.58		
	(1.0225)°	(1.3600)⁰	(1.4820)°	(1.2585)⁴		
Endosulfan 0.07%	0.91	1.24	1.60	1.17		
	(0.9550) <sup>ab</sup>	(1.1122)ª	(1.2634) <sup></sup> ,	(1.0808)⁰		
IPM	0.94	1.21	1.48	0.69		
	(0.9786)⁵	(1.1001)ª	(1.2160)ª	(0.8307)ª		
Control	1.65	2.26	2.60	1.84		
	(1.2843)⁴	(1.5044)⁴	(1.6135)⁴	(1.3552)°		
S.Ed.	0.011	0.019	0.016	0.025		
CD	0.024	0.041	0.034	0.053		

## Table 11 : Efficacy of the treatments in managing total larval load of H.armigera during rabi 1999-2000

(Figures in parenthesis are square root transformed values)

#### Pod formation stage

The crop was sprayed with the respective treatments by 78 DAS coinciding with pod formation stage. IPM which received neem as fourth spray and HaNPV were on par and significantly superior to the rest of the treatments in managing *H.armigera* larvae with 1.48 and 1.51 larvae per plant. Endosulfan (1.60 larvae/plant) and neem (1.60 larvae/plant) were on par and stood next in the order of efficacy. Bird perches (2.20) was less effective but significantly superior compared to control which recorded the highest mean number of larvae per plant (2.60).

#### Preharvest stage

The crop received one spray at 94 DAS during the preharvest stage. IPM treatment which received HaNPV as fifth spray was the most effective with the highest reduction in larval population during this stage (0.69). HaNPV with 0.82 larva per plant and neem with 0.90 larva per plant were on par and stood next in the order of efficacy and significantly superior compared to endosulfan (1.17). Bird perches was the least effective with 1.58 larvae per plant among the treatments but significantly superior compared to control which recorded 1.84 larvae per plant.

### 4.1.13 Mean Efficacy of the Treatments on Oviposition Behaviour of *H.armigera* in chickpea During Two Years

The mean data computed on oviposition behaviour of *H.armigera* during two years are presented in Table.12 (Figure.3).

#### Vegetative stage

The plots treated with neem recorded the lowest number of eggs per plant (0.35) during vegetative stage but it was on par with IPM (0.39) which received neem as first spray. The treatments HaNPV and endosulfan were on par and stood next with less number of eggs (0.49, 0.52 egg/

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage		(Mean numbe	r of eggs per plar	nt)
Neem	0.35	0.72	0.31	0
	(0.5874) <sup>*</sup>	(0.8485) <sup>°</sup>	(0.5522) <sup>°</sup>	(0.2236) <sup>"</sup>
HaNPV250LE/ha	0.49	0.83	0 .41	0.05
	(0.7000) <sup>bc</sup>	(0.9082) <sup>b</sup>	(0.6403) <sup>°</sup>	(0.3162) <sup>bc</sup>
Bird perches one/plo	t 0.60	0.96	0.42	0.07
	(0.7714) <sup>cd</sup>	(0.9772)°	(0.6442) <sup>°</sup>	(0.3391) <sup>°</sup>
Endosulfan 0.07%	0.52	0.97	0.38	0.03
	(0.7211) <sup>°</sup>	(0.9823)°	(0.6164) <sup>⊮⊄</sup>	(0.2738) <sup>ab</sup>
IPM	0.39	0.82	0.32	0.01
	(0.6204) <sup>®b</sup>	(0.9027) <sup>ab</sup>	(0.5656) <sup>ab</sup>	(0.2345) <sup>°</sup>
Control	0.76	1.17	0.55	0.08
	(0.8689) <sup>d</sup>	(1.0793) <sup>ď</sup>	(0.7416) <sup>ď</sup>	(0.3505) <sup>°</sup>
S.Ed.	0.046	0.027	0.030	0.028
CD	0.098	0.058	0.063	0.059

Table	12:	Mean	efficacy	of the	e treatments	on	oviposition	behaviour	of
		H. ar	migera						

(Figures in parenthesis are square root transformed values)





plant, respectively). Bird perches (0.60) showed poor efficacy and was found on par with control (0.76 egg/plant).

#### Flowering stage

During flowering stage also neem (0.72) was found superior in keeping the egg number at low level. IPM and HaNPV (0.82, 0.83 eggs/ plant, respectively) were found on par in reducing the oviposition. Bird perches and endosulfan plots (0.96, 0.97 egg/plant, respectively) were on par and recorded significantly reduced egg number compared to control (1.17 eggs/plant).

#### Pod formation

Neem and IPM which received neem as first and fourth spray were on par and recorded the lowest number of eggs/plant (0.31, 0.32 egg/plant, respectively). All the remaining treatments Viz., endosulfan (0.38), HaNPV (0.41) and bird perches (0.42) were on par and recorded significantly less number of eggs compared to control.

#### Preharvest stage

During this period also neem treatment (0) provided maximum repellency for *H.armigera* oviposition. IPM (0.01) and endosulfan (0.03) treatments were also on par with neem and recorded less number of eggs. HaNPV (0.05) and bird perches (0.07) were on par with control treatment (0.08)

## 4.1.14 Mean Efficacy of the Treatments Against Small Size Larvae of *H.armigera* in Chickpea During Two Years

The mean data of the two years on the effect of different IPM components on small size larvae of *H.armigera* are presented in Table 13 (Figure 4).

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#### Vegetative stage

The data revealed that during this stage the plots sprayed with endosulfan recorded the lowest number of larvae per plant (0.69). The treatments HaNPV (0.81) and IPM (0.84) were also as effective as endosulfan. Neem (0.90) was found next in the order of efficacy. Bird perches (1.08) showed no effect on small size larval population and was on par with control (1.20).

#### Flowering

During this stage neem (1.11), IPM (1.12), endosulfan (1.13) and HaNPV (1.18) were on par and recorded significantly less number of larvae compared to the remaining two treatments. Bird perches (1.57) showed no effect in managing small size larvae and was on par with control (1.61).

#### Pod formation stage

IPM treatment (0.89 larva/plant) was the most effective with the lowest number of larvae per plant. HaNPV (0.97) and endosulfan (1.00) and neem (1.01) came next and were on par and found to be significantly superior over the remaining treatments. Bird perches (1.16) showed no effect and was on par with control (1.26).

#### Preharvest stage

Based on the results, IPM proved significantly superior in keeping the larval population at the lowest level with 0.35 larva/plant. Endosulfan (0.73), HaNPV (0.74), neem (0.76) and bird perches (0.90) were on par and significantly superior over control which recorded the highest larval number (1.08).

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage		(Mean number	of larvae per pl	ant)
Neem	0.90	1.11	1.01	0.76
	(0.9460) <sup>♭</sup>	(1.0535) <sup>°</sup>	(1.0024) <sup>⁵</sup>	(0.8689) <sup>b</sup>
HaNPV250LE/ha	0.81	1.18	0.97	0.74
	(0.8972) <sup>#b</sup>	(1.0839) <sup>°</sup>	(0.9848) <sup>ab</sup>	(0.8573) <sup>b</sup>
Bird perches one/plo	ot 1.08	1.57	1.16	0.90
	(1.0368) <sup>°</sup>	(1.2529) <sup>⋫</sup>	(1.0747) <sup>°</sup>	(0.9460) <sup>be</sup>
Endosulfan 0.07%	0.69	1.13	1.00	0.73
	(0.8306) <sup>"</sup>	(1.0630) <sup>"</sup>	(0.9874) <sup>"b</sup>	(0.8544) <sup>"</sup>
IPM	0.84	1.12	0.89	0.35
	(0.9165) <sup>®</sup>	(1.0583) <sup>°</sup>	(0.9433) <sup>"</sup>	(0.5916) <sup>"</sup>
Control	1.20	1.61	1.26	1.08
	(1.0954) <sup>°</sup>	(1.2668) <sup>⊮</sup>	(1.1224) <sup>°</sup>	(1.0368) <sup>°</sup>
S.Ed.	0.042	0.042	0.028	0.045
CD	0.088	0.089	0.059	0.095

# Table 13: Mean efficacy of the treatments on small size larvae of H. armigera

(Figures in parenthesis are square root transformed values)



## 4.1.15 Mean Efficacy of the Treatments Against Medium Size Larvae of *H.armigera* in Chickpea During Two Tears

The data on cumulative effect of different IPM components on medium size larvae of *H.armigera* during both the years are presented in Table 14 (Figure.5).

#### Vegetative stage

HaNPV treated plot was superior in managing the medium size larvae with 0.41 larva/plant however it was found on par with neem with 0.42 larva/plant and IPM with 0.45 larva/plant. Endosulfan (0.47) stood next in the order of efficacy. Bird perches also showed significant reduction in medium size larvae (0.59) compared to control (0.71).

#### Flowering stage

During this stage IPM (0.39) was the most effective treatment with the lowest number of larvae/plant. HaNPV (0.43), endosulfan (0.45) and neem (0.46) were on par and significantly superior over the remaining two treatments in managing medium size larvae. Bird perches plot also gave significantly reduced number of larvae per plant (0.60) compared to control (0.80).

#### Pod formation stage

IPM was more efficient in managing medium size larvae with 0.64 larva/plant.  $H_{NPV}^{\circ}$  with 0.69 larva/plant was as effective as IPM. Neem (0.74 and endosulfan (0.78) were inturn on par with HaNPV and stood next in the order of efficacy. Bird perches plot also gave a significant reduction in the number of larvae per plant (0.88) compared to control (1.09).

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage		Mean number	of larvae per plan	t
Neem	0.42	0.46	0.74	0.50
	(0.6480) <sup>*</sup>	(0.6745) <sup>⋫</sup>	(0.8602) <sup>b</sup>	(0.7071) <sup>b</sup>
HaNPV250LE/ha	0.41	0.43	0.69	0.42
	(0.6403) <sup>ª</sup>	(0.6519) <sup>ab</sup>	(0.8276) <sup>ab</sup>	(0.6480) <sup>°</sup>
Bird perches one/plo	t 0.59	0.60	0.88	0.68
	(0.7648)°	(0.7745) <sup>°</sup>	(0.9380) <sup>°</sup>	(0.8215) <sup>d</sup>
Endosulfan 0.07%	0.47	0.45	0.78	0.59
	(0.6855) <sup>⁵</sup>	(0.6670) <sup>b</sup>	(0.8803) <sup>⁵</sup>	(0.7681) <sup>°</sup>
ЮМ	0.45	0.39	0.64	0.36
	(0.6708) <sup>ab</sup>	(0.6244) <sup>ª</sup>	(0.8000) <sup>a</sup>	(0.6000) <sup>°</sup>
Control	0.71	0.80	1.09	0.83
	(0.8426) <sup>ď</sup>	(0.8944) <sup>d</sup>	(1.0440) <sup>d</sup>	(0.9082) <sup>°</sup>
S.Ed.	0.015	0.018	0.020	0.024
CD	0.033	0.038	0.053	0.051

Table 14: Mean efficacy of the treatments on medium size larvae of H. armigera

(Figures in parenthesis are square root transformed values) \*Values followed by same letters in each column are statistically not significant



H. armigera in chickpea.



#### Preharvest stage

At preharvest stage also IPM (0.36) showed its efficiency in managing the medium size larvae. But HaNPV with 0.42 larva/plant was also as effective as IPM. Neem (0.50) came next and was significantly superior compared to endosulfan (0.59). Bird perches plot also gave a significant reduction in number of larvae per plant (0.68) compared to control (0.83).

### 4.1.16 Mean efficacy of the Treatments Against Large Size Larvae of *H.armigera* in Chickpea During Two Years

The data on cumulative effect of different IPM components on large size larvae of *H.armigera* during both years are presented in Table 15 (Figure 6).

#### Vegetative stage

HaNPV showed better effect in controlling large size larvae with 0.11 larva/plant during vegetative stage. Endosulfan and IPM were on par and stood next in the order of efficacy with 0.12 larva/plant. Neem (0.12) and bird perches (0.13) came next and found to be significantly superior over control (0.21).

#### Flowering stage

IPM was the most effective with 0.05 larva/plant and significantly superior compared to other treatments in managing large size larvae. HaNPV (0.08), neem (0.08) and endosulfan (0.08) were on par and stood next in the order of efficacy. Bird perches was the least effective (0.11) but significantly superior over control (0.25).

Treatment/ Crop stage	Vegetative	Flowering (Mean numbe	Pod formation	Preharvest	
orop orage		(mean namber et la rue per pr			
Neem	0.12	0.08	0.26	0.09	
	(0.3521) <sup>°</sup>	(0.2738) <sup>b</sup>	(0.5049) <sup>°</sup>	(0.30) <sup>°</sup>	
HaNPV250LE/ha	0.11	0.08	0.21	0.05	
	(0.3316) <sup>°</sup>	(0.2737) <sup>⁵</sup>	(0.4582) <sup>ªb</sup>	0.2236) <sup>°b</sup>	
Bird perches one/plo	t 0.13	0.11	0.24	0.07	
	(0.3535) <sup>°</sup>	(0.3316) <sup>°</sup>	(0.4847) <sup>bc</sup>	(0.2549) <sup>56</sup>	
Endosulfan 0.07%	0.12	0.08	0.22	0.08	
	(0.3391) <sup>ab</sup>	(0.2828) <sup>♭</sup>	(0.4690) <sup>ab</sup>	(0.2738) <sup>56</sup>	
IPM	0.12	0.05	0.21	0.04	
	(0.3449) <sup>∞</sup>	(0.2236) <sup>°</sup>	(0.4527) <sup>°</sup>	(0.2000)"	
Control	0.21	0.25	0.38	0.38	
	(0.4582) <sup>"</sup>	(0.5000) <sup>4</sup>	(0.6164) <sup>d</sup>	(0.6123) <sup>4</sup>	
S.Ed.	0.006	0.011	0.013	0.025	
CD	0.013	0.035	0.029	0.052	

Table 15: Mean efficacy of the treatments on large size larvae of *H. armigera* 

(Figures in parenthesis are square root transformed values)



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#### Pod formation stage

IPM was once again proved more effective with the lowest number of larvae per plant (0.21) during this stage, however it was found on par with HaNPV (0.21) and endosulfan (0.22). Bird perches (0.24) and neem (0.26) were on par and significantly superior over control (0.38) in managing large size larval population at pod formation stage.

#### Preharvest stage

IPM continued its superiority in managing large size larvae during this stage also with 0.04 larva/plant and was on par with HaNPV (0.05). Bird perches (0.07), endosulfan (0.08) and neem (0.09) were next being on par and significantly superior to control which recorded the highest larval population (0.38).

### 4.1.17 Mean efficacy of The Treatments Against the Total Larval Load of *H.armigera* in Chickpea During Two Years

Data on the mean effect of different IPM components in managing total larval load are presented in Table 16 (Figure 7).

#### Vegetative stage

The results (Table 15) revealed a significant reduction in larval number in the plots treated with endosulfan (1.25) and HaNPV (1.33) (Plate 7). Neem (1.44) and IPM (1.47) which received neem as first spray were on par and stood next in the order of efficacy. Bird perches was less effective with 1.58 larvae/plant but significantly superior over control (2.09).

#### Flowering stage

IPM was significantly the most effective treatment with 1.58 larvae/ plant. Endosulfan (1.66) and HaNPV (1.67) stood next and were on par

n. armigera				
Treatment/ Crop stage	Vegetative	Flowering Pod formation Mean number of larvae per plant		Preharvest
Neem	1.44	1.75	2.00	1.30
	(1.1979) <sup>⁵</sup>	(1.3228) <sup>°</sup>	(1.4124) <sup>b</sup>	(1.1401) <sup>⁵</sup>
HaNPV250LE/ha	1.33	1.67	1.87	1.21
	(1.1532) <sup>°</sup>	(1.2922) <sup>۴</sup>	(1.3656) <sup>ªb</sup>	(1.0977) <sup>6</sup>
Bird perches one/plo	t 1.58	2.27	2.27	1.63
	(1.2549) <sup>°</sup>	(1.5049) <sup>d</sup>	(1.5066) <sup>°</sup>	(1.2767) <sup>°</sup>
Endosulfan 0.07%	1.25	1.66	1.97	1.39
	(1.1157) <sup>°</sup>	(1.2884) <sup>⁵</sup>	(1.4035) <sup>ັ</sup> ້	(1.1768) <sup>°</sup>
.IPM	1.47	1.58	1.74	1.00
	(1.2124) <sup>⋫</sup>	(1.2569) <sup>°</sup>	(1.3190) <sup>*</sup>	(0.9998) <sup>°</sup>
Control	2.09	2.63	2.75	2.06
	(1.4439) <sup>d</sup>	(1.6217) <sup>°</sup>	(1.6568) <sup>d</sup>	(1.4352) <sup>d</sup>
S.Ed.	0.019	0.011	0.025	0.046
CD	0.041	0.024	0.053	0.096

Table 16: Mean efficacy of the treatments on total larval load of H. armigera

(Figures in parenthesis are square root transformed values)










Plate 7: HaNPV infected larvae of H. armigera in chickpea.



late 8: Natural incidence of larval parasitoid Campoletis chlorideae Uchida.

and significantly superior compared to neem (1.75). Bird perches was the least effective with 2.27 larvae/plant among the treatments but significantly superior over control (2.63).

#### Pod formation stage

IPM was once again the most effective with the lowest number of larvae per plant (1.74). HaNPV with 1.87 larvae/plant was found on par with IPM. Endosulfan (1.97) and neem (2.00) were on par and stood next in the order of efficacy. Bird perches (2.27) was the least effective but significantly superior over control which recorded high larval load (2.75).

#### Preharvest stage

IPM maintained its superiority during this stage also with 1.00 larve/ plant being the most effective and significantly superior to all the remaining treatments. HaNPV (1.21), neem (1.30) and endosulfan (1.39) were on page and stood next in the order. Bird perches was the least effective treatment with 1.03 larvae/plant but significantly superior over control (2.06).

## 4.2 EFFECT OF THE TREATMENTS ON NATURAL ENEMIES PRESENT IN CHICKPEA ECOSYSTEM

## 4.2.1 Effect of the Treatments on Soil Inhabiting Natural Enemies During rabi 1998-99

In order to assess the effect of different IPM components on soil inhabiting natural enemies, an experiment was conducted during *rabi* 1998-99 season and the results are presented in Table.17 (Figure.8). The number of natural enemies was observed in pitfall traps at ten days interval. Natural enemies such as ants, braconid wasps, ichneumonid wasps (Hymenoptera), ground beetles, coccinellids (Coleoptera), crickets (Orthoptera), earwigs (Dermaptera) and spiders were collected from the pitfall traps fixed in each treatment. A total of five sprays, two during vegetative, one during flowering and two during pod formation stage were given. The mean number of natural enemies present in different treatments was calculated.

### Vegetative stage

During the vegetative stage the plots treated with endosulfan recorded significantly less number of natural enemies (136.5 natural enemies/ trap). The plots treated with neem, IPM which received neem as first spray stood next with lowest number of natural enemies (245.8, 244.6 natural enemies/ trap, respectively). HaNPV (385.6/trap) and bird perches (331.5/trap) did not show any significant effect on natural enemies and were on par with control (380.0/trap).

#### Flowering stage

At flowering stage also endosulfan showed toxic effect on natural enemies present on ground (115.8/trap) and recorded the lowest number of natural enemies. Neem also affected the natural enemies (230.4/trap) followed by IPM (231.9/trap) that received endosulfan as third spray. Bird perches (302.8/trap) showed little but not significant effect on ground dwelling natural enemies compared to control. HaNPV (572.8/trap) and control (513.0/trap) were found to be on par.

## Pod formation stage

During podding stage also endosulfan was found toxic to ground inhabiting natural enemies up to 70% (40.8/trap) compared to control. Neem also reduced up to 50.5% (69.3/trap) of natural enemies compared to control. IPM that received neem as fourth spray and HaNPV as fifth spray reduced natural enemies up to 25% (104.6/trap) but not statistically

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage	(M	ean number of	f natural enemies	/trap)
Neem 0.006%	245.75	230.36	69.25	66.75
(AZA 3%)	(15.65) <sup>⊾</sup>	(15.17) <sup>ab</sup>	(8.32)* <sup>b</sup>	(8.17)*
HaNPV250LE/ha	385.63	572.75	138.38	62.50
	(19.64)⁰	(23.93)⁴	(11.76)⁰	(7.90)ª
Bird perches one/plot	331.50	302.75	111.38	69.50
	(18.20) <sup>⊳</sup>	(17.39) <sup>be</sup>	(10.55) <sup>bc</sup>	(8.33)"
Endosulfan 0.07%	136.50	115.75	40.75	48.25
	(11.68)ª	(10.76) <b>•</b>	(6.38)≝	(6.94)"
IPM	244.63	231.88	104.63	75.75
	(15.65)⁵	(15.22)ªb	(10.23) <sup>₅</sup>	(8.70)*
Control	380.00	513.00	140.00	73.25
	(19.47)⁰	(22.61) <sup>∞d</sup>	(11.80)°	(8.56)ª
S.Ed.	1.725	2.643	1.063	0.909
CD	3.678	5.634	2.266	1.912

Table	17:	Effect	of	the	treatmen	ts on	soil	inhabiting	natural	enemies	in
		chickp	ea	dur	ing <i>rabi</i> '	1998-1	999.				

(Figures in parenthesis are square root transformed values)

\*Values followed by same letters in each column are statistically not significant



significant from HaNPV and bird perches which showed no effect and were found on par with control (140.0/trap).

### Preharvest stage

At preharvest stage there was no significant difference in number of natural enemies among all the treatments.

## 4.2.2 Effect of the Treatments on Natural Enemies Present on Chickpea Crop Canopy During *rabi* 1998-99

To assess the effect of different IPM components on aerial natural enemies, a De Vac trap was used and the data on the natural enemies observed in the trap collection are presented in Table.18 (Figure.9). Natural enemies belonging to the order Hymenoptera such as braconids, chalcidids, ichneumonids, trichogrammatids, ants and others such as spiders, small crickets, tachinids etc., were considered for obsevations. During *rabi* 1998-99 at 22 days after sowing (one day after treatment) the first sampling of aerial natural enemies was done with De Vac. The results suggested a maximum reduction in natural enemies present on foliage in plots treated with endosulfan, neem and IPM (39.5, 50.0, 51.0 per two rows respectively). There was no significant reduction of the number of natural enemies present in plots treated with HaNPV (69.7/two rows) and bird perches (84.7/ two rows) which were found on par with control (87.1/two rows).

At 54days after sowing i.e., two days after 3<sup>rd</sup> spraying, the second sampling of aerial natural enemies was done. The results suggested a significant reduction in the number of natural enemies present on foliage in the plots treated with endosulfan and IPM treatment which received endosulfan as third spray (9.8, 11.4 per two rows, respectively). No

Treatment	Number of natural enemies/two rows (36m)				
	22DAS(1DAT)	54DAS(2DAT)			
Neem 0.006%	50.00	20.83			
(AZA 3%)	(7.06) <sup>ab</sup>	<b>. (4</b> .56)			
HaNPV 250LE/ha	69.73	21.45			
	(8.35) <sup>⊾</sup>	(4.63) <sup>b</sup>			
Bird perches one/plot	84.69	23.80			
	(9.20)°	(4.88) <sup>b</sup>			
Endosulfan 0.07%	39.45	9.79			
	(6.28) <b>*</b>	(3.13)"			
IPM	51.00	11.39			
	(7.08) <sup>ab</sup>	(3.37)*			
Control	87.06	23.79			
	(9.33)°	(4.86) <sup>b</sup>			
Sed	0.630	0.341			
CD	1.343	0.726			

Table 18 : Effect of the treatments on natural enemies present on crop canopy during rabi 1998-99

Mean of 4 replications

DAS = Days after sowing.

DAT = Days after treatment.

(Figures in parenthesis are square root transformed values)

\*Values followed by same letters in each column are statistically not significant



 $\mathsf{Fig.~9}$  : Effect of the treatments on natural enemies present on crop canopy during

significant reduction of natural enemies was observed in the plots treated with neem, HaNPV and bird perches compared to control (20.8, 21.5, 23.8, 23.8 per two rows, respectively).

## 4.2.3 Effect of the Treatments on Soil Inhabiting Natural Enemies During *rabi* 1999-2000

An experiment was conducted during *rabi* 1999-2000 in order to assess the effect of different IPM components individually and in combination on soil inhabiting natural enemies present during the crop growth. Observations on natural enemies in different treatments were recorded with the help of pitfall traps at ten days interval and the mean number of natural enemies per treatment at different crop stages were calculated. Sprayings were given at fifteen days interval during the period of crop growth (Table.19 & Figure.10).

#### Vegetative stage

During vegetative stage the plots treated with endosulfan recorded significantly less number of natural enemies present on the ground (79.0/ trap). The remaining treatments Viz., HaNPV (148.6/trap), neem (153.3/ trap), bird perches (158.5/trap) and IPM (217/trap) caused negligible reduction in number of natural enemies compared to control (224.7/trap) but the differences were not significant.

### Flowering stage

During flowering stage also the highest reduction of natural enemies was observed in the plots treated with endosulfan (208.3/trap). There was no significant difference among the plots treated with HaNPV, neem, bird perches and IPM which received HaNPV as second spray (236.5, 238.8, 257.5, 294.0 per trap, respectively) being on par and recorded significantly less number of natural enemies compared to control (397.9/trap).

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage	(Mea	n number of r	natural enemies/t	rap)
Neem 1750 ml/ha	153.25	238.75	133.50	62.55
(Nivaar 1500 ppm)	(12.38)⁵	(15.45) <sup>ab</sup>	(11.55)⁵	(8.91)⁵
HaNPV250LE/ha	148.60	236.50	111.43	53.45
	(12.19)⁵	(15.38) <sup>ab</sup>	(10.55) <sup>ab</sup>	(7.31) <sup>ab</sup>
Bird perches one/plot	158.50	257.52	157.25	49.55
	(12.59) <sup>ь</sup>	(16.04) <sup>ab</sup>	(12.54) <sup>₀</sup>	(7.04)≇ <sup>b</sup>
Endosulfan 0.07%	79.03	208.29	79.86	25.00
	(8.89)≋	(14.43)³	(8.93)*	(5.06)*
IPM	216.95	294.00	81.78	83.17
	(14.73)⁵	(17.14) <sup>ь</sup>	(9.04)ª	(9.12)⁵
	224.70	397.88	216.58	102.25
	(14.99)⁵	(19.94)⁰	(14.71)°	(10.11)⁵
S.Ed.	1.397	0.973	1.068	1.552
CD	2.978	2.073	2.246	3.263

# Table 19: Effect of the treatments on soil inhabiting natural enemies in chickpea during rabi 1999-2000

(Figures in parenthesis are square root transformed values)

\*Values followed by same letters in each column are statistically not significant





#### Pod formation stage

At pod formation stage control (216.6/trap) recorded the highest level of soil natural enemy fauna. But endosulfan spray and IPM that received endosulfan as third spray highly reduced the natural enemy fauna (79.9, 81.9/trap, respectively). HaNPV (111.4/trap) and neem (133.5/trap) were on par and stood next in reducing natural enemies that were present on ground. Bird perches (157.3/trap) caused no significant reduction of ground dwelling natural enemies compared to control (216.6/trap).

### Preharvest stage

During preharvest stage endosulfan spray was again found toxic to the ground natural enemies (25.0/trap). All the remaining treatments did not show any significant effect compared to control on soil dwelling natural enemies.

### 4.2.4 Mean effect of the treatments on soil inhabiting natural enemies

The mean data regarding the effect of different IPM components on soil dwelling natural enemies are presented in the Table 20 (Figure.11).

## Vegetative stage

The results revealed a significant reduction in number of natural enemies in plots treated with endosulfan (107.8/trap). The plots treated with neem stood next in reducing the natural enemies (199.5/trap). IPM (230.8), bird perches (245/trap) and HaNPV (267.1/trap) did not show any significant effect and were on par with control (302.4/trap).

### Flowering stage

At flowering stage also endosulfan was found highly toxic to ground dwelling natural enemies (162.0/trap). Neem (234.6/trap), IPM (262.9/trap)

Treatment/	Vegetative	Flowering	Pod formation	Preharvest
Crop stage	(N	lean number of	natural enemies	per trap)
Neem	199.5	234.58	101.38	64.65
	(14.12) <sup>⁵</sup>	(15.31) <sup>ªb</sup>	(10.06) <sup>ªb</sup>	(8.04) <sup>∎b</sup>
HaNPV250LE/ha	267.12	404.63	124.91	57.98
	(16.34) <sup>⊮°</sup>	(20.11) <sup>cd</sup>	(11.17) <sup>50</sup>	(7.61) <sup>ab</sup>
Bird perches one/plo	t 245.0	280.14	134.32	59.53
	(15.65) <sup>bc</sup>	(16.73) <sup>b</sup>	(11.58) <sup>bc</sup>	(7.71) <sup>at</sup>
Endosulfan 0.07%	107.76	162.02	60.31	36.63
	(10.38) <sup>°</sup>	(12.72) <sup>"</sup>	(7.76)"	(6.05) <sup>°</sup>
IPM	230.79	262.94	93.21	79.46
	(15.19) <sup>⊮</sup>	(16.21) <sup>ªbc</sup>	(9.65) <sup>#b</sup> ···	(8.91) <sup>ະ</sup>
Control	302.35	455.44	178.29	87.75
	(17.38)°	(21.34) <sup>d</sup>	(13.35) <sup>°</sup>	(9.36) <sup>⁵</sup>
S.Ed.	1.268	1.904	1.119	1.012
CD	2.658	3.989	2.345	2.121

Table 20: Mean efficacy of the treatments on soil inhabiting natural enemies

(Figures in parenthesis are square root transformed values)

Values followed by same letters in each column are statistically not significant



and bird perches (280.1/trap) were on par and stood next in reducing ground dwelling natural enemies. HaNPV did not show any significant effect on natural enemies compared to control (455.4/trap).

## Pod formation stage

During this stage also endosulfan (60.3/trap) was found highly toxic to the ground inhabiting natural enemies however it was found on par with IPM (93.2/trap) and neem (101.4/trap). HaNPV (124.9/trap) and bird perches (134.3/trap) did not show any significant effect compared to control on ground dwelling natural enemies.

## Preharvest stage

During this stage also endosufan (36.6/trap) was found toxic against the ground inhabiting natural enemies. All the remaining treatments did not cause any significant effect compared to control in reducing ground inhabiting natural enemies.

## 4.2.5 Effect of the Treatments on Natural Parasitism During rabi 1998-99

### Larval parasitism

The IPM treatments were compared for their role on the natural parasitism by *C.chlorideae* on the larvae of *H.armigera* and the results are presented in Table.21 (Plate.8). At 25 days after sowing i.e., 4 days after first treatment, maximum reduction in parasitisation (27%) by *C.chlorideae* was observed in endosulfan treatment. In the other treatments i.e neem and IPM which received neem, HaNPV and bird perches there was a non significant reduction in percentage parasitisation compared to control. At 58 DAS i.e., 6 days after third

Treatment	Parasitisation by Campoletis chlorideae(%)				
	25DAS(4DAT)	58DAS(6DAT)			
Neem 0.006 % (AZA 3%)	9.0 (17.41) <sup>ab</sup>	5.25 (13.50)			
HaNPV250LE/ha	9.25 (17.61) <sup>ab</sup>	5.50 (13.50)			
Bird perches one/plot	10.50 (18.89)⁰	7.00 (14.33)			
Endosulfan 0.07%	8.00 (16.33)ª	4.50 (12.01)			
IPM	9.00 (17.41) <sup>ab</sup>	5.25 (13.03)			
Control	11.00 (19.34) <sup>ь</sup>	7.25 (15.53)			
S.Ed.	1.04	1.28			
CD	2.21	NS			

## Table 21 : Effect of the treatments on natural larval parasitisation by Campoletis chlorideae Uchida during rabi 1998-99

DAS=Days after sowing

DAT=Days after treatment

(Figures in parenthesis are angular transformed values)

\*Values followed by same letters in each column are statistically not significant

treatment no significant difference between treatments in larval parasitisation was observed.

#### Pupal parasitisation

Apart from larval parasitoid, a larval-pupal parasitoid *Carcelia illota* Curron (Tachinidae:Diptera) was also observed in control plots. However its incidence was very low i.e 2% at 58 DAS sampling.

## 4.2.6 Effect of the Treatments on Natural Parasitism During *rabi* 1999-2000

#### Larval parasitism

The results pertaining to larval parasitisation during *rabi* 1999-2000 are presented in Table.22. At 36 days after sowing i.e., four days after treatment the highest reduction in larval parasitisation by *C.chlorideae* was observed in endosulfan treatment (43%). Neem and IPM plots were on par and stood next with reduced larval parasitisation by *C.chlorideae* (38, 33%, respectively). In the remaining treatments also a reduction in parasitisation was observed but not at significant level.

At 68 days after sowing i.e., 6 days after 3<sup>rd</sup> spraying there was no significant difference among the treatments in parasitisation by *C.chlorideae* compared to control (9.50%).

#### Pupal parasitisation

Larval-pupal parasitoid C.illota was also observed, however its incidence was very low i.e 4%.

## 4.2.6 Mean effect of the treatments on natural parasitism

Cumulative data regarding the effect of different IPM components on natural parasitisation by C.chlorideae are presented in Table 23. The

Treatment	Parasitisation by Campoletis chlorideae(% 36DAS(4DAT) 68DAS(6DAT)			
Neem 1750 ml / ha (Nivaar 1500 ppm)	3.25 (10.34)⁼ <sup>⊎</sup>	8.75 (17.14)		
HaNPV250LE/ha	4.00 (11.46) <sup>abc</sup>	9.00 (17.43)		
Bird perches one/plot	4.75 (12.53) <sup>⊳⊄</sup>	9.75 (18.16)		
Endosulfan 0.07%	3.00 (9.92)ª	8.50 (16.90)		
IPM	3.50 (10.67)ªÞ	.8.50 (16.87)		
Control	5.25 (13.03)⁰	9.50 (17.86)		
S.Ed.	1.04	1.38		
CD	2.21	NS		

Table	22:	Effect of	treatments	onr	natural	larval	parasitisation	by
		Campoletis	chlorideae	Uchida	a during	rabi 1	999-2000	

DAS = Days after sowing.

DAT = Days after treatment.

(Figures in parenthesis are arc sin transformed values)

\*Values followed by same letters are statistically not significant.

Treatment	1 <sup>st</sup> sampling	2 <sup>nd</sup> sampling
Neem	6.13 (14.33) <sup>sb</sup>	7.00 (15.34)
HaNPV250LE/ha	6.63 (14.92) <sup>ab</sup>	7.25 (15.62)
Bird perches one/plot	7.63 (16.03) <sup>⊾</sup>	8.38 (16.82)
Endosulfan 0.07%	້ 5.50 (13.56) <sup>°</sup>	6.50 (14.77)
IPM	6.25 (14.47) <sup>ab</sup>	6.88 (15.20)
Control	8.13 (16.56) <sup>⁵</sup>	8.38 (16.82)
S.Ed.	1.101	1.32
CD .	2.340	NS

Table 23: Mean effect of the treatments on natural parasitisation by Campoletis chlorideae Uchida in chickpea.

(Figures in parenthesis are arc sin transformed values) \*Values followed by same letters in each column are statistically not significant

Treatment	Pod damage(%)	Per cent reduction over control
Neem 0.006% (AZA 3%)	11.98 (20.23)⁵	39.37
HaNPV250LE/ha	12.55 (20.72)⁵	36.49
Bird perches one/plot	14.45 (22.32)°	26.87
Endosulfan 0.07%	11.21 (19.56)⁼ <sup>b</sup>	43.27
IPM	10.38 (18.77)ª	. 47.47
Control	19.76 (26.41)⁴	-
S.Ed.	0.550	
CD .	1.180	

Table 24: Effect of the treatments in reducing pod damage byH. armigera in chickpea during rabi 1998-99.

(Figures in parenthesis are arc sin transformed values)

\*Values followed by same letters in each column are statistically not significant



1998-99 1999-2000



results revealed the highest reduction in natural parasitisation of larvae in plots treated with endosulfan (32%). All the remaining treatments did not show any significant effect compared to control in reducing larval parasitism by *C.chlorideae*. During second sampling there was no significant difference among the treatments in parasitisation by *C.chlorideae* compared to control (8.38).

## 4.3.1 Effect of the Treatments on the Pod Damage by *H. armigera* in Chickpea During *rabi* 1998-99

A persual of the data presented in Table 24 revealed that the maximum percentage of pod damage was observed in control (19.8%) (Plate.10 & Figure 12). Among the treatments IPM was found to be the best by registering the lowest percentage of pod damage (10.4%) with 47.5% reduction over control. Endosulfan treatment was also effective as IPM by registering 11.2% of pod damage with 43.3% reduction over control. Neem spray was found next best and was on par with HaNPV spray with 39.4 and 36.5 per cent reduction in pod damage over control, respectively. Bird perches treatment significantly reduced pod damage (14.5%) compared to control (26.9%).

# 4.3.2 Effect of the Treatments on the Pod Damage by *H. armigera* in Chickpea During *rabi* 1999-2000

The results (Table 25 & Figure 12) from a field trial indicated that endosulfan was found to be the best treatment by registering the lowest percentage of pod damage (20.5) the reduction being 49.7% over control but was on par with IPM treatment with 22.6 per cent pod damage (44.4% reduction over control). The treatments with neem and HaNPV were on par in reducing pod damage by registering 26.4 per cent pod damage. Bird perches

Treatment	Pod damage(%)	Per cent reduction over control
Neem 1750 ml/ha (Nivaar 1500 ppm)	26.44 (30.88) <sup>bc</sup>	35.08
HaNPV250LE/ha	26.39 (30.84) <sup>₀</sup>	35.21
Bird perches one/plot	28.08 (31.98)⁰	31.06
Endosulfan 0.07%	20.50 (26.63)ª	49.67
IPM	22.60 (28.36)ª⁵	44.39
Control	40.73 (39.64)⁴	-
S.Ed.	1.718	
CD	3.610	

## Table 25 : Effect of the treatments in reducing pod damage by *H.armigera* in chickpea during rabi 1999-2000

(Figures in parenthesis are arc sin transformed values)

\*Values followed by same letters are statistically non significant

Treatment control	Pod damage(%)	Per cent reduction over
Neem	19.21 (25.99) <sup>∞</sup>	36.5
HaNPV250LE/ha	19.47 (26.18) <sup>bc</sup>	35.6
Bird perches one/plot	21.27 (27.46) <sup>°</sup>	29.7
Enclosulfan 0.07%	15.86 (23.46) <sup>°</sup>	47.6
IPM	16.49 (23.95) <sup>≇b</sup>	45.5
Control	30.25 (33.36) <sup>4</sup>	-
Ş.Ed.	1.137	
CD	2.39	

# Table 26: Mean effect of the treatments in reducing pod damage by H. armigera in chickpea

(Figures in parenthesis are arc sin transformed values)

\*Values followed by same letters are statistically not significant

were efficient in attracting many insectivorous birds, which can be viewed from the reduction in pod damage by 31.1% over control. The control plot recorded the highest pod damage of 40.7 per cent.

## 4.3.3 Mean effect of the Treatments on the Pod Damage by *H. armigera* in Chickpea

The mean data of the two years regarding pod damage are presented in Table 26. From the data it was clear that endosulfan was the best treatment by registering the lowest per cent of pod damage (15.9), the reduction being 47.6 per cent over control but was on par with IPM with 16.5 per cent pod damage (45.5 % reduction over control). The treatments neem, HaNPV and bird perches were on par in reducing pod damage with 19.2, 19.5 and 21.3 per cent pod damage, respectively. The control plot recorded the highest pod damage of 30.3%.

### 4.4.1 Effect of the Treatments on Chickpea Yield During rabi 1998-99

To assess the efficacy of different IPM components on the grain yield of chickpea, studies were conducted and the results are elucidated in Table.27 & Figure.13. The results revealed that IPM was significantly the best treatment by recording the highest yield, 1167 kg/ha, which was 57.3 per cent higher over control (741.8 kg/ha), followed by endosulfan spray 1054 kg/ha which recorded 42 per cent yield increase over control. HaNPV (963.8 kg/ha) and neem (961.8 kg/ha) stood next, being on par and significantly effective by recording 29.7 and 29.9 per cent yield increase over control, respectively. Even though the plots with bird perches recorded significantly less yield (858 kg/ha) than the remaining treatments, it was also found to be significantly effective by registering 15.7 per cent yield increase over control.

Treatment	Grain yield (kg/ha)	Per cent increase over control
Neem 0.006% (AZA 3%)	961.8 °	29.7
HaNPV250LE/ha	963.9 °	29.9
Bird perches one/plot	858 4	15.7
Endosulfan 0.07%	1054 <sup>b</sup>	42.
IPM	1167 *	57.3
Control	741.8 <sup>°</sup>	-
S.Ed.	34.15	
CD	72.77	

Table 27 : Effect of the treatments on the grain yield of chickpea duringrabi1998-99

Values followed by same letters are statistically not significant.

•



Treatment	Grain yield (kg/ha)	Per cent increas over control
Neem 1750 ml/ha	1298 <sup>b</sup>	51.9
HaNPV250LE/ha	1317 <sup>b</sup>	54.2
Bird perches one/plot	1096 <sup>°</sup>	28.3
Endosulfan 0.07%	1392 "	63.1
.IPM	1361 <sup>ab</sup>	59.4
Control	854 <sup>d</sup>	•
S.Ed.	34.5	
CD	72.4	

Table 28 : Effect of the treatments on the grain yield of chickpea duringrabi1999-2000

Values followed by same letters are statistically not significant.

# 4.4.2 Effect of the Treatments on Chickpea Yield During *rabi* 1999-2000

From the results presented in Table.28 & Figure.13 endosulfan was adjudged as the superior among the treatments by recording the highest yield of 1392 kg/ha which was 63 per cent increase over control. Due to integration of the pest management strategies, IPM was on par with endosulfan spray with 1361.7 kg/ha an increase of 59.4 per cent over control. The treatment HaNPV (1317 kg/ha) and neem (1298 kg/ha) were also found significantly effective in increasing the yield upto 54 and 52 per cent increase over control, respectively. Plots with bird perches also recorded 28 per cent increase in the grain yield over control mainly because of immense bird activity at ICRISAT location.

## 4.4.3 Mean Effect of the Treatments on Chickpea Yield

Mean data of the two years are presented in Table 29. The results revealed that IPM was found to be the best treatment, by recording the highest grain yield 1264.4 kg/ha, which was 58.5 per cent increase over control (797.9 kg/ha) but it was on par with plots treated with endosulfan spray (1223 kg/ha) with 53.3 per cent increase over control. HaNPV (1140.4 kg/ha) and neem (1129.9 kg/ha) were on par and recorded significantly higher yields with 42.9 and 41.6 per cent increase over control, respectively. Even though bird perches (977 kg/ha) was inferior than the other treatments it was also found to be significantly effective by registering 22.5 per cent increase over control.

## 4.5.1 The economics of the Treatments During rabi 1998-99

The cost-benefit ratio's which were worked out for different treatments (Table.30) showed a higher cost benefit ratio (1:3.32) with bird perches plot but the overall yield increase over control was very less. IPM due to the integration of different management strategies showed the highest cost

Treatment	Grain yield (kg/ha)	Per cent increase over control
Neem	1129.9 *	41.6
HaNPV250LE/ha	1140.4 <sup>b</sup>	42.9
Bird porches one/plot	977 <sup>°</sup>	22.5
Endosulfan 0.07%	1223 "	53.3
IPM	1264.4 "	58.5
Control	797.9 <sup>d</sup>	-
S.Ed.	34.26	
CD	72.02	

## Table 29: Mean effect of the treatments on the grain yield of chickpea.

\*Values followed by same letters are statistically not significant

	Grain	Yield (kg/ha)	Gross	Insecticidal &	Not	C:P (Cost
Treatment	Gross	Additional yield over control	income (Rs.)	application cost (Rs.)	income (Rs.)	Benefit ratio)
Neem 0.006% (AZA 3%)	961.8	220.0	9,618	1,750	7,868	<b>1</b> :1.26
HaNPV 250LE/ha	963.9	222.0	9,638	2,000	7,638	1:1.10
Bird perches one/plot	858.0	116.2	8,580	350	8,230	1:3.32
Endosulfan 0.07%	1054.1	312.3	10, <b>541</b>	1,725	8,816	1:1.81
IPM	1167.0	425.2	11,670	1,845	9,825	1:2.30
Control	741.8	-	7,418	-	7,418	-

Tabble 30: Cost Benefit ratio of IPM components in chickpea during rabi 1998-99

Cost of each spray/ha

Neem = Rs 350/-HaNPV = Rs 400/-Bird perches= Rs 350/ha Endosulfan = Rs 345/-Cost of chickpea = Rs10.0 / kg

Tabble 31: Cost Be	nefit ratio	of IPM c	components in chick	pea during <i>rabi</i>	1999-2000		
		Grain	Yield (kg/ha)	- Second	Incontinidal 8	4CM	
Treatment	<b>σ</b>	ross	Additional yield over control	income (Rs.)	application cost (Rs.)	income (Rs.)	U.B. (Cost Benefit ratio)
Neem 1750 m	il/ha 1;	298.0	444.0	19,470	2,297	17,173	1:2.90
(Nivaar 1500 ppm)							
HaNPV 250LE/ha	4	317.1	463.1	19,756	2,645	17,111	1:2.63
Bird perches one/plo	t 1(	0.96.0	242.0	16,440	350	16,090	1:10.37
Endosulfan 0.07%		392.0	534.0	,20,894	2,159	18,735	1:3.71
Mdi	1	361.7	507.0	20,426	2,025	17,990	1:3.76
Control		854.0	ı	12,810,	1	12,810	·
Cost of	each spray	y/ha					
Neem	= Rs	459/-					
HaNPV	= Rs	529/-					

Bird perches= Rs 350/ha

Endosulfan = Rs 432/-

Cost of chickpea = Rs15.0/kg

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	Grain	Yield (kg/ha)	Gross	Insecticidal &	Net	C:B (Cost
Treatment	Gross	Additional yield over control	income (Rs.)	application cost (Rs.)	income (Rs.)	Benefit ratio)
Neem	1129.9	332.0	14,544	2,023.5	12,520.5	1:2.05
HaNPV 250LE/ha	1140.45	342.55	14,697	2,322.5	12,374.5	1:1.84
Bird nerches one/nlot	977.00	179.1	12,510	350.0	12,160.0	1:6.40
Plid percense on propose	1223.05	425.15	15,717.5	1,942.0	13,775.5	1:2.74
Endosuitari 0.07 %	1264.35	466.45	16,048	1,935.0	13,907.5	1:3.01
Md	6.797.9	ı	10,114	•	10,114.0	•
Control						
Cost of eac	ch spray/ha					

Neem = Rs 405/-

HaNPV = Rs 465/-

Bird perches = Rs 350/ha Endosulfan = Rs 388/-

Cost of chickpea = Rs12.5/kg

benefit ratio (1:2.30) among the remaining treatments with considerable increase in yield over control. Endosulfan used in the experiment was comparatively cheaper chemical and proved to be cost effective with a cost benefit ratio of 1:1.81. Neem and HaNPV were next best treatments with 1:1.26 and 1:1.10 C:B ratio's, respectively.

## 4.5.2 The Economics of the Treatments During rabi 1999-2000

During *rabi* 1999-2000 the results (Table.31) revealed that bird perches treatment was most economical with 1:10.37 cost-benefit ratio. However the overall yield and the yield increase over control was much less. Among the remaining treatments IPM was the most economical treatment which registered the highest cost-benefit ratio of 1:3.76 followed by endosulfan treatment (1:3.71). Neem spray stood next in the order of efficacy with the cost-benefit ratio of 1:2.90 and the lowest cost-benefit ratio was obtained with HaNPV spray (1:2.60).

## 4.5.3 The Economics of the Treatments (mean of two years)

The cost-benefit ratios were worked out to know the economics of different IPM components individually and in combination. The results (Table 31) revealed that although bird perches gave the highest cost-benefit ratio (1:6.40) the overall yield and the yield increase over control was much less. Among the remaining treatments highest cost-benefit ratio (1:3.01) was obtained by integration of all IPM components. Endosulfan proved to be the effective and cheaper chemical with 1:2.74 cost-benefit ratio. Neem spray with 1:2.05 ratio proved economical compared to HaNPV (1:1.84).

## 4.6 Residues of Endosulfan in Chickpea Seed and Husk

The presence of residues were tested only in the sample of rabi season 1999-2000. The results (Table 33) suggested presence of residues

Sample	Alpha Endosulfan (mg/kg)	Beta Endosulfan (mg/kg)	Endosulfan sulphate (mg/kg)	Total endosulfan (mg/kg)
Endosulfan treatment				
Seed	0.322	0.461	0.022	0.805
Husk	0.460	0.475	0.415	1.350
IPM treatme	ent			
Seed	BDL	BDL	BDL	BDL
Husk	BDL	BDL	BDL	BDL
MRL				2.00

## Table 33 : Endosulfan residues in chickpea seed and husk

BDL-Below detectable limit

MRL-Maximum residual limit acceptable as per FAO standards.

in the samples collected from endosulfan treatment in seeds as well as in husk also. The seeds of chemical plot contained 0.81mg/kg endosulfan residues where as husk contained 1.35mg/kg endosulfan residues but both are lower than the maximum residual limit prescribed by FAO (2mg/kg). The samples collected from IPM plot showed residues below detectable limit both in seed and husk.

## 4.7 LABORATORY STUDIES

## 4.7.1 Effect of Neem as Oviposition Deterrent Against H.armigera Adults

To determine the oviposition deterrent effect of neem 30 days old chickpea plants in pots were taken and half of them were sprayed with neem and the remaining half with water. Fifteen pairs of *H.armigera* adults (12 hrs old) were released, and the egg number laid on chickpea plants was counted till the death of adults. The results (Table 34) showed significantly lower number of eggs on neem treated plants i.e., 111.60 compared to 287.80 eggs in control, which clearly revealed the oviposition deterrent effect of neem on chickpea against *H.armigera* adults.

## 4.7.2 Effect of neem as Antifeedant Against *H.armigera* In Choice Test (Table 35)

Small size larvae (first & second instars) were observed for their orientation towards neem treated and untreated chickpea leaves. The data showed 46% of larvae settled on neem treated leaves and 47% on untreated control, suggesting no significant difference between the treatments.

In medium larvae (third & fourth instars) 36% settled on neem treated leaves and pods and 54% on untreated chickpea leaves and pods. Where as 22% of large larvae (fifth & sixth instars) were settled on neem treated and 78% on untreated chickpea leaves and pods.
Treatment	Mean number of eggs laid/pot	S.Em.	
Neem 0.006%	111.60	6.63	
Control(water spray)	287.80	4.92	

Means are significantly different by the two sample t-test.

 Table 34
 Effect of neem on H.armigera oviposition

Based on the above experiment it was concluded that with increase in larval age, their differentiation between neem treated and untreated leaves increased.

#### In no choice test

Different age groups of larvae were observed for their preference towards neem treated chickpea leaves and pods in no choice situations. When larvae were given food mixed with neem under no choice situation, the initial behavior was evident in older larvae by moving around the food perhaps in search of better option. Since there was no option ultimately the larvae settled on the neem impregnated diet. It was clear from the study that the response of the larvae to neem increases as the larvae advance in age. 40% of the large larvae (fifth & sixth instar), 20% of the medium (third & fourth instars) and none of the small size larval group (first & second instars) showed initial avoidance of neem.

#### 4.7.3 Effect of neem on different age groups of H.armigera

Chickpea leaves sprayed with neem 0.006% were given to larvae of different age groups i.e. small (first & second instars), medium (third & fourth instars) and large (fifth & sixth instars) to assess the effect and the data are presented in Table 36.

#### Larval mortality

Neem treatment gave 47.9% larval mortality of small larvae as against 9.1% in control. While in medium larval group only 15.8% larval mortality was observed in neem treatment as against 5.3% mortality in control. Where as large larvae experienced no mortality.

# Table 35 : Neem as antifeedant in choice test against different age groups of *H.armigera*

#### I &II instars

Treatment	Mean number of larvae Settled	S.Em.	
Neem	4.6	0.617	
Control	4.7	0.539	

Means are statistically not significant by two sample t-test.

#### III & IV instars

Treatment	Mean number of larvae Settled	S.Em.
Neem	3.6	0.600
Control	5.4	0.539

Means are statistically significant by two sample t-test.

### V & VI instars

Treatment	Mean number of larvae Settled	S.Em.	
Neem	2.2	0.326	
Control	7.8	0.327	

Means are statistically significant by two sample t-test.

#### Larval duration

There was significant increase of mean larval duration in all the age groups when they received neem treatment. Small larval group took 14.9 days when they provided with neem treated food as against 12.9 days in control to complete larval stage. Medium size larvae took 9.4 days as against 8.3 days in control, where as large size larvae took 5.1 days with neem as against 3.8 days in control treatment to complete the larval stage.

#### Pupal weight

Small (285.6 mg) and medium (310 mg) age groups showed significant decrease in pupal weight compared to control (372.4, 370.3 mg resp.) when they were provided neem treated leaves as food compared to control, where as large size larvae (339.0 mg) showed non significant difference in pupal weight compared to control (357.7mg).

#### Pupal period

The small size larval group showed significant increase in pupal period (13.9 days) compared to control (10.3 days). Medium size larval group also showed significant increase in pupal period (12.0 days) compared to control (10.1 days) when they received neem treated leaves as food. But large size larval group showed non significant increase in pupal period (11.53 days) compared to control (11.48 days).

#### Oviposition period

There was a significant decrease in oviposition period of adults emerged from all the three age groups compared to control when fed with neem treated leaves during the larval stage. In the adults obtained from small larval group fed with neem treated leaves the oviposition period was 3.7 days as against 6.5 days in control, where as in medium group 4.5

	Neem	Control	Significance
Larval mortality(%)			
1&11	47.89	9.06	sig
111&IV	15.75	5.29	sig
V&VI	-	-	-
Larval duration(Days)			
181	14.85	12.9	sig
111&IV	9.43	8.3	sig
V&VI	5.12	3.84	sig
Rupplusicht (mg)			
Pupar weight (mg)	285.6	372 4	sia
	310.1	370.3	sig
V&VI	339	357.7	NS
Pupal period(Days)			
1&11	13.88	10.25	sig
111&IV	12.02	10.13	sig
V&VI	11.53	11.48	NS
Oviposition period(Da	ays)	<del>-</del>	
1811	3.72	6.47	sig
	4.51	6.93	sig
V&VI	4.7	6.03	sig
Fecundity(Number)			
1&11	838.7	1697	sig
III&IV	937.5	1779	sig
V&VI	1049.7	1679	sig
Egg hatchability (%)			
1&11	89.75	91.02	NS
111&IV	89.98	87.29	NS
V&VI	93.14	94.5	NS

Table 36 : Effect of neem on different stages of H.armigera

days as against 6.9 days in control. While in large size larval group it was 4.7 days compared to 6.0 days in control.

#### Fecundity

Significant decrease in number of eggs laid by the resultant females of all the age groups of larvae fed with neem treated food compared to control was observed. In small larvae group 838.7 eggs per moth were recorded as against 1697 eggs per moth in control. In medium larvae group 937.5 eggs per moth compared to 1779 eggs per moth in control. Where as in large larvae group 1049.7 eggs per moth compared to 1679 eggs per moth in control was observed.

#### Egg hatching

No significant difference was observed in egg hatching between neem treated and control in all the three age groups.

### 4.7.4 Efficacy of Robin Blue in Increasing the Persistence of HaNPV on Chickpea Foliage

To assess the efficacy of robin blue (1%) in increasing the persistance of HaNPV as UV rays protectant, an experiment was conducted by using 3<sup>rd</sup> instar larvae of *H. armigera* and the results are presented in Table 37 & Figure 14.

The data revealed that HaNPV and HaNPV+ robin blue (1%) treatments were on par with 75% and 76.2% mortality, respectively when fed with the leaves immediately after treatment, the corresponding mortality in control was 4.8%. When the treated leaves were fed one day after spray with HaNPV+ robin blue (1%) recorded significantly higher mortality i.e 51.2% as against 34.5% with HaNPV alone. Control registered no mortality. When

Days after Treatment	HaNPV alone (L	HaNPV+ Robin blue 1% arval mortality (%)	Control ))	S.Ed.	CD
0	75 (60.5) <sup>°</sup>	76.19 (61.7) <sup>*</sup>	4.76 (10.0) <sup>⊾</sup>	2.43	5.29
1 <sup>si</sup>	34.52 (35.9) <sup>ຫ</sup>	57.19 (45.7) <sup>°</sup>	0 (0.2238) <sup>°</sup>	1.26	2.7
2 <sup>nd</sup>	29.76 (33.0) <sup>ະ</sup>	45.24 (42.5) <sup>°</sup>	3.57 (5.8) <sup>°</sup>	1.77	3.85
3rd	22.62 (28.3) <sup>⊾</sup>	ັ 39.29 (38.66) <sup>*</sup>	5.67 (13.8) <sup>°</sup>	1.15	2.5
4 <sup>th</sup>	7.14 (13.0) <sup>b</sup>	27.52 (35.9) <sup>°</sup>	4.98 (12.9) <sup>b</sup>	1.57	3.42
5 <sup>m</sup>	4.76 (8.2) <sup>6</sup>	19.76 (32.9) <sup>°</sup>	1.19 (4.4) <sup>ະ</sup>	2.7	4.5
6"	2.58 (4.8) <sup>b</sup>	8.53 (14.1) <sup>°</sup>	2.38 (4.8) <sup>⁵</sup>	2.14	4.67

 Table 37: Persistence of HaNPV on chickpea foliage with / without

 Robin blue against 3rd instar larvae of H. armigera

(Figures in paranthesis are arc sin transformed values)

\*Values followed by same letters in each row are statistically not significant



the larvae were fed with the treated leaves after 48 hours once again HaNPV+ robin blue (1%) gave significantly high mortality (45.2%) compared to HaNPV (29.8%) and control 3.6%. Similarly HaNPV+ robin blue (1%) treatment showed significantly high rate of mortality (39.3%) compared to HaNPV (22.6%) and control (5.7%) when larvae were fed with the leaves 3 days after treatment. When the larvae were fed with the leaves 4 days after treatment again HaNPV+robin blue (1%) recorded significantly high mortality (27.5%) while HaNPV treatment (7.1%) was not significantly different from control (5%). Even 5 days after treatment HaNPV+ robin blue (1%) treated leaves when fed gave 19.8% mortality of the larvae and thus proved to be a good UV protectant by retaining HaNPV persistence, while HaNPV (4.8%) showed non significant mortality compared to control (1.2%). Robin blue continued to show UV protectant ability by causing 8.5% mortality when combined with HaNPV compared to 2.6% with HNPV alone and 2.4% on control when the larvae were fed with the treated leaves after six days.

#### 4.7.5 Effect of HaNPV on Three Different Age Groups of H.armigera

Effect of HaNPV on different age groups of *H.armigera* i.e., small (first & second instars), medium (third & fourth instars) and large (fifth & sixth instars) larvae in terms of larval mortality, pupal abnormality and death, pupal weight and fecundity were studied and the results are presented in Table 37.

#### Larval mortality

HaNPV treatment @ 6 x10° POB / I on small larvae (first & second instars) gave complete mortality as against 2.7% mortality in control. Medium larvae (third & fourth instars) registered 76.8% mortality as against 3.5% mortality in control. Large larvae (fifth & sixth instars) recorded 59.8% mortality as against no mortality in control.

	HaNPV	Control	Significance
Larval montality(%)			
1&11	100	2.68	Sig
111&IV	76.75	3.5	Sig
V&VI	59.80	-	Sig
Pupal abnormality(%	))		
1811	-	-	
III&IV	97.95	-	Sig
V&VI	86.90	-	Sig
Pupal weight (mg)			
1811	-	-	
III&IV	290.47	375.67	Sig
V&VI	349.43	357.93	NS
Fecundity (number)			
1811	•	-	
III&IV	397	1079	Sig
V&VI	689	865	Sig

# Table 38: Effect of HaNPV on different stages of H. armigera

#### Pupal abnormality and death

Pupal abnormality and death was not recorded in small size larvae as there was 100% mortality at larval stage itself. Medium larvae group recorded 98.0 % pupal abnormality and death where as large larvae group recorded 86.9% pupal abnormality and death.

#### Pupal weight

Small larvae did not reach pupal stage but in medium group there was significant reduction in pupal weight (290.5 mg) of the resultant pupae from the treated larvae as against control (375.7 mg). Where as in large larval group there was non significant difference between HaNPV treated and normal pupal weight (349.4 & 357.9 mg, respectively).

#### Fecundity

In the adults developed from the HaNPV treated medium larval group there was a significant reduction in number of eggs (397 per female) due to early death of male and female moths compared to control (1079 eggs per female). Similar trend was observed in the adults developed from the HaNPV treated large larval group with significantly lower number of eggs per female (689) compared to control (865 eggs per female).

# 4.7.6 Effect of HaNPV treatment at fifth instar larval stage on fecundity of *H.armigera*

A significant reduction in number of eggs laid per female was observed in all the treatments that included either male or female or both that emerged from HaNPV fed larvae at fifth instar stage compared to control (Table 39 & Figure.15). There was only 79.6% ovipositional potential realised in treated larvae in comparison with control where both male and female moths were from HaNPV infected larval population, where

Treatment	Number of eggs/ female	Reduction against control(%)	Hatchability(%)	Reduction against control(%)
HaNPVfemale + HaNPV male	689 (25.98) <sup>°</sup>	20.40	62.62 (52.80) <sup>*</sup>	30.47
HaNPV female + healthy male	718 (26.59) <sup>°</sup>	17	65.20 (54.00) <sup>°</sup>	27.60
Healthy female + HaNPVmale	780 (27.88) <sup>°b</sup>	9.83	83.73 (67.50)⁵	7.03
Healthy female + healthy male	865 (29.39) <sup>⁵</sup>	-	90.06 (73.90) <sup>°</sup>	-
S.Ed.	1.202		2.87	
CD	2.619		6.25	

Table 39: Effect of HaNPV treatment on fecundity and egg hatchability of *H. armigera* when received at 5th instar stage.

\*Values followed by same letters in each column are statistically not significant





as 83% of egg laying was observed in treatment where female from HaNPV infected population and male from normal population were allowed to mate. 90.2% egg laying compared to control was observed in the treatment that included female from normal population and male from HaNPV infected population. Thus both male and female adults that emerged from HaNPV fed larvae showed profound effect on the total fecundity.

# 4.7.7 Effect of HaNPV Treatment at Fifth Instar Larval Stage on Hatching of eggs of *H.armigera* (Table 39 & Figure 15)

When the fifth instar larvae of *H.armigera* were treated with HaNPV @ 6x10<sup>9</sup> POB/I, a significant reduction in hatching of the eggs laid by moths of resultant population was observed compared to control. When both male and female adults were collected from HaNPV treated population at fifth instar larval stage only 62.6% egg hatchability was observed i.e 30.5% reduction compared to control. Eggs from female collected from HaNPV treated population at fifth instar larval stage and male collected from normal population had 65.2% hatching i.e 27.60% reduction compared to control. When male was collected from HaNPV treated population at fifth instar larval stage and female from normal population the resultant eggs recorded 83.7% hatching i.e only 7.0% reduction compared to control. In the control where male and female adults were from normal population 90.1% hatchability of eggs was observed.

#### DISCUSSION

Helicoverpa armigera Hubner, a major pest on chickpea, has assumed major status because of its high fecundity, multiple generations, high generation turn over, polyphagy and migratory behavior. Although it attacks chickpea throughout the crop growth, the damage caused during flowering and pod formation stages results in substantial losses. To combat this pest till now the thrust was given mainly on chemicals, however their indiscriminate use resulted in the development of pest resistance, resurgence, environmental pollution, besides having adverse effects on bioagents. This ultimately led to adopt an appropriate IPM programme. Present studies were carried out in field and laboratory during rabi 1998-99 & 1999-2000 at ICRISAT Center, Patancheru, A.P. with a view to develop suitable and sustainable IPM strategies against H.armigera on chickpea by considering their safety against natural resources. The results of the experiments conducted are discussed in this chapter with the available literature.

#### 5.1 FIELD STUDIES

#### 5.1.1 Population Fluctuations of H.armigera in Chickpea

A sound knowledge on the population fluctuations of the chickpea pod borer helps to evolve suitable pest management strategies.

During rabi 1998-99 the peak moth activity was observed during initiation of flowering stage which was one week before the peak oviposition. During rabi 1998-99 the maximum number of eggs (Figure.16) and small size (Figure.17) larvae were observed during flowering stage of the crop with peaks at 50 & 57 DAS, respectively, which did not influence the yield directly. Medium (Figure.18) and large (Figure 19)











Plate 9 : Larva of *H. armigera* on the chickpea foliage.



Plate 10. Pod damage due to H. armigera.

size larval populations were more at pod formation and preharvest stages, respectively, but the peak population was observed stage at 71 & 85 DAS respectively, which directly influenced the yield by enhanced pod damage (Plate.9&10). Even though the egg and small larval population were high during flowering stage and medium and large larval population during pod formation stage, there was more or less uniform population throughout the crop growth suggesting the immigration of pest population from other fields.

The larvae were observed throughout the cropping period from 15 DAS to 99 DAS, even though it attained three peaks at 29, 57, 85 DAS the highest was at 57 DAS i.e first week of January which coincided with flowering & pod initiation stage. The pest activity started in the second fortnight of November and continued till the end of February (Figure 20).

During *rabi* 1999-2000 also the population fluctuation trend of *H.armigera* followed more or less the same as that of *rabi* 1998-99 The maximum moth activity was observed at initiation of flowering stage i.e one week before the maximum egg laying and remaining two peaks at 90 and 97 DAS, respectively. The maximum number of egg (Figure.21) and small larvae (Figure.22) of *H.armigera* were observed during flowering stage of the crop with peak at 54 days after sowing. Medium (Figure.23) and large (Figure. 24) size larval populations were more at pod formation stage, with peak population at 83 and 90 DAS, respectively, which directly caused the economic damage to the crop.

The larvae were observed in the field from 24 DAS to 112 DAS, even though it attained four peaks at 54, 68, 75 and 90 DAS the highest at 68 DAS i.e last week of December which coincided with pod formation











stage. The pest activity started in the second fortnight of November and continued till harvest i.e first week of February (Figure.25).

Mahajan *et al.* (1990) observed tha maximum pheromone catch during 3<sup>rd</sup> and 5<sup>th</sup> meterological week, but in the present study during *rabi* 1998-99 it was observed at 51<sup>st</sup> meterological week and during *rabi* 1999-2000 at 3<sup>rd</sup> meterological week.

Thakur (1990) observed the infestation of *H.armigera* on chickpea from 3<sup>rd</sup> week of October and first week of November upto the middle of March and also recorded the highest population in second week of December and the 2<sup>nd</sup> peak in first and 3<sup>rd</sup> week of January. Yadava and Lal (1988) reported two peaks in the *H.armigera* population in chickpea during the 47<sup>th</sup> to 50<sup>th</sup> and 11<sup>th</sup> to 15<sup>th</sup> standard weeks. The finding of the present studies were in conformity with the above reports.

On the overall basis the pest load was comparatively low during *rabi* 1999-2000 season compared to *rabi* 1998-99 which could be due to early sowing of the crop. The alteration of the sowing date itself is one of the IPM components. Chaudhary and Sachan (1995) conducted experiments during *rabi* 1990-91 and 1991-92 in chickpea and stated that October sown crop has the lowest population of *H.armigera* (0.42-0.5 larva/m row) and the greatest grain yield where as November sown crop had the greater number of insects and lowest yield irrespective of insecticidal use. According to Prasad and Singh (1997) chickpea crop sown on 25<sup>th</sup> September produced more yield, and recorded lower incidence of *H.armigera* compared to the sowing on 10<sup>th</sup> October. Both the above reports support the present finding.



















# 5 1.2 Efficacy of the treatments on the ovipositional preference of *Helicovepa armigera*.

During rabi 1998-99, at all the stages of crop growth period neem was the best effective treatment in reducing the oviposition of *H.armigera* on chickpea. Neem treatment gave 47, 33 and 30 per cent reduction of egg laying in comparision with control during vegetative, flowering and podding stages of the crop and complete reduction of egg laying during preharvest stage of the crop. IPM was the next best treatment which received neem as first and fourth spray with 29 to 86 per cent reduction of egg laying compared to control. Endosulfan also showed its efficacy in reducing egg laying by *H.armigera* moths during vegetative, podding and preharvest stages but not at flowering stage, particularly endosulfan showed its effect till few days after the treatment. Since IPM is a combination of neem, HaNPV, bird perches and endosulfan spray, it was found to be equally effective as that of neem as ovipositional deterrent.

During *rabi* 1999-2000 neem proved as the best ovipositional deterrent against *H.armigera*, and provided significant protection to chickpea crop from pod borer throughout the crop period, with lowest mean number of eggs in all the stages of the crop i.e 86, 67, 87, 100 per cent reduction of egg laying compared to control during vegetative, flowering, podding and preharvest stages of the crop, respectively. IPM which received neem as first and fourth spray also effectively reduced the percentage egg laying compared to control (32 to 100%). Endosulfan also showed some effect in reducing oviposition by *H.armigera* moths particularly immediately after spraying.

The ovipositional deterrent effect of azadirachtin was confirmed by Warthen (1979); Redfern et al. (1981); Rembold (1984) and Schmutterer

(1990); Murugan *et al.* (1995). Jeyakumar and Gupta (1999) reported that the neem seed kernel extract 10 & 7.5% treatment reduced the oviposition to 60.9 and 59 per cent by *H.armigera* compared to control. Ramachandra Rao *et al.* (1990) also reported the ovipositional repellent effect of neem products. The report by Rosaiah (1992) on the maximum oviposition repellency of Repellin, a neem product, to *H.armigera* on cotton also strengthen the present observation on the oviposition repellency of neem to *H.armigera* on chickpea.

### 5.1.3 Efficacy of the Treatments in Managing Small Size Larval Population of *H.armigera*.

During rabi 1998-99 endosulfan as a chemical was the most effective treatment in managing small size larvae (first & second instars) during vegetative stage with 38% reduction compared to control. But in the remaining stages of the crop i.e flowering, pod formation and preharvest stages IPM was the superior treatment with 31, 22, 79 per cent reduction compared to control, respectively. During vegetative stage IPM was on par with neem because in the first spraying it received neem. HaNPV and neem showed equall efficacy in managing small size larvae with a range of 14 to 26 per cent reduction compared to control. Bird perches showed no significant reduction of small size larvae compared to control during flowering, pod formation and preharvest stages.

During rabi 1999-2000 also endosulfan was the most effective treatment in managing small size larvae during vegetative stage with 50% reduction compared to control. Whereas neem was superior during flowering with 40.7% reduction of the larvae compared to control. This may be due to good oviposition deterrent effect of neem, which ultimately lead to less infestation. IPM was superior treatment during pod formation

and preharvest stages in keeping small size larvae at low level with 40 and 51% reduction compared to control, respectively. HaNPV was equally effective as that of endosulfan (7 to 44% reduction compared to control) in managing small size larvae. Bird perches showed no effect in managing small size larvae of *H.armigera*.

IPM initially did not show its superiority but later when it continued to receive different IPM components one after another, its superiority was observed.

Reports regarding the effect of different IPM components on small size larvae of *H.armigera* are not available.

### 5.1.4 Efficacy of the Treatments in Managing Medium Size Larval Population of *H.armigera*.

During *rabi* 1998-99 initially during vegetative stage due to its antifeedant and growth inhibiting effects, neem showed its superiority in managing medium size larvae of *H.armigera* with 40 per cent reduction compared to control followed by HaNPV with 37.5 per cent reduction compared to control. But during the remaining stage of crop, IPM treatment held its superiority in keeping medium size larvae at low level with 34.7, 32.6 and 41.5 per cent reduction over control during flowering, pod formation and preharvest stages, respectively. HaNPV showed superior effect than even endosulfan with 17 to 38 per cent reduction compared to control. HaNPV showed less effect at podding stage which may be due to presence of majority of medium size larvae in the pods and less access to potential POBs. Neem showed more effect at vegetative and flowering stages compared to remaining stages. Erecting bird perches was found to be as effective as endosulfan with 19 to 28 per cent reduction over control in different crop stages. During *rabi* 1999-2000 IPM and HaNPV treatments proved superior in managing medium size larvae with 45 to 72% reduction compared to control. Even though endosulfan given high initial kill, on the overall basis, neem showed superior effect compared to endosulfan with 42 to 60% reduction over control. Whereas endosulfan gave 29 to 64% reduction over control. Installation of bird perches was found useful by reducing 8 to 32% larval population compared to control.

Parasharya (1995) reported that birds prefer medium and large size larvae and assist in the spread of insect pathogens by eating NPV infected larvae, this report support the significant effect of bird perches in reducing medium size larvae compared to control in both the seasons. Reports regarding the effect of remaining IPM components on medium size larvae of *H.armigera* are not available.

# 5.1.5 Efficacy of the Treatments in Managing Large Size Larval Population of *H.armigera*

During *rabi* 1998-99 IPM maintained its supremacy in managing the fifth and sixth instar larvae of *H.armigera* by registering 47, 70, 59 and 84 per cent reduction over control in vegetative, flowering, podding and preharvest stages, respectively, followed by endosulfan with 47 to 84 per cent reduction compared to control. Neem and HaNPV showed almost similar efficiency by recording 39 to 87 percent reduction over control except at pod formation stage where HaNPV showed less efficiency. Erecting bird perches also reduced the larval number significantly compared to control and it was as effective as neem and HaNPV in managing large size larval population of *H.armigera*.

During *rabi* 1999-2000 during vegetative stage HaNPV was superior in managing large size larvae, but later in the remaining stages of crop growth IPM maintained its superiority with 35 to 90% reduction of larval population compared to control followed by HaNPV which also produced similar effect with that of IPM. At vegetative stage IPM was on par with neem because it received neem as first spray. Endosulfan and neem gave more or less uniform control with 24 to 85% reduction over control during various crop stages. Bird perches also proved effective and gave 28 to 66% reduction of large size larval population compared to control.

Parasharya (1995) reported that birds prefer medium and large size larvae and assist in the spread of insect pathogens by eating NPV infected larvae, this report support the significant effect of bird perches in reducing large size larvae compared to control in both the seasons. Reports regarding the effect of remaining IPM components on large size larvae of *H.armigera* are not available.

# 5.1.6 Efficacy of the Treatments in Managing *H.armigera* Total Larval Load

During *rabi* 1998-99 initially at vegetative stage endosulfan showed its superiority in reducing total larval load with 37% reduction compared to control. But in the remaining stages of crop growth IPM stood as a better treatment with 35, 31 and 43 % reduction over control during flowering, podding and preharvest stages, respectively. Based on average larval load in different crop stages the effect of endosulfan almost equaled to  $H_{P}^{A}NPV$  and neem. Bird perches even though were inferior compared to other treatments, contributed 11 to 26 % reduction of larval load compared to control without any investment.

During rabi 1999-2000 at vegetative stage of the crop HaNPV proved more effective with 47% reduction of total larvae compared to control. But in the remaining period of the crop growth IPM plots were found superior with 46, 43 and 62% reduction of larval population compared to control during flowering, pod formation and preharvest stages of the crop, respectively. HaNPV gave more or less equal control as that of IPM during flowering and pod formation stages of the crop. Even though endosulfan was superior in managing total larvae during vegetative and flowering stages, its effect was almost similar to neem during pod formation stage. Even though bird perches showed inferior effect compared to other treatments, it was found significantly superior compared to control and contributed 14 to 37% reduction of total larval load compared to control especially medium and large size larvae which had profound influence on the yield.

Thakur *et al.* (1988) reported that neem kernel extract and leaf extract recorded significantly less larval population in comparison to control, however it was less effective compared to chemicals and concluded that it can be used in place of highly toxic synthetic insecticides. Sehgal and Ujagir (1990) & Datkhile *et al.* (1992) stated that NSKE at 5% was less effective on gram pod borer *H.armigera* when compared to endosulfan but still significantly better than the control. The above reports support the present findings with regard to the superiority of endosulfan over neem in controlling small and medium size larvae, but on the overall basis neem was equally effective as that of endosulfan. This was supported by Sinha (1993) who reported that NSKE 5% gave 40% reduction of infestation of *H.armigera* in chickpea and was comparable to endosulfan 0.07%.

Anitha Mistry *et al.* (1984) reported that five sprays of HNPV @ 250 LE/ha gave satisfactory control of *H.armigera* in chickpea. Jayaraj *et al.* (1987) also found significant control of chickpea pod borer due to HNPV @ 250 LE/ha. The observations of Pawar *et al.* (1987) on the effectiveness of HNPV on chickpea pod borer, compared with endosulfan spray corroborate the present findings.

The findings of Ghode *et al.* (1988) on the high avian predation of *H.armigera* by cattle egrets and river tern in the month of January support the present findings, besides this, ICAR (1992) also reported 33% reduction of *H.armigera* population by birds on wheat and 20-40% only by house sparrow. In this study bird perches reduced only medium and large size larvae. Parasharya (1995) reported that birds prefer medium and large size larvae and assist in the spread of insect pathogens by eating NPV infected larvae. This report supports the present findings of heavy reduction of larval population of *H.armigera* in IPM plots, when HaNPV was sprayed twice apart from bird perches to encourage the predation by birds.

According to Sanap and Pawar (1998) IPM treatment for controlling *H.armigera* in gram includes three spray applications starting from initiation of flowering and subsequent two sprays at 15 days interval with first two sprays either with HNPV @ 250 LE/ha or NSKE 5% followed by a third spray with endosulfan 0.07% were most effective, this report support the present finding of superiority of IPM.

# 5.1.7 Effect of the Treatments on Natural Enemies Present in Chickpea Ecosystem

One of the main reasons for failure of pest control with chemicals is the distruction of natural enemies present in the agro-ecosystem which leads to pest build up in the absence of natural control. So at this juncture it is necessary to incorporate plant protection options into IPM system which are safer to natural enemies of the pest. For this reason the present study evaluated the effect of selected IPM components individually and in combination on the natural enemies fauna present in the crop.

#### 5.1.7.1 Effect of the treatments on soil inhabiting natural enemies

During rabi 1998-99 endosulfan treatment significantly reduced the ground dwelling natural enemies at all stages of the crop growth with 60 to 75% reduction compared to control up to pod formation stage. Neem spray also caused 8 to 50% reduction compared to control where as IPM recorded 54% reduction during flowering stage, which may be due to application of endosulfan during this stage. IPM also recorded 36, 25% reduction of natural enemies during vegetative and podding stage. respectively but no reduction was observed during preharvest stage. The reduction of natural enemies in IPM plot was mainly due to application of endosulfan as third and neem as first and fourth sprays. Even though the treatments with HaNPV and bird perches recorded little reduction of natural enemies throughout the crop period compared to control it was not significant, hence were concluded as safer to the soil dwelling natural enemies. Thus it was concluded that chemical as well as neem to some extent affect the natural enemy fauna in the chickpea while the remaining components were safer.

During rabi 1999-2000 endosulfan treatment once again proved to be a harmful component of IPM to soil dwelling natural enemies with 65, 47, 63 and 75% reduction over control during vegetative, flowering, pod formation and preharvest stages, respectively. Neem caused a significant reduction at flowering and pod formation stages with 40 and 38% reduction
compared to control, respectively. IPM treatment caused a significant reduction at flowering and pod formation stages with 26 and 62% reduction only compared to control, respectively, where it received endosulfan and neem treatments. IPM treatment did not show any significant effect on number of natural enemies at vegetative and preharvest stages. HaNPV and bird perches caused little disturbance but not at significant level compared to control at flowering and pod formation stages and were relatively safer to natural enemies. During peak period of birds activity some reduction in natural enemies fauna was observed which was not significant.

Parmar (1993) reported that neem can be used in IPM system because of its relative safety compared to highly toxic chemicals. According to Krishnamoorthy (1995) several insecticides including endosulfan were found toxic to both larvae and adults of *C. carnea*. Both spider and ground beetle populations were known to be reduced by regular applications of insecticides (Pfrimmer, 1964). Bijjur *et al.* (1991) reported that *Apis cerana indica* did not show any signs of abnormal development due to its exposure to NPV. All the above statements support the present findings of harmful nature of endosulfan and relative safety of HaNPV on natural enemies present on ground as well as on crop foliage.

# 5.1.7.2 Effect of the treatments on natural enemies present on crop canopy during rabi 1998-99

Among the treatments endosulfan spray was found to have more effect on natural enemies present on crop canopy and reduced significantly both at 22 DAS and 54 DAS with 54 and 58% reduction over control, respectively. Where as neem was found to reduce significantly at 22 DAS because of its repellent action on natural enemies, but at 54 DAS it did not cause any significant reduction. Bird perches did not show any significant effect. In IPM there was 19% reduction of natural enemies at 22 DAS mainly due to neem and 47% reduction at 54DAS due to endosulfan which was given as third spray in IPM. HaNPV had negligible effect.

According to Krishnamoorthy (1995) several insecticides including endosulfan were found toxic to both larvae and adults of *C. carnea*. This report support the toxic effect of endosulfan to natural enemies present on foliage as observed in the present study. Bijjur *et al.* (1991) reported that *Apis cerana indica* did not show any signs of abnormal development due to its exposure to NPV, and this observation support the present results in which NPV did not show side effects on natural enemies present on foliage.

5.1.7.3 Effect of the treatments on the natural parasitism of H.armigera

During both the seasons the egg parasitisation was observed to be nil. The dead *Trichogramma* adults were noticed on 10-15% of the chickpea plants, which indicate the non suitability of chickpea habitat for survival and effectiveness of *Trichogramma* species due to acid exudates. This was supported by report of Yadava *et al.* (1985) who reported that inundative release of *T.chilonis* to control *H.armigera* was found ineffective in chickpea, and Bhatnagar (1981) confirmed the deterrent role of leaf exudates of chickpea on the activity of egg parasitoid *Trichogramma*.

During *rabi* 1998-99 a significant reduction of natural parasitisation by *C. chlorideae* (27%) was observed at 25 DAS i.e 4 days after endosulfan treatment. Neem and IPM which received neem as first spray significantly reduced the natural parasitism by *C.chlorideae* to 38 and 33%, respectively

compared to control. Where as in the remaining treatments no significant reduction of parasitism was observed. At 58 DAS that is 6 DAT there was no significant difference among the treatments for the larval parasitism by *C. chlorideae*. Except endosulfan all the other treatments were found relatively safer to *C. chlorideae*. However endosulfan was found more toxic to larval parasitoid immediately after spray and later its toxic effect reduced drastically.

During the season a very low incidence of larval-pupal parasitoid *C.illota* was observed. It was found to be 2% only in control plot in 58 DAS sampling.

During rabi 1999-2000 at 36DAS i.e 4 days after treatment, only endosulfan spray significantly reduced the larval parasitism by *C.chlorideae* to the extent of 42% compared to control. Whereas the remaining treatments were found to reduce parasitism to some extent but not significantly compared to control. At 68 DAS i.e 6 DAT there was no significant difference among the treatments for the larval parasitism by *C.chlorideae*. The incidence of *C.illota* was only 4%.

Nagarkatti (1981) reported 20-80% larval parasitisation by *C.chlorideae* and observed the maximum during December and January months. In the present study the parasitism level was 11% during first fortnight of December and 7% during first fortnight of January in *rabi* 1998-99 and during second fortnight of November 5% and 10% during second fortnight of December in *rabi* 1999-2000. These findings are in agreement with Yadava (1990) who reported 10% parasitisation of *C. chlorideae* on *H.armigera* in chickpea with peak activity between September and February.

## 5.1.8 Effect of the Treatments on Pod Damage by H.armigera

The perusal of the data during *rabi* 1998-99 revealed that IPM and endosulfan were found to be the best treatments by recording the lowest percentage of pod damage with 47 and 43% reduction compared to control, respectively. Neem and HaNPV gave similar protection to crop from pod damage by *H.armigera*. Bird perches also reduced the pod damage up to 27% compared to control since the birds activity was more at ICRISAT Center due to prevailing favourable conditions for their survival. The per cent pod damage was observed to be low in IPM due to contribution of all IPM components.

During the *rabi* 1999-2000 endosulfan was proved to be the best treatment with 49.7 % reduction of pod damage compared to control. IPM and endosulfan were at par in reducing pod damage. Neem and HaNPV were found equally effective in reducing pod damage followed by bird perches. Bird perches contributed to the extent of 31% reduction of pod damage compared to control.

Thakur et al. (1988) reported 13 and 5% pod damage at green pod and harvest stages respectively in neem leaf extract 5% treatment and 3 and 4% in NSKE 5% treatment. Pawar et al. (1990) reported 46% reduction in pod damage by *H.armigera* over control when HNPV @ 250 LE/ha was sprayed twice in chickpea. Saxena (1980) reported the promising role of birds to reduce pod damage by *H.armigera*. The pod damage of 1.4% to 14% due to application of endosulfan against *H.armigera* in chickpea was reported by Sanap and Deshmukh (1987) and Ujagir et al. (1997). The pod damage was observed to be 6.7% when endosulfan 0.05% was sprayed after NPV @250LE/ha in chickpea against *H.armigera* (Pawar et al., 1990).

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All the above reports suggested that the pod damage caused by *H.armigera* can be reduced by different IPM components individually and contributed in a synergistic manner to reduce the pod damage when given in combination in IPM plot with out any harmfull effects associated due to use of chemicals.

### 5.1.9 Effect of the IPM Options on Chickpea Yield

During rabi 1998-99 the results suggested 60% extra yield in IPM plot followed by 42% yield increase in plots treated with endosulfan compared to untreated area. HaNPV and neem were found equally effective and gave around 30% extra yield compared to control. Plots installed with only bird perches gave 15% extra yield compared to control.

During rabi 1999-2000 season endosulfan treated plot recorded 63% additional yield compared to control followed by IPM with 59% additional yield over control. HaNPV and neem were found equally effective, and bird perches also contributed 28% extra yield compared to control.

From the results it was clear that birds activity was more during *rabi* 1999-2000 than the previous year. But the contribution of birds may not be to this extent in farmers fields because of the favourable conditions for birds activity present in ICRISAT may not prevail in farmers fields, however one should try to take advantage of these natural resources.

Thakur *et al.* (1988) reported 31% yield increase due to NSKE 5% treatment in chickpea against *H.armigera* which was in agreement with the present finding. Pawar *et al.* (1990) reported 14-47% yield increase due to HNPV @250LE/ha against *H.armigera* in gram. Birds contributed 218 g/m<sup>2</sup>

increase in yield of wheat (ICAR, 1992). The increase in yield by 45% in chickpea due to application of endosulfan 0.07% was reported by Thakur *et al.* (1988). Sanap and Pawar (1998) reported 26.9 and 27.3 % increase in yield during 1993-96 due to IPM treatment by controlling *H.armigera*. All the above reports support the present findings of yield increase due to different IPM components which contributed in a synergistic way in IPM plot

#### 5.1.10 Economics of the IPM Components

During both the years bird perches showed the highest cost-benefit ratios due to less cost involved, but showed very less increase in yield compared to control. During *rabi* 1998-99 the highest cost benefit ratio was obtained with IPM treatment (1:2.3) followed by endosulfan (1:1.81). During *rabi* 1999-2000 also IPM gave highest cost benefit ratio of 1:3.76 followed by endosulfan (1:3.71). Even though HaNPV recorded the lowest cost benefit ratio due to its high cost of production it can be reduced by educating the farmers about its preparation. Even for neem also if the farmers prepare NSKE at farmhouse itself with locally available neem seeds the cost of production can be minimized which inturn increases the C<sup>·</sup>B ratio. All these finally reduce the cost of plant protection of IPM and it may prove much better than chemical treatment.

The cost benefit ratio of endosulfan 0.07% was reported as 1:5.2 by Parsai *et al.* (1989), as1:12 by Gupta *et al.* (1991) in chickpea against *H.armigera.* Thakur *et al.* (1988) also recorded the highest C:B ratio of 1:10 with endosulfan 0.07%, with NSKE5% as 1:7.7 and with neem leaf extract 5% as 1:3.9. Datkhile *et al.* (1996) reported that endosulfan 0.07% gave 1:5.3 C:B ratio where as it was 1:2.6 for HNPV @250LE/ha, 1:2.7 for neemark 0.2% and 1: 2 for NSKE in chickpea. All these reports conform the superiority of endosulfan compared to neem and HaNPV. Reddy and Manjunatha (2000) conducted experiment in consolidated block of 40 ha

cotton fields at two locations and demonstrated the superiority of IPM strategy in terms of both cost versus benefit ratio and environmental safety over that used in the farmers fields where only conventional control methods were followed. This report confirm the superiority of IPM in C:B ratio in the present experiment.

In the present study the cost of neem and HaNPV was considered along the market price and there is every possibility of producing them at farm level hence the C:B ratio with these treatments can be improved.

#### 5.1.11 Residues of Chemicals

Plants treated with endosulfan contained 0.81mg/kg residue in seed and 1.35 mg/kg in husk at harvest stage of the crop but no residues were found in IPM treatment plot. Even though the residues are less than the maximum residual limit given by FAO but may affect the health of consumers to some extent if consumed at green pod stage and the animals if they consumed green foliage of chickpea. This risk must be kept in mind while using chemicals on crops like chickpeas.

Pandey *et al.* (1977)<sup>b</sup> reported presence of residues at much higher than the tolerance limit even 25 days after spraying of endosulfan 0.07% both in plant and grain of chickpea and also reported the translocation and accumulation of residues in the grain. Verma (1983) stated that the residues of endosulfan fell below the tolerance limit in 12 days in grain after spraying with endosulfan 0.07%, and this report confirm the results in the present experiment.

## 5.2 LABORATORY EXPERIMENTS

#### 5.2.1 Oviposition Deterrenncy Effect of Neem Against H.armigera

The present studies clearly demonstrated the ovipositional deterrenncy effect of neem against *H.armigera*. There was a significantly lower mean number of eggs laid per pot (two plants) which were sprayed with neem 60 ppm compared to water sprayed pots (111.60, 287.8 eggs respectively).

This was in confirmity with the results of Jeyakumar and Gupta (1999) who reported that NSKE 10 and 7.5% treatments reduced oviposition by 60.9 and 59 % compared to control by *H.armigera* in chickpea. Present field results also support this statement that neem effectively deterred the egg laying by *H.armigera*.

### 5.2.2 Antifeedant Effect of Neem Against H.armigera.

When different age groups of *H.armigera* were given choice to choose their food between neem treated and untreated chickpea leaves and pods their capacity to differentiate was increased with age of the larvae. The data showed that the small larval group (first & second instars) did not show any significant differentiation between neem treated and untreated food, where as the medium sized group (third & fourth instars) and large sized group (fifth & sixth instars) showed significant differentiation between neem treated and untreated neem treated and untreated food.

When the *H.armigera* larvae were given neem treated food under no choice situation the initial behavior was more pronounced in older larvae with faster movement around the food, and tried in search of any other food. Since there was no alternative food the larvae finally settled on the neem treated food. It was clear that 40% of older larvae, 20% of medium sized larvae and none of the younger larvae showed initial avoidance of neem.

The above two experiments clearly indicated the antifeedant effect of neem on *H.armigera*. Contact with azadirachtin makes disruption in food intake and increases the locomotory activity of insects as reported by Schoonhoven *et al.* (1987) which was also observed in the present study. The antifeedant effect of neem extract was also reported by Murugan *et al.* (1993).

## 5.2.3 Effect of Neem on Different Age Groups of H.armigera

Unlike a chemical which cause direct kill of larvae neem shows different types of effects like repellency, feeding & oviposition deterrency and hormone like growth disrupting activity throughout the lifecycle of the pest and even affect fecundity also.

In the present study the results suggested that the effects of neem were more pronounced when *H.armigera* received treatment at early age. There was about 40% mortality in small larvae and 10 % in medium larvae and no mortality was recorded in large larvae.

Larval duration was significantly increased when fed with neem in all the three age groups compared to control. But even with more larval duration it can not cause more damage to crop because it suffers with several abnormalities like disruption of cuticle, reduced food intake and even activity also.

Pupal weight was significantly reduced in first and second age groups but not in third age group because of less time to experience the neem effect. Due to less food intake, reduced growth index, efficiency of conversion of ingested food and efficiency of conversion of digested food leads to reduced pupal weight compared to control. The effective oviposition period and fecundity were reduced in all the three age groups significantly compared to control. Where as no significant effect was observed in egg hatching with neem. Reduced fecundity in *H.armigera* due to NSKE was observed by Joshi and Sitaramaiah (1979) and reduced fecundity and hatching was also observed by Brattsen (1983) which confirmed the present results.

According to El-Sayeed (1985) 0.2-0.5% suspension of ground neem seeds caused complete mortality of *S. littoralis* by the end of larval period and also caused pupal mortality and adult deformity. Growth inhibitory effect of neem limnoids in *S.litura* was reported by Murugan and Jeyabalan (1995) confirming the effects of neem which were observed during the present experiment.

# 5.2.4 Efficacy of Robin Blue as an Ultraviolet Ray Protectant to Heliothis armigera Nuclear Polyhedrosis Virus

Major obstacle in the use of insect viruses in field situations is the rapid inactivation by ultraviolet rays. The results suggested that robin blue is a good ultraviolet ray protectant. Several workers tried with different adjuvants and UV ray protectants to increase the persistence and effectiveness of HaNPV under field conditions. But detailed studies work not taken up with robin blue which was cheap and readily available and easy to use.

Immediately after treatment there was no difference between HaNPV + robin blue 1% and HaNPV treatments in their effect against third instar larvae of *H.armigera*. But at 24 hours after treatment due to addition of robin blue 16.67% extra mortality of III instar larvae of pod borer was recorded which shows the effectiveness of robin blue1% in increasing the

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persistence of HaNPV. Where as 2<sup>nd</sup> and 3<sup>rd</sup> day after treatment there was 15.48 and 16.67% extra mortality, respectively compared to HaNPV treatment with out robin blue. But at fourth day after treatment the effect of HaNPV was very low and found to be on par with control but due to the addition of robin blue significantly higher mortality compared to control was recorded. At 5<sup>th</sup> and 6<sup>th</sup> day after treatment the effect of HaNPV was almost nil but due to addition of robin blue significant of robin blue significant treatment the effect of the effect of the effect.

Previously Rabindra and Jayaraj (1988) and Rabindra et al. (1989) reported increased efficacy of HaNPV with UV protectant like Robin blue which support the present findings.

#### 5.2.5 Effect of HaNPV on Different Age Groups of H.armigera.

Treatment of HaNPV @6x10<sup>9</sup> POB/I caused high mortality of *H.armigera* larvae at early stage compared to later stages indicating the capacity of larvae to with stand infection by HaNPV as age increases. This was supported by Ignoffo (1966) who reported that as the age of the *Heliothis zea* and *H.virescens* larvae increases their susceptibility to the HaNPV virus decreases.

When HaNPV treatment was given at first & second instar stage there was 100 % larval mortality, when it was given at third & fourth instar stage only 70% larval mortality but more pupal abnormality and death (97.9 %) was observed. In case of fifth & sixth instar stage the mortality was only 60 % but there was 87% pupal abnormality and death.

There was significant reduction in pupal weight when third & fouth instar larvae were fed with HaNPV. But this was not observed when larvae were treated with virus at fifth & sixth instar stage. This was supported by

Kencharaddi and Jayaramiah (1997). It was reported that the growth rate, gross and net efficiency of food utilization for body matter observed to be decreased during the course of HaNPV infection of *H.armigera* which ultimately leads to low pupal weight, but the time for showing this effect was less for fifth and sixth instar age group.

All these effects ultimately led to lower fecundity compared to control, where it was more pronounced in third & fourth instar age group compared to fifth & sixth instar age group. This finding was supported by Patil *et al.* (1989) where the HNPV treatment to *M.seperata* revealed an increase in development duration of larvae and pupae and also there was reduction in pupation, adult emergence rate, growth index, fecundity and average egg production, per cent egg hatchability. The same effects mentioned above were demonstrated in boll worm, *H.zea* by Luttrell *et al.* (1982).

#### 5.2.6 Effect of HaNPV Treatment on Fecundity of H.armigera

The moths collected from HaNPV treated population have less viability and capacity to mate. Most of the adults failed to emerge, some dead immediately after emergence, and some failed to mate and laid nonviable eggs, and, if mating occurred laid eggs only for few days and died.

When both male and female were collected from HaNPV treated population, there was significantly less number of eggs (689) compared to control (865). When only female was taken from HaNPV treated population, the fecundity was 718 eggs and with only male from HaNPV treated population the fecundity was 780 eggs compared to control (865).

This suggested the reduced fecundity of H.armigera by HaNPV treatment which was supported by the report of Luttrell et al. (1982) in the case of H.zea.

# 5.2.7 Effect of HaNPV Treatment on Egg Hatchability of H.armigera

A significant effect was observed on egg hatchability when larvae at fifth instar stage were treated with HaNPV. More significant reduction (30.5%) of hatchability was observed when male and female were taken from HaNPV treated population than with single sex from HaNPV. This result was supported by previous work of Patil *et al.* (1989) who observed the reduction of per cent egg hatchability in case of *M.separata* and Luttrell *et al.* (1982) in case of *H.zea* due to NPV treatment at larval stage.

## SUMMARY

Investigations were carried out on the effect of different IPM components on *H.armigera* and its natural enemies in chickpea ecosystem during 1998-99 and 1999-2000 *rabi* seasons at ICRISAT Center and the results obtained are summarized in this chapter.

- 1. The pest infestation was observed throughout the cropping period during both the years with peak population at 57 DAS i.e first week of January during rabi 1998-99 and at 68 DAS i.e last week of December during rabi 1999-2000
- 2. During both the years neem treatment effectively reduced the egg laying by *H.armigera* moths by acting as a oviposition deterrent and IPM which included neem as one of its component also effectively reduced the oviposition by *H.armigera* in chickpea under field conditions.
- 3. In both the years endosulfan was proved to be the best treatment in managing small size (first & second instars) larvae especially in the vegetative stage of the crop whereas IPM was superior in the flowering, pod formation and preharvest stages. HaNPV showed equal efficiency as that of neem during *rabi* 1998-99 but equally effective as that of endosulfan during *rabi* 1999-2000 in reducing the population of small size larvae.
- 4. IPM followed by HaNPV showed more effect in reducing medium size larvae in both the seasons than even endosulfan. Erecting bird perches was as effective as endosulfan in reducing medium size larvae during the peak period of bird activity.

- 5. At vegetative stage HaNPV was superior in managing large (fifth & sixth instar) larvae in *rabi* 1999-2000, but in the remaining period and during *rabi* 1998-99 throughout the crop period IPM maintained its superiority in managing the large size larvae. Erecting bird perches was as effective as endosulfan in reducing both medium and large size larvae during the peak period of bird activity.
- 6. Endosulfan and HaNPV proved effective in reducing total larval load during rabi 1998-99 & 1999-2000, respectively during vegetative stage but in the remaining stages IPM stood as a superior plant protection strategy in managing total larval load. Installation of bird perches contributed up to 26 and 37% reduction of larval load compared to control during the two years.
- 7. Endosulfan was observed to be harmful IPM component as it significantly reduced the total number of soil dwelling natural enemies in both the years and also the number of natural enemies present on crop canopy during *rabi* 1998-99. Neem also showed its ill effect on natural enemies and significantly reduced their number in both the seasons, present on ground as well as on foliage of the crop. Bird perches showed little disturbance to natural enemies during peak period of bird activity.
- 8. Egg parasitism by *Trichogramma* was not observed in both the years even though 10-15% of plants were observed with dead *Trichogramma* adults. Up to 11 and 10 % natural parasitism by *Campoletis chloredeae* Uchida, was recorded during *rabi* 1998-99 and 1999-2000 years, respectively. All IPM components except endosulfan were proved to be safe to natural parasitism by

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C.chlorideae. A very low level of larval-pupal parasitoid Carcelia illota Curron (2-4%) incidence was observed in both the years.

- 9. Even though endosulfan recorded the lowest pod damage and higher yield during 1999-2000, IPM treatment proved more economical than other components individually except bird perches with 1:2.30 and 1:3.76 cost benefit ratio's in both the years.
- 10. The plots treated with endosulfan were found to have residues of 0.81mg/kg in seed and 1.35mg/kg in husk at harvest stage, but in IPM plots the residues were found to be below detectable limit.
- 11. Neem effectively showed its oviposition deterrence effect with 111 60 eggs on neem treated foliage as against 287.8 eggs on control foliage which was 63% less.
- 12. When *H.armigera* larvae were given a choice to choose its food between neem treated and untreated food the capacity to differentiate increased with age of the larvae. When *H.armigera* was given neem treated food in no choice situations the searching behavior for another food source also increased with age.
- 13. Neem treatment showed different types of effects on *H.armigera* throughout its life cycle. The effects were more pronounced when treatment was given at early period of larval stage. The effects observed were increased larval duration, reduced pupal weight, reduced effective oviposition period and fecundity with no significant effect on egg hatching.

- 14. Robin blue 1% was proved to be a good ultraviolet light protectant and increased the persistence of HaNPV up to six days under field conditions with increased efficiency of HaNPV from 24 hours after treatment.
- 15. HaNPV @ 6x10<sup>9</sup> POB/I was found to have more impact on early stages of larvae than later stages. HaNPV treatment in addition to higher larval mortality resulted in pupal abnormality and death, reduced pupal weight, adult emergence.
- 16. When *H.armigera* larvae received HaNPV infection at fifth instar stage they may escape from higher rate of mortality but the fecundity of adults was reduced up to 20% and the egg hatching up to 30% The reduction in egg hatching was pronounced when male and female were from HaNPV treated population rather than of single sex from HaNPV treated population.

- Abhisek Shukla and Goydani B M 1996 Evaluation of nuclear polyhedrosis virus for the control of *Helicoverpa armigera* (Hubner) on chickpea under the agroecosystem of Satpura plateau region of Madhya Pradesh. Advances in Plant Sciences 9:143-146.
- Achan P D, Mathur K C, Dharmadhikari P R and Manjunath T M 1968 Parasites of *Heliothis* Spp. in India. Technical Bulletin, Common Wealth Institute of Biological Control 10:129-149.
- Ahmed K, Lal S S, Morris H, Khalique F and Malik B A 1990 Insect Pest problems and recent approaches to solving them on chickpea in South Asia. In, Chickpea in the nineties:Proceedings of the second International workshop on chickpea improvement, ICRISAT. Patancheru, India, 4-8 December 1989. pp.165-168.
- Ahmed R, Katti G, Saxena Hem and Sachan J N 1996 Evaluation of REF Chemicals for management of *Heliothis armigera*. Paper present in Indian Science Congress Association, 83<sup>rd</sup> Session, January 3 – 8, 1996. University, Patiala.
- Ali Salim 1996 Book of Indian Birds. Bombay Natural History Society, Mumbai, India.
- Allen G E and Ignoffo C M 1969 The nuclear polyhedrosis virus of Heliothis zea quantitative in vivo estimates of virulence. Journal of Invertebrate Pathology 13:378-381.
- Anitha/Mistry, Yadava D N, Patel R C and Parmar B S 1984 Field evaluation of Nuclear Polyhedrosis Virus against Heliothis armigera Hubner (Lepidoptera : Noctuidae) in Gujarat. Indian Journal of Plant Protection 12: 31-33.
- Armes<sup>/</sup>N J, Jadhav D R, Bond G S and King A B S 1992 Insecticide resistence in *Heliothis armigera* in South India. Pesticide Science 34 : 355-364.
- \*Ascher K R S 1993 Non-conventional insecticidal effects of pesticides available from the neem tree *Azadirachta indica*. Arch. Insect Biochem. Physiology 23: 433-449.

- Balandrin M S, Lee S M and Klock J A 1988 Biologically active volatile organosulphur compounds from seed of the neem tree, Azadirachta indica (Meliaceae). Journal of Agricultural and food chemistry 36:1048-1054.
- Banken A Julie and Stark D John 1997 Multiple routes of pesticide exposure and the risk of pesticides to biological controls : A study of neem and seven spotted lady beetle (Coleoptera : Coccinellidae). Journal of Economic Entomology 91:1-6.
- Banken A O Julie and Stark D John 1998 Stage and age influence on the susceptibility of *Coccinella septempunctata* (Coleoptera : Coccinellidae) after direct exposure to Neemix, a neem insecticide. Journal of Economic Entomoloty 90:1102-1105.
- Barkhade U P, Pawar W S and Thokal M R 1991 The effect of pesticidal application at different growth periods of the level of infestation of pod borer *Helicoverpa armigera* (Hubner) on chickpea. Journal of Soils and Crops 1:182-183.
- Barrett J H 1967 *Heliothis* species Lepidoptera : Noctuidae in Papua and New Guinea. Papua New Guinea agricultural Journal 19:61-66.
- Bell M R 1982 The potential use of microbials in *Heliothis* management. In, Reed W and Kumble V (Eds) Proceedings of the International Workshop in *Heliothis* Management, 15-20 November, 1981., Patancheru, India, ICRISAT. pp.123-135.
- Bhagwat V R 1997 ICRISAT's ecofriendly gift to check chickpea pod borer. SAT news 20:6-8.
- Bhatnagar V S 1981 Are effective parasites of *Helicoverpa* eggs found on Pigeonpea and Chickpea?. International Pigeonpea Newsletter 1:32.
- Bhatnagar V S and Davies J C 1978 Factors affecting population of gram pod borer, *Heliothis armigera* (Hubner) in the period 1974-77 at Patancheru (Andhra Pradesh). Bulletin of Entomology 13:52-64.

- Bijjur S, Kulkarni K A and Lingappa S 1991 Safety of *Heliothis armigera* (Hubner) NPV to some non-target beneficial organisms. Indian Journal of Entomology 53:475-478.
- Bilapate G G, Mokat R B, Lovekar R C and Bagade D N 1988 Population ecology of *Heliothis armigera* (Hubner) and its parasites on pulses. Journal of Maharashtra agricultural Universities 13:299-302.
- Boucias D G, Johnson D W and Allen G E 1980 Effect of host age, virus dosage and temperature on the infectivity of a nuclear polyhedrosis virus against velvet bean caterpillar, Anticarsia gemmatalis larvae. Environmental Entomology 9:59.
- \*Brattsen L B 1983 Cytochrome P 450 involvement in the interactions between plant terpens and insect herbivores. In, Plant resistance to insects. (ed Hedin P A), ACS Symposium Series 208 : 173 – 195.
- \*Breethaupt J 1995 Untersuchungen zur Bekampfung des Asiatischen Maiszunslers ostrinia furnacalis Guence (Lepidoptera : Pyralidae) unter vermeidung des einsatzes synthetischer Insektizide im Markhamtal und im sudlichen Hochland in Papua Neuguinea. Doctor thesis, Unversity of Giessen, Germany.
- \*Breuer M and Schmidt G H 1996 Wirkung einermit Melia azedarach extract behandelten Reupendiat anf wachsttom entwick lund and Fekunditat Von Spodoptera frugiperda (Smith J C) (Lep : Noctuidae). Z. Pflanzenkr Pflanzenschutz 103 : 171 – 194.
- Butani P G and Mittal V P 1993 Comparative efficacy of botanical insecticide (neem seed kernal suspension) and other insecticides against gram pod borer (*Heliothis armigera* Hubner). In, Botanical pesticides in Integrated Pest Management, Indian Society of Tobacco Science, ISTS, Rajahmundry, India. pp. 276-281.
- \*Butterworth J H and Morgan E D 1968 Isolation of a substance that suppresses feeding in locusts chemicals. Commun. 1 : 23 24.

- \*Cano V and Gladstone S M 1994 Effecto de insecticida botanica, NIM-20, sobre el parasitismo por *Trichogramma pretiosum* an hueros del *Heliocoverpa zea* en el cultivo de melon. Manecho Integrado de plagas 33 : 23 - 25.
- Chaudhary R R P and Sachan R B 1995 Influence of sowing dates and use of insecticides on the infestation of gram pod borer in chickpea in western plain zone of Uttar Pradesh. Bhartitya Krishi Anusandhan Patrika 10:143-150.
- Chaudhary S 1997 Effect of age of Spilosoma obliqua larvae on their susceptibility to nuclear polyhedrosis virus. Indian Journal of Entomology 59:59-61.
- Chauhan R and Dahiya B 1991 Estimation of avoidable losses to chickpea due to pod borer in Haryana. Legume Research 14 205-207.
- Chawla R P and Joia B S 1992 Evaluation of carbaryl, endosulfan and quinalphos residues in pigeonpea grains. International Pigeonpea Newsletter 15:31-32.
- Chen C C, Yaw-Jen Dong, Ling-Lan Chang and Roger F Hoel 1996 Deterrent effect of neem seed kernel extract on oviposition of the fruit fly (Diptera : Tephritidae) in Guava. Journal of Economic Entomology 89:462-466.
- Chhabra K S 1990 Present status of *Helicoverpa* on pulses and strategies for its management in Punjab. In, Sachan J N (Eds)
   First National Workshop on *Heliothis* management: Current status and future strategies. Directorate of Pulses Research, Kanpur, India, 30-31 August, 1990. pp. 57-70.
- Chundurwar R D and Pawar V M 1991 Natural incidence of nuclear polyhedrosis virus infection in *Heliothis armigera* (Hub.) on chickpea. Journal of Maharashtra agricultural Universities 16:420-421.

- \*Cilgi T, Framptom G K, Wratten S D and Holland J M 1993 The long term effects of pesticides on benefical invertebrates lessons from the Bonworth project. Pesticide outlook 4:30-35.
- Cowgill S E and Bhagwat V R 1996 Comparison of the efficacy of chemical control and Helicoverpa NPV for the management of *Helicoverpa armigera* (Hubner) on resistant and susceptible chickpea. Crop protection 15:241-246.
- Datkhile R V, Pawar S A and Mote U N 1996 Efficacy of different insecticides against pod borer on chickpea. Journal of Maharashtra agricultural University 21:204-206.
- Datkhile R V, Pawar S A, Mote U N and Khaire V M 1992 Bioefficacy of different insecticides against gram pod borer *Heliothis armigera* Hubner on chickpea *Cicer arietinum* L. Bioecology and control of insect pests. In, Goel S C (Eds) Proceedings of the National Symposium on Growth, Development and control technology of insect pests, Uttar Pradesh Zoological Society, Muzaffarnagar, India. pp.156-160.
- Daware D G and Dhanorkar B K 1981 Chemical control of pod borer of red gram. Pesticides 150:35.
- Deka N K, Prasad D and Chand P 1989 Chemical control of *Helicoverpa* armigera (Hubner) in chickpea. Research and Development Reporter 6:130-137.
- Dethe M D and Kale V D 1991 An analysis of HCH and endosulfan residue in chickpea seed. International Chickpea Newsletter 24:30-31.
- Dhamdhere S G and Khaire V M 1986 Field evaluation of different doses of nuclear polyhedrosis virus of *Heliothis armigera* (Hubner). Current Research Reporter 2:221-226.
- Dhurve S B and Borle M N 1985 Chemical control of gram pod borer (*Helicoverpa armigera* Hubner). PKV Research Journal 9:83-85.

- Dorn A, Rademacher J M and Sehn E 1986 Effects of azadirachtin on the moulting cycle, endocrine system and ovaries in last instar larvae of the milk weed bug Oncopeltus fasciatus. Journal of Insect Physiology 32:321-328.
- \*Eisenlohr K, Dimange A L, Lenfant C and Sauphanor B 1992 Effects of Neem Azal-F on aphids and beneficial insects in peach orchards in France. In, Kleenberg H (Eds) Proceedings of 1<sup>st</sup> Workshop Practice Oriented Results on use and Production of Neem Ingradients. Giessen : Druck and Graphic. pp. 27-40.
- El-Sayeed El 1985 Evaluation of the insecticidal properties of the common Indian neem seeds against the Egyptian leaf worm *Spodoptera littoralis*. Bulletin of Entomological Society of Egypt 13:39-47.
- Évans H F 1981 Quantitative assessment of the relationship between dosage and response of the nuclear polyhedrosis virus of *Memestra brassicae*. Journal of Invertebrate Pathology 37:101-109.
- \*Fagoonee I 1981 Behavioural responses of *Crocidolomia binotalis* on neem. In, Proceedings of 1<sup>st</sup> Intenational Neem Conference, Rottach-Egern, 1980. Eschborn : GTZ. pp. 109-20.
- Farrar R R JR and Ridway R L 1999 Relative Potency of Selected Nuclear Polyhedrosis Viruses against five species of Lepidoptera. Journal of Agricultural and Urban Entomology 16:187-196.
- Fernandez N J, Palanignan E L, Soon L L and Botrell D G 1992 Impact of neem on non target organisms. In, Proceedings final workshop Botanical Pest Control Project Phase 2, IRRI, Los Banos, Philippines. pp.117 – 121.
- Fitt G P 1989 The ecology of *Helicoverpa* spp. In relation to agroecosystems. Annual Review of Entomology 34:17-52.
- Garg D K 1989 Campoletis chlorideae Uchida, a larval parasite of Helicoverpa armigera (Hubner) in the Kumaon Hills, Uttar Pradesh, India. International Chickpea Newsletter 20:8-9.

- Ghode M K, Nayak U K, Ghosh P K and Pawar A D 1988 Avian predation of gram pod borer *Helicoverpa armigera* Hubner in Orissa. Journal of Advanced Zoology 9:148.
- Ghosh P K, Devendra P, Prasad and Shaw S P 1989 Management of pod borer, Helicoverpa armigera (Hubner) in chickpea. In, Das D K and Sarkar K R (Eds) Proceedings of the National seminar on Integrated Management Approach for maximizing crop production in rainfed and problem areas, Indian Society of Agricultural Sciences, IARI, New Delhi, India, 26-28 February 1986. pp.122-126.
- Giraddi R S, Goudreddv B S and Patil B V 1994 Critical time of spray in chickpea for the control of gram pod borer, *Heliothis armigera* (Hubner). Karnataka Journal of Agricultural Sciences 7:79-81
- Gopal M and Mukharjee I 1993 Determination of residues of endosulfan and endosulfan sulphate on egg plant, mustard and chickpea. Pesticide Science 37:67-72.
- Gopal M, Mukharjee I, Dikshit A K, Roy N K, Nagia D K, Sharma M L and Kumar S 1988 Persistence of endosulfan and its metabolites on brinjal and gram crop. Indian Journal of Plant Protection 16:177-183.
- Greathead D J and Girling D J 1982 Possibility for natural enemies in Heliothis management and contribution of the Common Wealth Institute of Biological Control. In, Reed W and Kumble V (Eds) Proceedings of the International Workship in Heliothis Management, 15 – 20 November, 1981. Patancheru, India, ICRISAT. pp.147-158.
- Gudauskas R T and Canerday D 1968 The effect of heat, buffer salt and H-ion concentration and ultraviolet light on the infectivity of *Heliothis* and *Trichoplusia* nuclear polyhedrosis viruses. Journal of Invertebrate Pathology 126:405-411.
- Gunasekaran K and Balasubramanian D M 1987 Field efficacy of diflubenzuron against *Heliothis armigera* Hb. in chickpea. Madras Agricultural Journal 74:52-53.

- Gupta M P and Thakur B S 1990 Avoidable losses in grain yield of chickpea due to damage caused by pod borer. Indian Journal of Pulses Research 3:45-47.
- Gupta M P, Parsai S K and Gupta D P 1990 Bioefficacy and economics of certain insecticides and vegetable oils against gram pod borer, *Helicoverpa armigera* Hubner on chickpea. Indian Journal of Plant Protection 18:207-211.
- Gupta M P, Thakur B S and Shrivastava S K 1991 Spray schedule of endosulfan for gram pod borer (*Helicoverpa armigera* Hubner) in chickpea (*Cicer arietinum*). Indian Journal of Agricultural Sciences 61:860-861.
- Hardwick D F 1965 The corn ear worm complex. Mem. Ext. Soc.Canada 40 : 247.
- Heinz K M Mc, Cutchen B F, Herrmann R, Parrella M P and Hammock B D 1995 Direct effects of recombinant nuclear polyhedrosis viruses on selected nontarget organisms. Journal of Economic Entomology 88:860-861.
- \*Hellpap C 1985 Populationsokologie and biologischbiotechnische Bekampfung vond Spodoptera spp. in Nicaragna. Doctor thesis, University of Frankfurt and Giessen, Germany.
- Hongo H and Karel A K 1986 Effect of plant extracts on insect pests of common bean. Journal of Applied Entomology 102:164-169.
- Ibrahim Ali M, Dand Miah M and Karim M A 1993 Efficacy of two bioinsecticides in controlling the *Helicoverpa armigera* (Hubner) in chickpea. Legume Research 16:91-94.
- ICAR 1992 Research highlights of AICRP on agricultural ornithology (ed A K Raheja) pp.16.
- Ignoffo<sup>-/</sup>C M 1966 Effect of larvae age on mortality of *Heliothis zea* and *Heliothis virescens* larvae exposed to a nuclear polyhedrosis virus. Journal of Invertebrate Pathology 8:279-282.

- \*Ignoffo C M and Falcon L A 1978 Formulation and application of microbial insecticides. Misc. publ ent. Soc. Am., 10:550-575.
- Ignoffo C M and Garcia C 1992 Estimation of environmental factors and sunlight affecting activity of inclusion bodies of the *Heliothis* (Lepidoptera : Noctuidae) nuclear polyhedrosis virus. Environmental Entomology 210 – 213.
- Ingalhalli S S, Savanurmath C J and Hinchigeri S B 1995 Carbohydrate contents at the primary active site of Nuclear Polyhedrosis Infection in the armyworm Mythimna (Pseudoletia) seperata. Entomon 20: 49-51.
- International Crops Research Institute for Semi-Arid Tropics Chickpea Biotic Stresses, Insect Pests. Annual Report for 1987, Patancheru, Andhra Pradesh, India. 138-146.
- Isman M B, Lowery D T and Koul O 1992 Laboratory and field evaluation of neem for control of aphid and Lepidopteran pests. In, Resources for sustainable agriculture :The use of neem and other plant materials for pest control and rural development. Proc. XVII Pacific Science Cong., Honolulu.
- Jabbar A, Anwar T and Khalique F 1988 Suitability of neem (*Azadirachta indica*) for the control of agricultural insect pests. In, Proceedings of 8th Pakistan con. Zool., Peshawar, pp. 147-148.
- Jain P C and Singh S 1988 Management of gram pod borer Heliothis armigera Hubner on bengal gram by insecticides. In, Integrated pest control : progress and perspectives : proceedings of the national symposium, Trivandrum, Kerala, India 15 – 17 October, 1987. pp. 309-312
- Jaques R P 1985 Stability of insect viruses in the environment. pp. 285-360, In Viral insecticides for biological control (ed Maramorosch K and Sherman K) Academic press, England.

- Jayachandran G and Chaudhari S 1996 Effect of age-related response of Spodoptera litura Fab. larvae on susceptibility to nuclear polyhedrosis virus. Indian Journal of Entomology. 58:275-279.
- Jayaraj S 1992 Pest Management in pulses on overview. In, Sachan J N (Eds) Proceedings of the National Symposium on New Frontiers in Pulses Research and Development, Directorate of Pulses Research, Kanpur, India, 10-12 November, 1989. pp.154-165.
- Jayaraj S, Rabindra R J and Santharam G 1987 Control of Heliothis armigera (Hubner) on chickpea and lablab bean by nuclear polyhedrosis virus. Indian Journal of Agricultural Sciences 57:738-741.
- Jeyakumar P and Gupta G P 1999 Effect of neem seed kernel extract (NSKE) on *Helicoverpa armigera*. Pesticide Research Journal 11:32 - 36.
- Jodha N S and Rao S K V 1987 Chickpea worlds importance and distribution. In the chickpea in ninetees (ed Saxena M C and Singh K B) Wallingford, Oxon, UK : CAB International. pp.1-10.
- Joginder Singh, Sadhu S S and Singla M L 1990 Ecology of Heliothis armigera (Hub) on Chickpea in Punjab. Journal of Insect science 3:47 – 52.
- Joshi B G and Sitaramaiah S 1979 Neem seed kernel as on oviposition repellent for *Spodoptera litura* (F) moths. Phytoparasitica 7:199-202.
- Joshi B G, Ramaprasad G and Sitaramaiah S 1982 Effect of neem seed kernel suspension on *Telenomus remus* on egg parasite of *Spodoptera litura*. Phytoparasitica 10:61-63.
- \*Kaethner M 1991 Keine Nebenwirkungen Von Niemprodukten auf die aphidophagen pradatoren Chrysoperla carnea (Steph.) ond coccinella septempunctata L. Anz. Schadling S K., Pflanzensch. Umweltsch. 64 : 97 – 99.

- Karnavar G K 1987 Influence of azadirachtin on insect nutrition and reproduction. Proceedings of Indian Academy of science (Animal Science) 96:341-347.
- Kaul C K, Mehrotra P and Singh S D 1988 Chemical control of gram pod borer. Indian Journal of Entomology 50:532-533.
- Kencharaddi R N and Jayaramiah M 1997 Dosage mortality response of nuclear polyhedrosis viruses in two species of field bean pod borers. Mysore Journal of Agricultural Sciences 3:47-50.
- Khan M 1996 Use of newer insecticides for the control of pod borer, *Helicoverpa armigera* Hb. on chickpea. In, Wanjari K B, Raut B T and Potdukhe S R (Eds) Seminar on Strategies for increasing pulse production in Maharashtra, Mumbai, India, 7-8 March 1996. pp.39-40.
- Khan M M, Rustamani M A, Talpur M A, Balouch H B and Chhutto A B 1993 Efficacy of different insecticides against *Helicoverpa armigera* (Hubner) on gram. Pakistan Journal of Zoology 25:117-119.
- King A B S and Sawicki R M 1990 Insecticide resistance of Helicoverpa and its management. In, Chickpea in the ninetees : Proceedings of the Second International Workshop in Chickpea Improvement, ICRISAT Center, Patancheru, Andhra Pradesh, India, 4-8 Dec 1989. pp.195-201.
- King E G, Powell J C and Smith J W 1982 Prospects for utilization of parasites and predators for management of *Heliothis* Spp. In, Reed W and Kumble V (Eds) Proceedings of the International Workshop in *Heliothis* Management, 15 – 20 November, 1981. Patancheru, India, ICRISAT. pp.103-122.
- Kostandy S N 1995 The simultaneous effect of early using of insecticides on cotton pests and its related natural enemies. Annals of Agricultural Sciences, (Ain Shams University, Cairo) 40:877-889.
- Koul O 1987 Antifeedant and growth inhibitory effect of calamus oil and neem oil on Spodoptera litura under laboratory conditions. Phytoparasitica 15:169-180.

- Koul O, Shankar J S and Kapil R S 1996 The effect of neem allelochemicals on nutritional physiology of larval *Spodoptera litura*. Entomologia Experimentalis et. Applicata. 79 : 43 50.
- Krishnamoorthy A 1995 Effect of several pesticides on eggs, larvae and adults of the green lace wing Chrysoperla carnea Stephens. Entomon 10:21-28.
- \*Kubo I and Klocke J A 1982 Azadirachtin : Insect ecdysis indibitor. Agricultural Biological Chemistry 46:1951-1953.
- Lal S S 1990 Present status of Helicoverpa armigera (Hubner) on pulses and future strategies for its management in Uttar Pradesh. In, Sachan J N (Eds) First National workshop on Heliothis management: Current status and future strategies. Directorate of Pulses Research, Kanpur, India, 30-31 August, 1990. pp.34-41.
- Lal S S 1992 Scope and limitation of integrated pest management in chickpea. In, Sachan J N (Eds) Proceedings of the National Symposium on New Frontiers in Pulses Research and Development, Directorate of Pulses Research, Kanpur, India, 10-12 November, 1989. pp.139-155.
- Lamb R and Saxena R C 1988 Effect of neem seed derivatives on rice leaf folders (Lepidoptera : Pyralidae) and their natural enemies. In, Proceedings of final workshop IRRI-ADB-EWC Project on Botanical Pests Control in Rice based cropping systems, IRRI, Los Banos, Philippines. pp. 47.
- Li K H, Xu X, Li Y F, Mexg Q Z and Zhav L C 1986 Determination of toxicity of 29 chemicals to *Trichogramma japonicum* at various developmental stages. Natural Enemies of Insects 8:187-194.
- Lowery D T and Isman M B 1995 Toxicity of neem to natural enemies of aphids. Phytoparasitica 23:297-306.
- Luff M L 1987 Biology of Polyphagous ground beetles in agriculture. Agricultural Zoology Review 2:237-278.

- Luttrell R G, Yearian W G and Young S Y 1982 Effect of Elcar Heliothis zea nuclear polyhedrosis virus treatment on Heliothis species. Journal of Georgea Entomological Society. 17:211-221.
- Mahajan S V, Sable K R and Thorat R N 1990 Present status of Helicoverpa on pulses and strategies for its management in Maharashtra. In, Sachan J N (Eds) First National workshop on Heliothis Management: Current status and future strategies. Directorate of Pulses Research, Kanpur, India, 30-31 August, 1990. pp.71-77.
- Mala S R, Peter C and David B V 1993 Ovipositional behaviour and eclosion of eggs of *Heliothis armigera* as affected by insecticides. Entomon 17:171-181.
- Malathi S, Sriramulu M and Ramesh Babu T 1999 Effect of certain ecofriendly insecticides on the egg parasitoid *Trichogramma chilonis* (Ishii) (Hymenoptera : Trichogrammatidae). Journal of Research ANGRAU 27:1-4.
- Mansour F A, Ascher K R S and Abo-Mosh F 1993 Efffect of Margosan-O <sup>™</sup> Azatin <sup>™</sup> and RD9 Repellin on spiders, and on predacious and phytophagous mites. Phytoparasitica 21:205-211.
- Markandeya V and Diwakar B J 1999 Effect of a neem formulation on four bioagents. Plant Protection Bulletin 15:28-29.
- Mc Kinley D J 1982 The prospects for the use of nuclear polyhedrosis virus in *Heliothis* Management. In, Reed W and Kumble V (Eds)
  Proceedings of the International Workshop in *Heliothis* Management 15 20, November, 1981. Patancheru, India, ICRISAT. pp.123-135.
- Mehto D N, Singh K M and Singh R N 1986 Natural enemy complex on insect pest complex in chickpea *Cicer arietinum* Linn. Bulletin of Entomology 27:1-12.
- Mishra B K, Mandal S M A and Tunga S 1992 Seasonal activity of parasitoids of *Helicoverpa armigera* (Hubner) in the eastern ghat

high land zone of Orissa. Orissa journal of Agricultural Research 5:170-173.

- Misra M P, Pawar A D and Ram N 1991 Use of NPV in management of the insect pest, *Heliothis armigera* (Hubner) in gram. Journal of the Andaman Science Association 7:75-78.
- Mordue (Luntz) A J and Blackwell A 1993 Azadirachtin Annual update. Journal of Insect Physiology 39:903-924.
- Morian D F, Larew H G, Krodel J J and Natoli W 1990 Systemic activity of neem extract against the leaf birch miner. Journal of Arboriculture. 16:12-16.
- Morris O N 1977 Long term effects of aerial application of virus fenitrothion combinations against spruce budworm, *Choristoneura fumiferana* (Lepidoptera : Noctuidae). Canadian Entomologist 109:1-14.
- Murugan K and Jeyabalan D 1995 Antifeedant and ovipositional effect of neem root extract (*Azadirachta indica* A.Juss.). Neem Newsletter 12:45-46.
- Murugan K, Jahanmohini P and Babu R 1993 Effect of neem kernel extract and neem oil on nutritive and reproductive physiology of *Heliothis armigera* Hubner In, Proceedings of World Neem Conference, Bangalore, India. 1:321-334.
- Murugan K, Jeyabalan D, Senthil L and Babu R 1998 Antifeedant and growth inhibitory properties of neem limnoids against the cotton boll worm, *Helicoverpa armigera* (Hubner). Insect Science and Application 18:157-162.
- Murugan K, Senthil Kumar N, Babu R and Senthil Nathan S 1995 Antifeedant and ovipositional deterrent effects of *Azadirachta indica* A. Juss. Neem Newsletter 12:43-44.
- Murugesan S and Jacob J P 1994 Antifeedant and growth disruption activity of azadirachtin on *Heliothis armigera* (Hubner), *Spodoptera litura* (F.) and *Atractomorpha crenulata* (F). Journal of Arid Zone 20:21-25.

- Nagarkatti 1981 The utilization of biological control of *Helicoverpa* management in India. In, Proceedings of the International workshop on *Heliothis* management, 1981, ICRISAT, Patancheru, India. 15-20 November.
- Nagarkatti S 1982 The utilization of biological control in Heliothis management in India. In, Reed W and Kumble V (Eds) Proceedings of the International Workshop in Heliothis Management, 15 – 20 November, 1981. Patancheru, India, ICRISAT. pp.159-167.
- Naseema Beevi S, Thomas Biju Mathew and Visalakshi A 1997 Dissipation of endosulfan in cowpea. Journal of Tropical Agriculture 35:41-43.
- Neumann K and Isman M B 1995 Evaluation of neem Azadirachta indica seed extracts and oils as oviposition deterrents to noctuid moths. Entomologia Experimentalis et. Applicata 76:115-120.
- Neumann K and Isman M B 1996 Toxicity of neem (Azadirachta indica A.Juss.) seed extracts to larval honey bees and estimation of dangers from field applications. American Bee Journal 136: 518-520.
- Noorani A M, Shah A D, Jugtani T K and Lohar M K 1994 Efficacy of different insecticides against gram pod borer, *Helicoverpa armigera* Hubner on gram crop under field conditions. Sarhad Journal of Agriculture 10:183-186.
- Odak S C 1982 Integrated Control of gram pod borer, Heliothis armigera Hubner (Lepidoptera : Noctuidae). ICRISAT Annual report for 1982. pp. 106.
- Odak S C, Srivastava D K, Mishra V K and Nema K K 1982 Preliminary studies on the pathogenicity of *Bacillus thuringiensis* and nuclear polyhedrosis virus on *Heliothis armigera* host in the laboratory and in pot experiments. Legume Research 5:13-17.
- Padmaja P G and Rao P J 2000 Efficacy of certain plant oils on the American boll worm Helicoverpa armigera Hubner. Pesticide Research Journal. 12:107-111.

- Pandey S Y, Dixit A K, Jain H K and Agnihothri N P 1977\* Residue of lindane and endosulfan in/on pea plant (*Pisum sativum*). Indian Journal of Entomology 39:85-87.
- Pandey S Y, Jain H K, Agnihothri N P, Dewan R S and Saxena H P 1977<sup>b</sup> Residues from foliar application of carbophenothion, tetrachlorvinphos, dicrotophos, trichlorofos and endosulfan on bengalgram (*Cicer arietinum*). Indian Journal of Plant Protection 5:47-50.
- Parasharya B M 1995 Role of beneficial birds in agricultural ecosystem. Journal of the Bombay Natural History Society 92:11-15.
- Parihar N S, Gupta A and Singh V 1990 Endosulfan residue in chickpea. International Chickpea Newsletter 23: 16-17.
- Parmar B S 1993 Scope of botanical pesticides in Integrated Pest Management. Journal of Insect Science 6:15-20.
- Parsai S K, Chaudhary R K and Sahu H R 1989 Comparative efficacy and economics of some pyrethroid and non pyrethroid insecticides against *Helicoverpa armigera* (Hubner). Indian Journal of Pulses Research 2:147-151.
- Parsai S K, Sahu H R and Chaudhary R K 1990 Effect of common insecticides and synthetic pyrethroids on flowers of chickpea and pigeonpea. Indian Journal of Plant Protection 18:284.
- Patel A J, Dhulia F K and Bharodia P S 1990 Present status of Heliothis armigera (Hub.) in pulses and strategies for its management in Gujarat. In, Sachan J N (Eds) First National Workshop on Heliothis management : Current status and future strategies, Kanpur, Uttar Pradesh, India. pp.188-192.
- Patel J J, Patel N C, Jayani D B, Patel J R and Patel B D 1997 Bioefficacy of synthetic and botanical insecticides for controlling pod borer (*Helicoverpa armigera*) and pod fly infesting vegetable pigeonpea (*Cajanus cajan*). Indian Journal of Agricultural Sciences 67:117-119.

- Patel M H 1988 Studies on predation of Helicoverpa armigera Hubner and Spodoptera litura Fab. by insectivorous birds with special reference to Acridotheres tristis (L). M.Sc. (Ag.) thesis, submitted to Gujarat Agricultural University, Anand, Gujarat.
- Patil U R, Savanurmath C J, Mathad S B, Aralaguppi P I and Ingahalli S S 1989 Effect of nuclear polyhedrosis virus on the growth, development and reproduction in surviving generations of the armyworm Mythimna (Pseudoletia) separata (walker). Journal of Applied Entomology 108:527-532.
- Patnaik H P, Rath L K, Senapathi B and Behera P K 1991 Incidence of Helicoverpa armigera Hubner on chickpea and its population phenology in north central plateau zone of Orissa. Orissa Journal of Agricultural Research 4:137-142.
- Pawar C S, Bhatnagar V S and Jadhav D R 1986 Host plants and natural enemies of *Helicoverpa* species in India: a compendium ICRISAT progress report 13, pp107.
- Pawar C S, Bhatnagar V S and Jadhav D R 1989 Campoletis chlorideae Uchida (Hymenoptera : Ichneumonidae) as a parasite of Heliothis armigera (Hub.) (Lepidoptera : Noctuidae) in southwest India. Proceedings of the Indian Academy of Sciences (Animal Scieces) 98:259-264.
- Pawar V M, Aleemuddin M and Bhosle B B 1987 Bioefficacy of HNPV in comparison with endosulfan against pod borer on chickpea. International Chickpea Newsletter 16:4-6.
- Pawar V M, Chundurwar R D, Kadam B S, Thombre U T, Dhawandkar S D and Seeras N R 1990 Field efficacy of nuclear polyhedrosis virus against *Heliothis* (Lepidoptera:Noctuidae) on gram (*Cicer arietinum*) in Maharashtra. Indian Journal of Agricultural Sciences 60:287-289.
- Pfrimmer T R 1964 Populations of certain insects and spiders on cotton plants following insecticide application. Journal of Economic Entomology 57: 640-644.

- Pimbert M P 1990 Some future research directions for integrated pest management in chickpea. In, Chikpea in the ninetees : Proceedings of the Second International Workshop on Chickpea Improvement, ICRISAT Center, Patancheru, Andhra Pradesh, India, 4-8 Dec1989. pp.151-163.
- Prabal Saikia and Rameswaran S 2000 Repellant and antifeedant effect of EC and dust formulations of plant derivatives against rice folder, *Cnephalocrosis medinalis* Guenee. Pestology XXIV:32-34.
- Pradan S, Jotwani H G and Rai B K 1962 Use of neem as natural pesticide. Indian forming 12:7-11.
- Prasad C S and Singh V P 1997 Impact of variety, sowing date and control measures on incidence of pod borer, *Helicoverpa armigera* (Hub.) and yield of chickpea. Annals of Plant Protection Sciences 5:26-28.
- Prasad D and Chand P 1986 Campoletis chlorideae Uchida a new parasite of Helicoverpa armigera (Hubner) in Ranchi, Bihar. Indian Journal of Entomology 48:231-232.
- Prasad J V and Ramakrishnan N 1993 Late larval resistance of Spodoptera litura (Fab.) to nuclear polyhedrosis virus. In, Proceedings of Indian National Science Academy, 59:543-548.
- Rabindra R J and Jayaraj S 1988 Efficacy of nuclear polyhedrosis virus with adjuvants as high volume and ultra low volume applications against *Heliothis armigera* Hubner in chickpea. Tropical Pest Management 3:441-444.
- Rabindra R J, Sathiah N and Jayaraj S 1992 Efficacy of nuclear polyhedrosis virus against *Helicoverpa armigera* (Hubner) on *Helicoverpa* resistant and susceptible varieties of chickpea. Crop Protection 11:320-322.
- Rabindra R J, Sathiah N, Muthiah C and Jayaraj S 1989 Controlled droplet application of Nuclear polyhedrosis virus with adjuvants and

UV protectants for the control of *Heliothis armigera* Hubn. on chickpea. Journal of Biological control 2:102-105.

- Raguraman S and Singh R P 1999 Biological effect of neem (Azadirachta indica) seed oil on an egg parasitoid, Trichogramma chilonis. Journal of Economic Entomology 92:1274-1280.
- Rajasekaran and Kumaraswamy Y 1985 Antifeedant properties of certain plant products against Spodoptera litura Fab. In, Proceedings of National Seminar on Behavioural Physiology Appr. Mrmt. Crop Pests. TNAU:25-28.
- Ramachandra Rao G G, Ragavaiah G and Nagalingam B 1990 Effect of botanicals on certain behavioural responses and on growth inhibition of tobacco caterpillar *Spodoptera litura*. In, Proceedings of National symposium on problem and prospects of botanical pesticides in integrated pest management, CTRI, Rajahmundry, A.P., India. pp. 12.
- Rao N V, Reddy A S and Reddy P S 1990 Relative efficacy of some new insecticides on insect pests of cotton. Indian Journal of Plant Protection 18:53-58.
- Ravi D and Verma S 1997 Persistence and dissipation of insecticides against *Heliothis armigera* on chickpea. Indian Journal of Entomology 59:62-68.
- Ravi G and Verma S 1997 Evaluation of pesticides against Heliothis armigera and its parasitoid Campoletis chlorideae on chickpea. Indian Journal of Entomology 59:69-77.
- \*Rayner O Coates M and Newby R 1996 Consequences of pesticide use in spider communities to mango orchards. In proceedings of the XIII International Congress of Arachnology Geneva, 3 – 8 September 1995. Revue Suirs de Zoologie (1996) Horrserie. pp. 537-547.
- Reddy G V P and Manjunatha M 2000 Laboratory and field studies on the integrated pest management of *Helicoverpa armigera* (Hubner) in
cotton, based on pheromone trap catch threshold level. Journal of Applied Entomology 124:213-221.

- \*Redfern R E, Warthen J D Jr, Uebel E C and Mills G D Jr 1981 The antifeedant and growth disrupting effects of azadirachtin on *Spodoptera frugiperda* and *Oncopeltus faciatus*. In, Proceedings of 1<sup>st</sup> International Neem Conference, Rettach-Egern 1980. Eschborn : GTZ. pp.297.
- Reed W and Pawar C S 1982 *Heliothis* : a global problem. In, Proceedings of the International Workshop on *Heliothis* Management. International Crops Research Institute for the Semi-Arid Tropics, Patancheru, A.P., India. pp. 9-14.
- \*Rembold H 1984 Secondary plant products in insect control with special reference to the azadirachtins. pp. 481, In advances in Invertebrates Reproduction, 3<sup>rd</sup> Edn., (ed Engles W) Elsevier, Amsterdam.
- Rembold H and Sieber K P 1981 Effect of azadirachtin on oocyte development in *Locusta migratoria migratorioides*. In, Proceedings of 1<sup>st</sup> International Neem Conference. Rottach-Egern, 1980. pp.75-80.
- Rembold H, Sharma G K and Czoppelt C H 1981 Growth regulating activity of azadirachtin in two hemimetabolus insects. pp. 121-128, In Natural Pesticides from the neem tree (ed Schmutterer H, Ascher K R S and Rembold H) GTZ, Eschborn, Germany.
- Rice M, Sexton S and Esmail A M 1985 Antifeedant phytochemical blocks oviposition by sheep blowfly. Journal of Australian Entomological Society 24:16.
- Rizvi S M A, Chaudhary M B, Pandey V and Upadhyay V K 1986 Efficacy and Economics of some insecticides in management of *Heliothis armigera* Hubner. Indian Journal of Plant Protection 14:47-50.
- Rosaiah 1992 Bioecology and management of *Helicoverpa armigera* (Hubner) on cotton. Ph.D. thesis, submitted to Acharya N G Ranga Agricultural University, Hyderabad.

- Ruberson J R, Young S Y and Kring T J 1991 Suitability of prey infected by nuclear polyhedrosis virus for development, survival and reproduction of the predator Nabis roseipennis (Heteroptera Nabidae). Environemental Entomology 20:1475-1479.
- Sachan J N 1990 Present status of Helicoverpa on pulses and strategies for its management. In, Sachan J N (Eds) First National workshop on Heliothis management:Current status and future strategies. Directorate of Pulses Research, Kanpur, India, 30-31 August, 1990 pp.8-33.
- Sachan J N and Lal S S 1993 Role of botanical insecticides in Helicoverpa armigera management in pulses. In, Botanical pesticides in Integrated Pest Management, ISTS, Rajahmundry, India. pp.261-269.
- Sajap A S, Kostulai J R, Kadir H A and Hussein M Y 1999 Impact of Prey infected by nuclear polyhedrosis virus on a predator, Sycanus leucomesus Walk. (Hemiptera : Reduviidae). Journal of Applied Entomology 123: 93-97.
- Samu F and Vollrath F 1992 Spider orb web as bioassay for pesticide side effects. Entomologia Experimentalis et. Applicata 62: 117-124.
- Sanap M M and Deshmukh R B 1987 Testing of different insecticides for the control of *Helicoverpa armigera* (Hubner) on chickpea. International Chickpea Newsletter 17:15-16.
- Sanap M M and Pawar V M 1998 Integrated management of *Helicoverpa* armigera on gram (*Cicer arietinum*). Indian Journal of Agricultural Sciences 68:162-164.
- Santharam G and Kumaraswami T 1985 Effect of some insecticides on emergence of the parasitoid, *Trichogramma chilonis* Ishii (Hymenoptera : Trichogrammatidae) Entomon 10: 47-48.

- Sarode S V and Sarnaik D N 1996 Integrated pest management in pulses. In, Wanjari K B, Raut B T and Potdakhe S R (Eds.) Strategies for increasing pulses production in Maharastra, Mumbai, India, 7-8 March, 1996. pp.7.
- Sarode S V, Deotale R O and Patil P P 1995 Performance of Helicoverpa nuclear polyhedrosis virus (HNPV) combined with neem seed kernel extract (NSKE) against the pod borer on chickpea. International Chickpea and Pigeonpea Newsletter 2:35-37.
- Saucke H 1995 Biological and integrated control of diamond back moth *Plutella xylostella* Linnaeus (Lepidoptera : Yponomeutidae) and other major pests in brassica crops. In, SPC/GTZ Biological control Project, Final report 1992-1995, Canedobu, Papua New Guinea.
- Saxena H P 1980 Report of research work on pests of pulses 1979-80. All India Coordinated Project on Improvement of Pulses IARI, New Delhi, pp.65.
- Saxena K N and Rembold H 1983 Orientation and oviposition responses of Heliothis armigera to certain neem constituents. In, Schmutterer H and Askher K R S (Eds) Natural pesticides from neem and other tropical plants. Proceedings of the 2<sup>nd</sup> International Neem Conference, Rausichholtzhausen. Eischborn. GTZ pp. 199-210.
- Saxena R C 1987 Neem seed derivatives for the management of the rice insect pests, a review of recent studies. In, Schmutterer H and Ascher K R S (Eds) Natural Pesticides from the neem tree and other tropical plants. Proceedings of the 2<sup>nd</sup> International Neem Conference, Rausichholtzhausen. Eischborn. GTZ. pp.81-93.
- \*Saxena R C 1989 Insecticides from neem (ed Arnason J T, Philogene B J R and Moroand O) presented at ACS symp. Scr. 387, American Chemical Society, Washington, D. C., 110 – 135.
- Schmutterer H 1981 Ten years of neem research in the Federal Republic of Germany. In, Schmutterer H, Ascher K R S and Rambold H (Eds) Proceedings of 1<sup>st</sup> International Neem Conference, Rottach-Egern. Germany, 1980. GTZ Eschborn, Germany, pp.21-51.

- Schmutterer H 1990 Properties and potentials of natural pesticides from the Neem tree, Azadirachta indica. Annual review of Entomology 25:271-97.
- \*Schmutterer H 1992 EinfluB von Azadirachtin einer azadirachtinfrein Fraktion eines alkoholischen Niemsamenextrakte und von formulieten Extrakten auf verpuppung, Schlupf und Imagines der kohlweiBlings brackwespe Apanteles glomeratus (L.) (Hymen., Braconidae). Journal of Applied Entomology 113:79-87.
- \*Schmutterer H and Holst H 1987 Untersuchungen under die wirkung des angereicherten und formutierten. Niemsamenextrakts AZT-VR-K auf die Honigbiene Apis mellifera L. Journal of applied Entomology 103:208-213.
- \*Schoonhoven L M, Greenhalgh R and Roberts T R 1987 Pesticide Science and Biotechnology. Black well Scientific, London, UK.
- \*Schulz C, Kinzle J, Harrmann P and Zebtiz C P W 1997 (Neem Azal-T/ s-a new botanical insecticide for fruit growing) Neem Azal-T/s-Einnuues botanishces Insektizid fur den Obstban. Gesunde Pflanzen (1997) 49 :97- 99.
- Sehgal V K and Ujagir R 1990 Effect of synthetic pyrethroids, neem extracts and other insecticides for the control of pod damage by *Helicoverpa armigera* (Hubner) on chickpea and pod damage and yield relationship at Pantnagar in North India. Crop Protection 9:29-32.
- Senapathi H K, Sahoo B K, Pattnaik M R and Pal A K 1992 Persistence of some commom pesticides in pigeonpea. Orissa Journal of Agricultural Research 5:100-103.
- \*Serra A C 1992 Untersuchungen zum Einsatz Von Niemsamenextrakten im Rahmen integrierter Ansatzezur Bekampfung vas Tomatenschadlingen in der Dominikanischen Repubik Doctor thesis, University of Giessen, Germany.

- Sharma M L, Rai H S and Verma M L 1997 Biopesticides for management of *Helicoverpa armigera* (Hubner) in chickpea. International Chickpea and Pigeonpea Newsletter 4:26-27.
- Shrivastava S K and Yadav K P 1991 Distribution of *Heliothis armigera* (Hub.) and its biocontrol agents in Chhattisgarh, Madhya Pradesh. Agricultural Science Digest 11:107-109.
- Singh G and Balan J S 1986 Host plant and natural enemies of Heliothis armigera (Hubner) in Haryana. Indian Journal of Ecology 13:175-178.
- Singh R P, Devakumar C and Dhingra S 1988 Activity of neem (Azadirachta indica A. Juss.) seed kernel extracts against the mustard aphid, Lipaphis erysimi. Phytoparasitica 16:225-229.
- Singh Y P, Gangwar S K and Azad Thakur N S 1990 Extent of endosulfan residues on pods of soyabean. Legume Research 13:9-12.
- Singh Y P, Srivastava A S and Singh S V 1988 Residues of phosphamidon, endosulfan and monocrotophos in/on sorghum grains Indian Journal of Entomology 50:17-23.
- Singla M L, Mahal M S and Balraj singh 1989 Assessment of loss in yield of gram by the pod borer, *Heliothis armigera* (Hub.). Journal of Insect Science 2:38-43.
- Sinha S N 1993 Control of Helicoverpa armigera Hubner infesting chickpea:Field efficacy of neem products and insects growth regulators. Indian Journal of Plant Protection 21:80-84.
- Sinha S N and Mehrotra K N 1988 Diflubenzuron and neem (Azadirachta indica) oil in control of Heliothis armigera infesting chickpea (Cicer arietinum). Indian Journal of Agricultural Sciences 58:238-239.
- Sithanantham S 1987 Insect pests of pigeonpea and chickpea and their management plant protection in field crops : lead papers of the

National Seminar on Plant Protection in Field Crops, CPPTI, Hyderabad, India, 29-31 January 1986, A.P., India. pp.159-173.

- Sithanantham S, Rao V R and Ghaffar M A 1984 International review of crop losses caused by insects in chickpea. In, Proceedings of the National seminar on crop losses due to insect pests. 7 – 9 January 1983, Hyderabad, India (Rao B H K and Murth K S E K (Eds.) Hyderabad, A. P., India : Entomological society of India. (Special issue, Indian Journal of Entomology, Vol – 11). pp. 263 – 283.
- Smits P H and Vlak J M 1988 Biological activity of Spodoptera exigua nuclear polyhedrosis virus against S. exigua larvae. Journal of Invertebrate Pathology 51:107-114.
- Srinivas G and Sundara Babu P C 2000 Effect of neem products on predatory green lace wing, *Chrysoperla carnea* Stephens (Chrysopidae : Neuroptera). Pesticide Research Journal 12:123-126.
- Srinivas P R 1989 Extent of parasitism of gram pod borer (*Helicoverpa armigera*) by ichneumonid larval parasites. Indian Journal of Agricultural Sciences 59:377-72.
- Srinivas P R and Jayaraj S 1989 Record of natural enemies of *Helicoverpa* armigera from Coimbatore district, Tamilnadu. Journal of Biological Control 3:71-72.
- Srivastava C P and Srivastava R P 1990 Antibiosis in chickpea (*Cicer* arietinum) to gram pod borer, *Heliothis armigera* (Hub.) (Noctuidae : Lepidoptera) in India. Entomon 15:89-94.
- Srivastava K P and Parmar B S 1985 Evaluation of neem oil emulsifiable concentrate against sorghum aphids. Neem Newsletter 2:7.
- Tahir Anwar, Takir S and Jabbar A 1993 Effect of neem oil on the longevity and fecundity of chickpea pod borer. Pakistan Journal of Agriculture Research 14:340-343.

- Tanwar R S and S K Honda 1998 Persistence, translocation and metabolism of endosulfan residue on Pigeonpea (*Cajanus cajan* L. Mill sp.). Pesticide Research Journal 10:73-79.
- Thakur R C 1990 Present status of Helicoverpa on pulses and strategies for its management in Madhya Pradesh. In, Sachan J N (Eds) First National Workshop on Heliothis Management:Current status and future strategies, Directorate of Pulses Research, Kanpur, India, 30-31 August, 1990. pp.91-99.
- Thakur R C, Nema K K and Kango K N 1988 Comparative efficacy of neem seed kernel and some insecticidal formulations against the gram pod borer, *Heliothis armigera* (Hubner). Legume Research 11:114-116.
- Tinśley T W 1979 The potential of insect pathogenic viruses as pesticidal agents. Annual Review of Entomology 24: 63-87.
- Udean A S, Joia S S and Chawla R P 1991 Endosulfan residues in Mustard seed at harvest. Journal of Insect Science 4:99-100.
- Ujagir R, Chaubey A K, Sehgal V K, Saini G C and Singh J P 1997 Evaluation of some insecticides against *Helicoverpa armigera* on chickpea at Badaun, Uttar Pradesh, India. International Chickpea and Pigeonpea Newsletter 4:22-24.
- Usha G Patel and Patel J R 1997 Ovicidal effect of botanicals alone and incombination with synthetic insecticides on eggs of *Helicoverpa armigera*. Indian Journal of Entomology 59: 326-329.
- Van Den Berg H, Hassan K and Marzuki M 1998 Evaluation of pesticide effects on Arthropod predator populations in soybean in farmer's fields. Biocontrol Science and Technology 8:125-137.
- Verma S 1983 Persistence of insecticides on Bengalgram. Journal of Research, Assam Agricultural University 4:136-140.

- \*Vogt H 1993 Einsatz Von Niempraparaten gegen Adoolophyes orana F V R und Untersuchungen Zu Ne Benwirkungen. In : 6, Intern. Exfahrungsaust Uber For Schungserg. Zum Okologischen Obstbau, LVWO Weinsberg. pp. 51 – 55.
- Vyas H G and Lakhchaura B D 1996 Effects of nuclear polyhedrosis virus of *Helicoverpa armigera* on pod damage and yield of chickpea at Pantnagar. Journal of Maharastra agricultural Universities 21:302-303.
- Warthen J S Jr 1979 Azadirachta indica : a source of insect feeding inhibitors and growth regulators. US Department of Agriculture, Agricultural Research Results, ARR-NE-5.
- Weigand S and Tahhan O 1990 Chickpea insect pests in the Mediterranean zones and new approaches to their management. In, Chickpea in the nineties : Proceedings of the Second International Workshop on Chickpea Improvement, ICRISAT Center, Patancheru, A.P., India, 4-6 Dec 1989. pp.169-175.
- Whitlock V H 1977 Effect of larval maturation on mortality induced by nuclear polyhedrosis and granulosis virus infection of *Heliothis armigera*. Journal of Invertebrate Pathology 30:80-86.
- Wightman J A, Anders M M, Rao V R and Reddy L M 1993 Cattle egrets may be important predators of *Helicoverpa armigera* on chickpea. International Chickpea Newsletter 29:19.
- Wightman J A, Anders M M, Rao V R and Reddy L M 1995 Management of *Helicoverpa armigera* (Lepidoptera : Noctuidae ) on Chickpea in Southern India : thresholds and the economics of host plant resistance and insecticide application. Crop Protection 14:37-46.
- Wilson L J, Bauer L R and Lally D A 1998 Effect of early season insecticide used in predators and out breaks of spider mites (Acarina : Tetranychidae) on cotton. Bulletin of Entomological Research 88: 477-488.

- Wu H L 1986 Determination of toxicity of some insecticides to Lycosa pseudoannulata and Apanteles cypris. Natural enemies of Insects 8:230-231.
- Yadava C P 1990 Need of ecological studies in developing effective Heliothis management in Uttar Pradesh. In, Sachan J N (Eds) Status and future strategies, Kanpur, U.P, India, 30-31 Aug 1990. pp.42-51.
- Yadava C P and Lal S S 1988 Relationship between certain abiotic and biotic factors and the occurence of gram pod borer, *Helicoverpa armigera* (Hubner) on chickpea. Entomon 13:269-273.
- Yadava C R 1996 Improved package of practice of chickpea production in Nepal. International Chickpea and Pigeonpea Newsletter 3:7-8.
- Yadava D N, Patel R C and Patel D S 1985 Impact of inundative releases of *Trichogramma chilonis* Ishii against *Heliothis armigera* (Hubner) in Gujarat (India). Journal of Entomological Research 9:153-159.

\*Originals not seen

Note:- The literature is cited as per the "Thesis Guide lines" Prescribed by Acharya N.G.Ranga Agricultural University, Rajendranagar, Hyderabad.

4	ppendix   :	Efficacy of	different IF	M compor	ients on o	viposition t	sehaviour c	of H.armige	ra during r	abi 1998-95	) (No. of eg	gs / plant)	
	15 DAG	22 DAS	29 DAS	36 DAS	43 DAS	50 DAS	57 DAS	64 DAS	71 DAS	78 DAS	85 DAS	92 DAS	99 DAS
Nem Nem	2 1625	0.35	0.325	0.7875	1.1375	17126.1	0 6625	1 4125	0 1675	1.55	0 0125	0	0
	11.4705	0.3669]	002500	[0.6872]	[1 0662]	[1:3083]	[0 5284]	[: :63]	[0.4529]	[1.2447]	[0.1116]	[0:2236]	leszz ol
NPV	23125	0.55	0.4125	0.8875	1.4375	2.05	-	3.6	0.3	1.8625	0.0375	01	0.0375
	[1.5207]	[0.5914]	[0.6421]	0.9416]	[1.1986]	[1.4314]	[ <b>Z</b> ]	(c; c; c; c)	[0.5475]	[1:3646]	[0.1934]	[Z286:0]	[0.4742]
Bird perches	2.3	0.575	0.475	0.85	1 85	25	1 0625	15125	0.325	1.625	0.0375	0.0875	0.0375
	[1 5165]	<b>[0.5913]</b>	(8585 Ci	[7126:0]	[1 3600]	[1.5811]	[9063 i]	[1622 :]	lo 5700]	[1 3607]	(0 1335)	[0 3708]	[1:27:0]
Endosulfan	2.1875	527-0	0 3625	0 9775	165	28	5,8125	15:25	0 2575	1 7375	00125	0 0375	0
	11,4788	0.4741	[0 5020]	[0 9681]	(1 2842)	(1 6730)	io 9012)	[c;2]	(0 5359)	[1 3188]	[7111 O]	(0 4743)	(szz)
Md	2175	0.3625	0.25	0 7375	1 425	1 9125	0 75	1 525	0 1575	15125	0 025	0 025	0
	[; 4742]	<b>[0.4028]</b>	[05 C!	0 6585]	[+ 1935]	[1 3625]	[9555 C]	(s :2 :)	(5257 Cİ	[12221]	[0 1579]	[0 2738]	[0 22:9]
Control	22125	0 675	0 625	1 3625	22125	3 3625	1 075	: 525	0 4575	2 0375	0 CE75	C 1375	990 0
	[1 4871]	10 6885	(cce) (	[1.167.1]	(197 L)	[2633 []	11 0366]	[535] J	1.859 Cl	[: 0163]	10 2097)	(0557 O)	(2318.0)
Sed	0099	0 072	1000	014	0 108	C 107	5 CG4	840 C	Z- 0	0 067	0.067	0 046	250.0
8	0 184	0 151	38 13	6Z ()	C 22.2	0 225	0:16	55	1220	0 121	11 de 1	5:5 0	83

	Appendi	x II: Efficac	y of differe	nt treatment	s against sı	mall sized I	arvae of <i>H</i> .	armigera dı	uring rabi	0N) 66-8661	. of larvae /	plant)	
Treatment	15 DAS	22 DAS	29 DAS	36 DAS	43 DAS	50 DAS	57 DAS	64 DAS	71 DAS	78 DAS	85 DAS	92 DAS	SYQ 66
Neem	0 275	205	1 0375	0 7825	0.8625	0.975	1 9875	1 3375	0.925	11125	18	185	0 2875
	[0.5244]	[1:4317]	[1.0165]	[0.6702]	[0.9283]	[0.9869]	[260; 1]	[1 1562]	[5096]0]	[1 0541]	[1 3412]	[1.2841]	lo 5359)
NPV	0:30	1.9375	0.75	0.8125	0.85	1.025	1.525	1.2875	0.9125	105	1.75	1,5125	0 3875
	[0.5477]	[1.3919]	(0 <del>308</del> .0)	(1006;0]	[0.9213]	[1.0119]	[13201]	[1.1341]	[0.9647]	[1:0242]	[1.3129]	[1 2292]	0 6221)
Bird perches	0.275	1.875	1.15	0 6375	1.125	1.4375	2525	145	<b>1</b> .	1,3125	1 625	1.65	036
	[0.5244]	[1.3693]	[1 0723]	[0-91-49]	[1 0601]	[1 1982]	[1 5200]	[1 2038]	[1 0241]	[1 1451]	[1 2742]	[1 2839]	[0 5911]
Endosulfan	0.3125	150	0 7525	977.0	0 6375	0 9125	15875	12125	0 9875	1 0375	1 825	1 525	0.35
	10 5530J	[1 2247]	[0 8732]	0 6501)	(07979)	(ආශා ර	[1823]]	(1 1008)	ize66 ol	(10167)	(ece 1)	1 2343	[0 59 <b>0</b> 8]
Wdi	925 0	2 025	90	0.7125	0.775	0 6875	17225	1 225	0 8875	1 0875	1 5125	1 3125	0 225
	[0 5701]	(1 4230)	[97 <i>1</i> 5 0]	[0 <del>1</del> 122]	[0 8800]	[2:25 0]	1923 1	[501 t]	[0 9418]	[15:01]	[2622 1]	[2571 ]]	[1272.0]
Control	03125	2 10	1 3675	83 11 1	1 275	1 475	2 55 2	1 4525	1 (625	다. 11	20:25	(7) ***	ŝ
	[0 5590]	[: 4491]	(11779)	[1785 i]	11 1257	(1712 L)	15551	[1 2087]	[‡ 0302]	(229. t)	h 473	[+ 578t]	[0 7741]
Sed	6100	0 023	0.05	3.68	0 023	900	850 B	£20 C	0014	ф С	6:03	2 053	016
9	0 14	0.21	0 126	810 19	9 C 26	8		2, 3	101 O	43 1.1	621	·	S S

	×	ppendix III	: Efficacy o	f different tr	eatments a	gainst med	lium sized l	larvae of H.	armigera	during <i>rabi</i>	1998-99 (No.	of larvae / pla	nt)
Treatment	15 DAS	22 DAS	29 DAS	36 DAS	43 DAS	50 DAS	57 DAS	64 DAS	71 DAS	78 DAS	85 DAS	92 DAS	99 DAS
Neem	0.0125	0 3000	0 9250	C 2750	0 4000	0.3750	1.0375	0.55.0	1.4375	0 4375	0 220	1 6125	0 3375
	[0.1117]	[0.5470]	[0 <b>3</b> 615]	lia 52 al	[0.6321]	[0 6121]	[i 0184] r .	, [0:5"38]	[1 1966]	lus oj	(cxc)	[1 0051]	[0 5603]
HNPV	0.0375	0.3625	0 9375	0.2575	0.3875	0.3750	1.0125	9237 0	1 3750	0 3375	0 7575	6975	0.1875
	[0.1 <b>3</b> 36]	<b>[0.6019]</b>	[1896.0]	(8905 D)	[0 6222]	0.6122]	[1:0060]	(ocss a)	[1.1724]	[C 5607]	[0 55/3]	(c.285.0)	[0.4329]
<b>Bird perches</b>	0.025	0.375	1.175	52170	0.5125	0.55	105	83	1.15	0.475	08125	095	0.2625
	<b>(0.1580)</b>	<b>[0.6121]</b>	[1 0838]	(02~S C)	<b>(0.7156)</b>	[0.7415]	[1 0244]	[ <b>;;;</b> ]	[1:0721]	0685 01	(11:06 Oİ	<b>[9</b> 744]	<b>[0.5122]</b>
Endosultan	<u>80</u> 0	0.5125	0 9125	C 4375	0.3875	0.45	0.9625	ទេ	1 4625	6.3625	C 2015	060	0 325
	[FCZZ:0]	[0.5292]	[0 9550]	[0 <del>3</del> 613]	[0 6223]	[0 <b>6706</b> ]	6066 0]	(oc)	[1 2091]	[ຄາວຣ ວ]	(67:5 C)	jo 3486]	(553 <b>5 0</b> )
M	. 0 052	<u>0</u> .6	0 9875	C 2375	0.375	0 3625	C 6575	0.4575	1 2125	C 1975	523 J	C 5125	0175
	0 2437)	0 5587]	[0 9934]	(1 5368)	[0 6120]	[0 6018]	[2175 0]	06:33	[1 1003]	(6Z\$7 2)	free ei	(Z108 0)	[0 41EC]
Control	0 65	0 7375	1 325	15 15	0 625	0 6875	1 3626	92.20	1 45	9259 C	8	: 2575	0 40
	lo 2235)	[0 5125]	[1 1506]	(cees c)	io 7504]	[0 8 <b>2</b> 30]	lars: 1	621 C	[1 2033]	Б С	ត្ ទំនួន	[950 J	jo 6575]
Sed	0 053	620.0	0.015	8 10 10	9 C 2 9	3 025	:00:	ф і і і і	0 630	01 10 10	<b>N</b>	85	810 O
8	0 174	0 051	Q 032	2400	0.061	C 052	C C22	Ř	0 127	100 100	2::3	1221	0 642

	Annendi	r IV • Effica	cv of differe	int treatmer	nts against l	large sized	larvae of H	, armigera	during <i>rabi</i>	1998-9 <del>9</del> (N	do. of larvat	e ( plant)	
Treatment	15 DAS	22 DAS	29 DAS	36 DAS	43 DAS	50 DAS	57 DAS	64 DAS	. SAC 17	78 DAS	SS DAS	92 DAS	99 DAS
		2	92.4 Q	100	0.175	0.0975	0 0875	0 175	0.075	0175	C 375	0 075	9 022
Neem	Þ	3					10 2657	10 41821	0 2736	[1812 C]	[2 6121]	[1573 J]	(0 1560)
		[0.2236]	0.4181	[1:512:1]	0.4102	funez ni	Pictorol			0 135	50	10	0.0125
NPV	0	0.025	0.2125	0.0675	0.15	0.125	0.0625	0.2	67611D	771.0	3		
		IQ 1581]	10.4607	0.2956]	[1.78C.0]	[0 3534]	(c.25)	[0.4472]	(0.25)	10 3534)	[0.5]	[0.3151]	0.1117
Died march ac	c	5200	0.2625	0.125	0.0875	0.1125	0.0875	0.2125	0.0625	0 1125	0 3875	0.1	<u> 3,0625</u>
	>	1757C (U	0.5122	0.3534]	10 2958)	[0.3352]	0.2956]	[0.4607]	(0 2498)	[C 3353]	lo 62231	[0,3162]	[0.25]
Endoculture	c	500	03	. 90	0.1	01125	<u>0.05</u>	0.125	0 075	61	1325	0 075	900
	•	ID 15791	10.4471)	10 ZZ3	0.3160	[0.3353]	leszz al	[0.3535]	(96.7.2 D)	ໂເຊເຊິ	(00 <i>1</i> 5 c)	[7.5.7.7]	[0 2235]
	c	0.875	0.2125		0.1125	0 125	0 0625	0065	5 2 2	C :375	920	01	0025
	5	n 1936	10.46081	10 2735	0 3353]	0 3534)	[cz:0]	[0 2548]	ic 2236]	(2025 cl	<b>(966</b> ≯ 0.	[0 3150]	(0 1580)
Control	c	0.1875	035	0 175	0 3625	0 2625	0.2575	033	C 125	80	C 6	9.20	Q; 0
	•	0 43301	[0:56:5]	1;3; <b>7</b> 0	[0 6C19]	(c 2.22)	ie a oi	[0:5700]	[CESE 0]	[0 53 16]	<b>isr</b> 22	[0 530]	10 3534)
Sed		0.016	620 0	30	0 052	0 (58	7:00	0048	C 108	an 1.1	0.024	900	6-30
6		0 160	0 092	.80	0 (53	0 121	0 03	0 101	0.227	<b>5</b> 80 I	0.061	83	150
3													

	~	Appendix V :	Efficacy of (	different trea	utments agai	inst all sized	larvae of H.	armigera du	ser labi gun	10 .0N 22-01	larvae / psain	-	
Treatment	15 DAS	22 DAS	29 DAS	36 DAS	43 DAS	50 DAS	57 DAS	64 DAS	71 DAS	TE DAS	85 DAS	92 DAS	32 C 25
Neem	C.2875	2.4	2.1375	1.1125	1.4375	1.4375	3.2	1.8525	2.5375	1.525	2.7	2.6875	0.7:25
	[2.5362]	[1.5492]	[1.462]	[1.0547]	[1.1959]	[1.1989]	.7888]	[1.3547]	[1.5929]	[- 3374]	[1.6431]	[1.6393]	[;; <del>;;</del> 8:0]
Adnh	0.3375	2.325	1.9	1.1875	1.3875	1.525	3.0375	1.8125	2.4125	:.375	2.6125	2.5	0.6375
	[0:5809]	[1.5247]	[1.3784]	[1.0897]	[1.1779]	[1.2349]	1.7428]	[1.3453]	[1.5532]	[:.2796]	[1.6163]	[1.5811]	[ <del>5</del> 537]0]
<b>Bird perches</b>	0.3	2.325	2.5875	1.375	1.725	2.1	3.8875	2.0525	2.3125	2.175	2.5375	2.6625	<u>1</u> 0
	[ <b>3.54</b> 77]	[1.5248]	[1.6085]	[1.1726]	[1.3134]	[1.4491]	[1:9716]	[1:4351]	[1.5207]	[1:,4747]	[1.5929]	[1.6317]	[0.8353]
Endosulfan	0 3625	1.8	1.875	1.2125	1.125	1.475	2.975	1.7875	2.55	.725	2.7375	2.475	0 725
	[].6021]	[1.3416]	[1.3693]	[1.1011]	[1.101.1]	[1.2144]	7248	[1.3359]	[1.5968]	["3134]	[1.6545]	[1.5732]	(7, <u>23</u> 0)
١٩٩	0.387	3.2125	2.15	1.075	1.2625	1.375	2.715	1.7525	2.2375	: 525	2.2375	2 15	0.4525
	[3:6221]	[1.7923]	[1.4662]	[1.0385]	[1 1236]	[1.1726]	[2279].	[1.3276]	[8567 ]]	6562 .	[1.4655]	1 4563	jc est i
Control	0.3625	2.8	3.0625	1.7625	2 2625	2.425	1375	22	2.8625	1 6325	3.2525	3.3125	1 25 5
	[0.6021]	[1.6735]	[1.75]	[1.3275]	[1534]]	[1.5572]	[3 0916]	[1,433]	[1:6918]	[/ ess/]	[1.8034]	[t 6201]	11 11 24
Sed	300	0 025	6100	C C55		30	3049	3	2014		5:00	••••••	 
9	20	120	SI 13	0 126	271.0	;;; · ,			: 	:"; ; ;	12	24 1 - 2	3. .:

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dix V - Efficacy of different treatments against all sized larvae of H. armigera during rabi 1998-99 (No. of larvae / plant)

								- Anrino Cabi	1999-2000 INo. 0	f eggs/plant}		
		-	iv VI-Fffi	acy of different (	treatments on o	riposition behav	iour of H.armige	Summe B	Rt DAS	SAC 33	90 DAS	5:25
		Appen	11.11.11	0.00	54 DAS	65 0AS	EB DAS	2	25-10	2010-0	0 0025	.,
	33 DAS	SHO ST	12 CFS	4/ UAS			570 C	8	0.012			1252
Treatment	0.0125	92310	0(375	0.625	0 :575	671 0	0.02 C	[FC72 0]	(560; OJ	[g] 1: 2 <sup>:</sup>	(5570 D	. 1615. . 1615
Neem	0.1117	(1991 C)	0 2557)	10 155C	lezez di	[0 3532] 2 2 2 2 2	(u 1625	;;	0.037	<b>99:</b> 0	0000	1526 -
10000	0175	10	0.2125	0.2	0.425	c717 ()	10,4030)	[13161]	12261 O	[ <b>210:</b> -0]		. 0
AANH	10,4182	03150	10.4507	107±4.0	(0.6517)		0 2375	C.375	900	C 069		المحكومة
	0.25	0.2375	0.25	0.2575	0 2625	c1.0	11287 00	(9952 C)	lo 2234]	(r292 d)	0 1935	
Bird perces	10501	[Z187 0]	0.4933	11722-01	[0.5120]	invec.ol	0 13/5	10.1	C0125	3.62	00200	1925 C
-	6787.0	0 175	0.2375	0 6525	0 25	c792.0	0 170K	0.534	[21110]	(C 2486)	(0 i624)	
Endosultan	10,4326	0 4:8:1	l0:457	1 102-5-]	(1997) 10.4997)	105120	in 176	C. 375	0 0065	0.018	9 <b>200</b> 0	
ļ	10	0 0525	0:	0175	0 3625	015	1835 U		(008C 0	[600; C	[S0 0]	
15 M	:03161	157 Q)	<b>JO 3:9</b>	23:7 c) lo	j0 6018J	10 3305 D	04	St	G115	396	770.0	
1040	0 275	0 2625	0.32	5 0 2525	517 0		, 10.5322	0037 1	[2 1182]	0995	100-101 0.011	5N:
Control	10 5245	9 (D5:22)	35 Ol	17:30 IS:	10699 01	17575 ()	690	8	950 0	516	1970	3
Ţ	. 0 0	30055	00	7500 T			9 <b>1</b> 10	57	0.205	5	2011	
8	500	· · · · ·	3	3 0135	315							

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Treatment	24 DAS	33 DAS	40 DAS	45 DAS	47 DAS	54 DAS	66 DAS	68 DAS	75 DAS	81 DAS	83 DAS	90 DAS	97 DAS	98 DAS	105 DAS	112 DAS
Neem	0.45	0.5375	0.586	0.7125	0.65	0.9625	1.413	1.325	0.625	0.6125	0.625	0.4375	0.7875	0.725	0.375	0.2625
	[0.6707]	[1557.0]	[0.7415]	[0.8439]	0.8062]	[6086.0]	[1.1883]	[1.1509]	[0.7904]	<b>[0.7825]</b>	0.9905]	[0.6612]	<b>[0.8871]</b>	[0.8513]	0.6121)	[0.5121]
Adnh	0.48	0.5125	0.4375	0.6125	0.63	0.900	1.375	1.35	0.5625	0.63	0.6	0.39	0.7	0.775	0.3375	0.25
	[0.6927]	[0.7157]	[0.6613]	0.7825]	[0.7904]	[0.9485]	[1.1725]	[1.1617]	[0.75]	[0.7904]	[0.7744]	[0.62247]	[0.8364]	[0.8801]	[0.5806]	0.50
<b>Bird perches</b>	0.55	0.7625	1.1375	0.81	0.8000	1.713	1.3750	1.2875	0.9875	0.7625	0.8125	0.9875	0.7875	0.8875	0.8125	0.6875
	[0.7415]	[0.8731]	[1.0665]	[0.9012]	[0.8943]	[1.3085]	[1.1726]	[1.1344]	(9666 <sup>.</sup> 0)	[0.8731]	[0.9011]	[0.9935]	[0.8872]	[0.9417]	[0.9012]	0.8289
Endosultan	0.5	0.2875	0.3875	0.725	0.238	1.125	1.4	0.6375	0.9125	0.675	0.475	0.6375	8375	0.425	0.4625	0.3625
	[0.7071]	[0.5360]	<b>[0.6223]</b>	[0.8513]	[0.4872]	[1.0604]	[1.1831]	[0.7983]	[0.9551]	[0.8214]	[0689:0]	[0.7982]	[[0:9150]	[0.6518]	[0.6800]	[0.6017]
. Wdi	0.525	0.4875	0.49	0.70	9.0	0.925	1.325	0.525	0.85	0.6375	0.6125	0.4625	0.6625	0.625	0.25	0.2
	[0.7243]	[0.6981]	[0.6981]	0.8365]	[0.7744]	[0.9616]	[1.1509]	0.7244	[0.9217]	[0.7982]	<b>[0.7823]</b>	<b>į0 6800</b> ]	[0.8138]	(2057.0)	[0 <del>2</del> 0]	0 4470]
Contrei	0.475	0.788	1.175	0.825	0.7750	1.7625	1.463	1,3750	1.0375	0.7675	0 850	1.075	0.8875	150	6.0	0.8875
	[0.6891]	[0.8873]	[1.0837]	[0.9081]	[0.8802]	[1.3273]	[1:2081]	[1.1725]	[1.0184]	[0.8874]	[0.9217]	[1 0368]	0 9419]	[0:36:0]	[0.9483]	0.9419]
Sed	0.048	0.042	C.038	0.048	0.033	0.019	0 015	0 023	0 014	0 033	0.048	0 033	0.023	0.028	0 023	0.042
ទ	0.101	0.091	0.082	0.101	1/0.0	0.041	0 042	0.053	0.032	1200	0.101	0 072	0 051	0.063	0 052	60.0

Appendix. VII : Efficacy of different treatments against small size larvae of H. armigera during rabi 1999-2000 (No. of larvae / plant)

	Appen	dix.VIIt:Ei	flicacy of	different	treatmen	ts agains:	t medium	size larv.	ae of H. a	rmigera (	during ral	oi 1999-2(	000 (No. o	f larvae /	plant)	
Treatment	24 DAS	33 DAS	40 DAS	45 DAS	47 DAS	54 DAS	66 DAS	68 DAS	75 DAS	81 DAS	83 DAS	90 DAS	° DAS	98 DAS	105 DAS	112 DAS
Neem	0	0.4625	0.2	0.425	0.2875	0.213	0.375	0 475	0.5625	0.3875	0 9875	523-3	: 422	0 3375	0 2625	0 2125
	0.2236	00890	[0.4471]	<u> 0.6518]</u>	[0:5360]	[0.4607]	peizz	(Je88 OJ	(57.0)	lo.6223J	(0 9836)	[0:99: 3]	1 6:35	(0 580B)	(0 5122)	(0.4608)
NUN	•	0.4625	0.1375	0.3875	0.275	0.1625	0.2375	0.6125	0.325	. 0.36	0.9375	97 13	3675	0 2625	0.2215	0.175
	0.2236)	0.67983	[0.4471]	0.6222	0.5243	[0:4030]	10.4372)	[C297.0]	[0:5700]	0.6123	[0.9681]	[81920]	1.6223	[0.5122]	[0.4742]	[0.4182]
ird perche	•	0.5375	0.3875	0.725	0.4875	0.4375	05125	0.9375	0.8875	0.775	1.3125	05126	1 0625	0.825	0.5715	0.4875
	0.2236)	<b>(0557.0)</b>	0.6224	[0.8513]	[0.6981]	[0.6614]	[0,7157]	( <u>2396 O</u>	0.9419]	[2088:0]	[1.1455]	[c:c:c]	1000 L	[0.9081]	10957.01	[0.6981]
Endosulfa	0	0.275	0.2875	0.5875	0.1125	0.225	6:25	0 3625	0.4875	0.9875	0 825	(23)	: :629	05125	0 525	0.375
	0 2236	0.5243]	0.5360]	[0.7661]	[0.3353]	[0 47 42]	[7133 O]	0 6015]	0.6980)	[0.9935]	[0 9081]	[ <u>2951</u> ]	1 1253	(0512 O)	[0 7244]	<b>jo</b> 6120J
Mdi	0	0.4	0.19	0 44	0 2625	0.13	02	0 7625	0 4875	C 7125	06125	9197 C	3255	0.2375	0213	0.125
	0.2236	0.6327]	0.4328)	0.6631)	[0 5121]	jo 3538)	loz= ol	1578 G	(6269°C)	[0 8440]	[0.7824]	ខ្មែរទ្ធ ជ	6.20	[0 1570]	[0 4607]	(0 3233)
Control	0	06125	0.4625	0 625	0 5825	0 725	C STC	1 275	1113	1 025	1 4875	97.7	0 	0.575	0 675	0 5375
	lo 2236)	[0 7825]	0.6800]	[0000]	(c 7831)	[0 8513]	(c. cs o)	1125	[1 0546]	(cz:o :i	[12155]	[cay: .]	1788) 17880	(5353)	0 62:4]	[0 7330]
Sed	0	<del>5</del> 10 0	0.023	0 019	0 058	0 023	80	3-00	0.023	001	0 019	÷:	\$ZC :	6:00	0.014	0 019
8	000	20	90.0	0 040	012	0 020	12:0	5 12 12	50 20	800	200	2:	:* <u>:</u>	50	ខ្ល	100

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Treatment	24 DAS	SYU EE	40 DAS	45 DAS	SFI Lt	S4 DAS	66 DAS	68 DAS	75 DAS	SYQ 18	SYD 23	90 DAS	97 DAS	9% DAS	105 D.AS	112 DAS
Neem	0	0	0.1125	0.0625	0.075	0	0	0.25	0.525	0.1	0.2	0.46	0.075	0.375	0.025	0.05
	[0.2236]	[0.2236]	[0.3353]	[0.2498]	[0.2737]	[0.1580]	[0.2236]	[0.5]	[0.7243]	[0.3161]	[0.4028]	[0.6780]	[0.2737]	[0.1935]	[0.2737]	[0.3161]
HNPV	0	0	0.075	0.025	0.05	0.0125	0	0.2625	0.4375	0.088	0.175	0.3800	0.0375	0.0375	0.0375	0.05
	[0.2236]	[0.2236]	[0.2737]	[0.1580]	[0.2234]	[0.1117]	[02236]	[0.5122]	[0.6612]	[1282.0]	[0.4181]	[0.6202]	[0.1933]	[0.1934]	[0.2957]	[0.3161]
Bird perches	0	0.0375	0.0375	0.0375	0.0625	0.05	0.1125	0.2125	0.5125	0.1	0.2	0.39	0.0375	0.0375	0.075	0.0625
	[0.2236]	[0.1935]	[0.1936]	[0.1935]	[0.2497]	[0.2236]	[0:4030]	[0.4607]	[0.7156]	[0.3162]	[0.4470]	[0.6301]	[0.1935]	[0.1936]	[0.3534]	[0.3353]
Endosulfan	Ö	0	0.0625	0.125	0.0125	0.0375	0.1375	0.15	0.5375	0.1625	0.0875	0.41	0.1375	0.025	0.025	0.125
	[0.2236]	[0.2236]	[0.25]	[2E3E.0]	[0.3533]	[0.1934]	[0.4329]	[0.3870]	[0.7329]	[0.4029]	[0.2956]	[0.6401]	[0.3706]	[0.1580]	[0.2736]	[0.4182]
. Wal	¢	o	0.13	0.05	0.1	0.05	0.025	0.1375	0.45	0.1375	0.175	0.3975	0.0625	0.025	•	0
	[0.2236]	[0.2236]	[0.4162]	[0.3352]	[0:4030]	[0.4608]	[0.2737]	[0.6323]	[0.6706]	[0.4329]	[0.5242]	[0.6302]	[0.4329]	[0.2957]	[0.2236]	[0.2236]
Control	0	0	0.175	0.1125	0.1625	0.2125	0.24	0.4	0.55	0.1875	0.275	0.575	0.1875	0.1	0.1	0.1625
	[0.22 <b>36</b> ]	[0.2236]	[0.41£2]	0.3350]	0.4029]	[0.4607]	[0:5360]	[0.6322]	[0.7413]	[0.4327]	[0.5241]	[0.7580]	[0.4328]	[0.2956]	[0.3871]	[0.4608]
Sed	0	0.004	0.019	0.014	0.05	0.014	0.025	0.057	600.0	0.019	0.057	0.53	0.028	0.043	0.014	0.019
c	0	0.01	<del>2</del> 0.0	0.03	0.04	0.03	0.06	0.121	0.02	0.04	0.101	0.121	0.06	60.0	0.03	0.04

Appendix.IX:Efficacy of different treatments against large size larvae of *H. armigera* during *rabi* 1999-2000 (No. of larvae / plant)

Appendix	k.X:Eff	icacy of	f differ	ent tre:	atment	s again	st all si	ze larv	ae of H	armiş	<i>iera</i> du	ring <i>ra</i>	666T 10	M07-		
Treatment	24 DAS	31 DAS	40 DAS	45 DAS	A DAS	54 DAS	66 DAS	68 DAS	75 DAS	81 DAS	SAD 53	90 DAS	97 DAS	SE DAS	105 DAS	112 DAS
Neem	0 1 1 1 1 1	0.9625 10.9287	1.2 [1.0862]	1.0125	1.2 [1.095]	1.7875 1.7805 [1.3369]	, 2.05 [1.4317]	17125 1.3064)	1.1 [1.04668]	1.775 [1.3223]	1.4 [1.1659]	1.225 [1 1511]	1.1 [1.0488]	0.6625 [0 8139]	0.525 10.7245	0525 107245
VINH	0.975 10.9874)	0.65 [0.8062]	1.025 [1.0122]	0.95  0.9746]	1.075 [1.0368]	1.6125 [1.2696]	2225 [1.4916]	1.325 [1.1508]	1.0675 [1.0428]	1.713 [1.3066]	1.1975 [1.0941]	1.125 [1.0806]	1.075 [1.0368]	0.6 10.7745	0.475 10.6882]	0.475 [0.6590]
<b>Bird perches</b>	1.3375 [1.1565]	1.565 [1.25]	1.575 1.2548]	1.35 [1.1618]	22 [14832]	20 [1.4142]	2.4375 [1.5612]	2.3875 [1.5450]	1,6275 [1,2796]	2325 [1.5247]	2.1975 [1.4820]	1.8375 [1.3736]	1.75 [1.3228]	1.4625	1,2305 [1,1124]	12375 2021
Endosultan	5295.0 5295.0	0.7375 [0.8688]	1.4375 [1.1987]	0.3855 10.6021]	1 3875 [1 1779]	1.9025 [1.4009]	1.15 [1.0724]	1.3375 [1.3917]	1.825 [1.3609]	1.3875 [1.7779]	1.685 [1.2978]	1 8375  1 255 <b>5</b> ]	5296 0 [1196:0]	1.0125 [1.0062]	0.9625 10.92387]	0.625 10 9235]
PM	278870 (12%20)	0.8 [0.6944]	1 1875 [1.0695]	2360 11960	1.1 [1 0486]	1.2429] 1.2429]	1.1937] [1.1937]	1.7875 [1.3366]	1 4675 [1 2196]	1 4 [1.1832]	1.2575 [1 1367]	1 (575 1 (575	0.8875 (0.9421)	0 4625 [0 6801]	0 325 [0 5701]	0.375 10 57 30]
Control	1.4 [1.1832]	1.8125 [1.3463]	1 7375 [1.3180]	1 58 [1.2566]	27 [: 6431]	25125 [1.5651]	305 [17464]	2.7 [1 6429]	20 [14142]	2.6125 [1 6133]	27 [; 206]	22:22 [: 4374]	1 875 [1 3693]	1 675 [1 2942]	1 5875  1 2599	1523 1253
<u> 8</u> 8	0019	0.048 0.101	0053	0 014 0 032	C033 C071	0.025 0.063	0023 0052	0.042 0.091	003 0671	2100 0042	0.036 0.052	CC23 0552	006 0127	0015 0042	900 0 061	0015 0642

during rahi 1999-2000 f H. armige •